



Determination of Pesticide Residue Present in Cumin Plant (*Nigella orientalis* L.) with LC-MS/MS and GC-MS

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In this study, pesticide residues were investigated in black cumin (*Nigella orientalis* L.) seeds which grown in Turkey. GC-MS and LC-MS/MS analytical instruments are used in high precision, when determining residue limits. A total of 100 pesticide active ingredients in LC-MS/MS devices have been performed in *Nigella orientalis* L. seeds samples. In this plant, 103 pesticide active ingredients were analyzed in GC-MS. This study conducted in 2012 and 2013. Samples residues were not found in detectable levels for 2 years.

Keywords: Pesticide, Residue, Black cumin.

INTRODUCTION

Kinds of *Nigella* L. are originated from Mediterranean Region and Westerns Asia; they are members of *Ranunculaceae* family and they form 14 species [1]. They contain several important medicinal and aromatic species. It has been found that *N. sativa* and *N. damascena* L. Seeds have pharmacological effect [2,3].

Nigella orientalis is an annual plant and it can grow up to 30 cm; its flowers are yellow and its seeds are shaped as discs. They spread from North-East Mediterranean region, Caucasia and Crimeato Syria. Cumin has been used as a natural medicine in Middle-East and Far-East countries, over 2000 years now. They are grown in Southern Europe in a limited amount [4].

There are 15 species and 19 taxons in the flora of Turkey [1,5]. The plant is grown especially Afyon, Burdur, Isparta, Kütahya and Konya regions [6].

Its seeds are used as a flavour and the plant, itself, is grown as a foliage plant. Kokoska *et al.* [7] defined 18 compounds in *N. orientalis* L. seeds through their analyze of essential oils. They determined β -elemene (68.8 %), δ -cadinene (5.2 %) β -selinene (4.3) and α -selinene (3.3 %) in the highest rates. Noting that their study is the first on the field so far, as far as they have known, they claimed that, *N. orientalis* and *N. domascena* are more similar than *N. sativa*, in terms of essential oils.

Cumin seeds have been used as spice and nutrient protectors. The seed sand oil of the plant have potential medical

features, which can be used in traditional medicine. Cumins therapeutich (healing) feature and its role in cure of diseases and effect on human body was examined in this compilation. Cumin has the effect of starting antioxidant system, which cleans oxidants. It shows anti-inflammatory property and also supports the defense system (immune) [8].

Wagner and Fransworth [9], suggested that there is 6.4 % water, 4 % ash, 32 % oil, 20.2 % crude protein, 6.6 % crude fiber and 37.4 % carbohydrate in cumin seeds. As an addition, cumin seeds contain B1, B2, B6 vitamins in small amounts and iron, calcium, magnesium, zinc and selenium, as they suggested [10]. Farag *et al.* [11] defined 62 metabolites in 6 *Nigella* species, also including *Nigella orientalis* and revealed 52 of these metabolites. They claimed that their study was the first to form a comparing metabolites profile among *Nigella* species.

Baser *et al.* [12] claimed that linoleic and oleic acids are observed as the main fatty acids, in their study, conducted with GC-MS and LC-MS/MS, to examine the chemical components of *Nigella* seeds.

Meziti *et al.* [13] suggested that cumin plants are traditionally used in the cure of asthma, coughing, bronchitis, head ache, rheumatism, high-fever, flu and eczema. The same researchers claimed that *Nigella* seeds are reported to have some biological activities, including antioxidant activity. They claimed that the chloroform extract and ethyl acetate are the highest rated phenolic components in *Nigella* seed sand ordinarily they contain 81.31 and 72.43 μg , by using Folin-Ciocalteu method.

Wajs and Kalembe [14] defined 30 components in *Nigella orientalis* L. seeds in Poland and claimed that 16 of these components are new in these species. Researchers suggested that the most known *Nigella* species are *N. sativa* (black cumin) and *N. damascena* L. (love in the mist) and naturally they spread through West-Europe, North-Africa and South west Asia. As an edition, they stated that researches on *N. orientalis* are rather limited.

Sirma and Kadioglu [15] studied weed species on wheat cultivation areas through their study in 2006. They defined 51 weed species belonging 20 families in total. They included *Nigella arvensis* into these species as well.

Tutukcu and Sagdic [16] defined linen, nigella, thyme, nettle and camomille a highly requested plants, their study to examine medical and their drugs, sold in herbal shops through the years 2008 to 2009 in Konya.

Kim et al. [17] in their study to investigate nematode activity in plant oils observed that 17.2 % nematode activity took place in the oil from India-originated *N. sativa*.

Pesticides are used as protectors in agriculture and their over/misuse lead toxic effects in nature. Their misuse cause acute and permanent health problems on people. Although restrictions on pesticide use are imposed, pesticides remain as

residues on food chain as a result of reaping and marketing the products prematurely. Active substances of pesticides are known to cause acute toxic and geno toxic effects on people [1].

Highly sensitive analytical devices such as GC-MS and LC-MS/MS are used to define World Health Organization (WHO) and Food Agriculture Organization (FAO) international pesticide residue limits [18]. In this study, pesticide residues on Turkey originated Cumin (*Nigella orientalis* L.) seeds were investigated.

EXPERIMENTAL

In the study, the weeds obtained from *Nigella orientalis* L. plants grown by a farmer in Tefenni town of Burdur city in years of 2012-2013 are used as material. Cumin production is very prevalent around the region. Examples were gathered approximately 1 month later hence the harvest. As the purpose, 1 kg seed examples from 10 different sacks in the farmer's cumin stock were gathered and mixed together. 15 g of weed examples are studied for extractions 3 times repeatedly.

All the solvents and chemicals (water, acetonitrile, methanol, formic acid, acetic acid, ammonium formate) used as mobile phases in example extractions are chosen in accordance to a profound quality. Pesticides given in Tables 1 and 2 are searched in the examples of cumin seeds, which are the

TABLE-1
ACTIVE SUBSTANCES EXAMINED IN CUMIN SEEDS EXAMPLES ON GC-MS/MS DEVICE

| Analyte | LOQ (µg/kg) | Analyte | LOQ (µg/kg) | Analyte | LOQ (µg/kg) |
|-------------------------|-------------|---------------------------|-------------|--------------------|-------------|
| 1-3 Hexachlorobutadiene | 10 | Ditalimphos | 10 | Oxyfluorfen | 10 |
| Total DDT | 10 | α,β-Endosulfan sulfate | 50 | Pebutale | 10 |
| Acetochlor | 10 | Endrin | 10 | Penconazole | 10 |
| Alachlor | 10 | Endrin aldehit | 20 | Pendimethalin | 10 |
| Aldrin+Dieldrin | 10 | Endrin keton | 20 | Pentachloroanilin | 10 |
| α-BHC | 10 | EPN | 20 | Pentanachlor | 10 |
| Azinphos methyl | 10 | Ethalfuralin | 10 | Permethrin | 10 |
| Azobenzene | 15 | Ethiofencarb | 10 | Phorate | 10 |
| β-BHC | 10 | Ethofumasate | 10 | Piperonyl butoxide | 10 |
| Bifenthrin | 10 | Ethoprophos | 10 | Procymidone | 10 |
| Bitertanol | 10 | Etoxazol | 10 | Propanil | 10 |
| Bromophos ethyl | 10 | Fenamiphos | 10 | Propargite | 10 |
| Bromophos methyl | 15 | Fenarimol | 10 | Prothiofos | 10 |
| Bromopropylate | 10 | Fenclorfos | 10 | Pyrimethanil | 10 |
| Bupirimate | 10 | Fenitrothion | 10 | Pyrimidifen | 10 |
| Buprofezin | 10 | Fenpropathrin | 10 | Quinalphos | 10 |
| Cadusafos | 10 | Fenson | 10 | Quinomethionate | 10 |
| Captan | 50 | Fenthion | 10 | Quinoxifen | 10 |
| Chlorfenapyr | 25 | Fenvalerate+Esfenvalerate | 10 | Quintozene | 10 |
| Chlorfenson | 10 | Flamprop methyl | 10 | Rabenzazole | 10 |
| Chloroneb | 10 | Flucythrinate | 10 | Resmethrin | 10 |
| Chlorpropam | 10 | Fluotrimazole | 10 | Simazine | 10 |
| Chlorthal dimethyl | 10 | Flusilazole | 10 | Sulprofos | 10 |
| Chlorthalonil | 20 | Folpet | 10 | Taufluvalinate | 10 |
| cis + trans-Chlordane | 10 | Fuberidazole | 10 | Tebuconazole | 10 |
| Cyfluthrin | 10 | Heptachlor+izomerleri | 10 | Tebufenpyrad | 10 |
| Cypermehtrin+izomerleri | 10 | Hexachlorobenzene (HCB) | 10 | Tecnazene | 10 |
| Cyromazine | 10 | Hexaconazole | 10 | Tefluthrine | 10 |
| δ-BHC | 10 | Iodofenfos | 15 | Terbacil | 10 |
| Deltamehtrin | 15 | Isazofos | 10 | Tetraconazole | 10 |
| Demeton-S-methyl | 10 | Isodrin | 10 | Tetradifon | 10 |
| Desmethrine | 10 | Isopropalin | 10 | Tetrasul | 10 |
| Dialifos | 10 | λ-Cyhalothrin | 10 | Thiobencarb | 10 |
| Dichlofluanid | 15 | Lindane (γ-BHC) | 10 | Thiometon | 10 |
| Dicofol | 10 | Linuron | 10 | Tolclofos Methyl | 10 |
| Dinobuton | 15 | Methoxychlor | 10 | Trichlorfon | 10 |
| Disulfoton | 10 | Nuarimol | 10 | Trifluralin | 10 |
| Disulfoton sulfone | 10 | Ofurace | 10 | Vinclozolin | 10 |
| Disulfoton sulfoxide | 10 | Oxidixyl | 10 | - | - |

TABLE-2
ACTIVE SUBSTANCES EXAMINED IN CUMIN SEED EXAMPLES ON LC-MS/MS DEVICE

| Analyte | LOQ (µg/kg) | Analyte | LOQ (µg/kg) | Analyte | LOQ (µg/kg) |
|-----------------------------------|-------------|----------------------------|-------------|-------------------------|-------------|
| 3-Hydroxy carbofuran + Carbofuran | 10 | Epoxiconazole | 10 | Parathion methyl | 15 |
| Acephate | 50 | Ethion | 10 | Pencycuron | 10 |
| Acetamiprid | 10 | Ethirimol | 10 | Phentoate | 10 |
| Aclonifen | 10 | Ethoxyquin | 30 | Phosalone | 10 |
| Aldicarb+sulfone+sulfoxide | 50 | Etrifos | 10 | Phosmet | 10 |
| Allaxydim Na | 10 | Famoxadone | 10 | Phosphamidon | 10 |
| Amitraz+Metabolitleri | 30 | Fenamidone | 20 | Phoxim | 10 |
| Anilofos | 10 | Fenazaquin | 10 | Picloram | 10 |
| Atrazine | 10 | Fenhexamid | 10 | Pirimicarb | 10 |
| Atrazine Desethyl | 10 | Fenoxaprop P | 10 | Pirimiphos ethyl | 10 |
| Azadirachtin | 20 | Fenoxycarb | 10 | Pirimiphos methyl | 10 |
| Azoxystrobin | 10 | Fenpiclonil | 10 | Pirimisulfuron methyl | 10 |
| Azpyrotryn | 10 | Fensulfothion | 10 | Prochloraz | 10 |
| Benalaxyl | 10 | Fenuron | 10 | Profenefos | 10 |
| Bendiocarb | 10 | Fluquinconazole | 10 | Profoxydim | 10 |
| Benfurocarb | 10 | Flurochloridone | 10 | Promecarb | 10 |
| Benomyl+Carbendazim | 10 | Flutriafol | 10 | Prometryn | 10 |
| Bensulfuron methyl | 20 | Fonofos | 25 | Propachlor | 10 |
| Boscalid | 10 | Formothion | 20 | Propamocarb | 20 |
| Bromacil | 10 | Furathiocarb | 10 | Propaquizafop | 10 |
| Bromuconazole | 10 | Haloxypop etoxyethyl | 10 | Propazine | 10 |
| Butocarboxim | 50 | Haloxypop methyl | 10 | Propiconazole | 10 |
| Butylate | 30 | Heptenephos | 10 | Propoxur | 10 |
| Carbaryl | 10 | Hexythiazox | 10 | Propyzamide | 20 |
| Carbosulfan | 10 | Imazalil | 10 | Proquinazid | 10 |
| Carboxin | 10 | Imazaquin | 20 | Prosulfuron | 10 |
| Carfentazone ethyl | 10 | Imidocloprid | 10 | Pymetrozine | 20 |
| Chlofentezine | 10 | Indoxacarb | 10 | Pyraclostrobin | 10 |
| Chlorbromuron | 10 | Iodosulfuron methyl sodium | 10 | Pyrazophos | 10 |
| Chlorfenvinphos | 10 | Iprodion | 20 | Pyrazosulfuron | 10 |
| Chloridazon | 10 | Iprovalicarb | 10 | Pyridaben | 10 |
| Chlormequat chloride | 50 | Isoprotun | 10 | Pyridaphention | 10 |
| Chloroxuron | 10 | Kresoxim methyl | 10 | Pyriproxyfen | 10 |
| Chlorpyrifos | 10 | Lenacil | 10 | Quizalofop P ethyl | 10 |
| Chlorpyrifos methyl | 10 | Malaoxon | 10 | Rimsulfuron | 10 |
| Clodinafop propargyl ester | 10 | Malathion | 10 | Spinosad | 20 |
| Clomazone | 10 | Mecarbam | 10 | Spiroxamin | 10 |
| Coumachlor | 20 | Mefenpyr diethyl | 10 | Tulfotep | 10 |
| Coumaphos | 10 | Mephosfolan | 10 | Temefos | 50 |
| Cyanazine | 10 | Mesosulfuron methyl | 10 | Tepraloxymid | 20 |
| Cycloate | 10 | Metalaxyl+Metalaxyl M | 10 | Terbufos | 50 |
| Cymoxanil | 25 | Metamitron | 10 | Terbutylazine | 10 |
| Cyproconazole | 10 | Metazachlor | 10 | Terbutryn | 10 |
| Cyprodinil | 20 | Metconazole | 10 | Tetrachlorvinphos | 10 |
| Demeton S methyl sulfoxide | 30 | Methacrifos | 10 | Thiabendazole | 10 |
| Diazinon | 10 | Methamidophos | 10 | Thiacloprid | 10 |
| Dibrom | 10 | Methidathion | 20 | Thiamethoxam | 10 |
| Diclorvos (DDVP) | 10 | Methiocarb | 10 | Thiophonate ethyl | 10 |
| Dicrotophos | 10 | Methomyl+Thiodicarb | 10 | Thiophonate methyl | 10 |
| Diethofencarb | 10 | Metolachlor | 10 | Tolyfluanide | 30 |
| Difenoconazole | 10 | Metoxuron | 10 | Tralkoxydim | 20 |
| Dimefox | 10 | Metribuzine | 10 | Triadimefon+Triadimenol | 10 |
| Dimethachlor | 10 | Metsulfuron methyl | 10 | Triallate | 10 |
| Dimethenamid | 10 | Mevinphos | 10 | Triasulfuron | 10 |
| Dimethoate+Omethoate | 10 | Molinate | 10 | Triazophos | 10 |
| Dimethomorph | 10 | Monocrotophos | 10 | Trifloxystrobin | 10 |
| Diniconazole | 10 | Monolinuron | 20 | Triflumizole | 10 |
| Dioxacarb | 10 | Myclobutanil | 20 | Triflusulfuron methyl | 15 |
| Diphenamid | 10 | Nicosulfuron | 10 | Triticonazole | 10 |
| Dipropetryn | 10 | Oxamyl | 10 | Vamidothion | 10 |
| Diuron | 10 | Paraoxon Ethyl | 10 | Vernolat | 10 |
| Dodine | 50 | Parathion ethyl | 20 | Zoxamide | 10 |

materials. All extraction studies and residue analysis of the examples by The Directorate of Food Control Laboratory of Izmir, connected to The Food, Farming and Livestock Ministry.

Pesticide standards are prepared at least a 90 % rate of purity. Extractions and clearance of the examples are generalized in accordance with AOAC (International Official Methods of Analysis) methods [19].

Preparation for analysis: 15 g examples were homogenized in a mechanical shredders. Other similars of the same example were put into same processes separately. Example amounts that put into extraction were taken from these homogenized examples after weighing.

Extraction: Whole examples were homogenized with steel blenders by shredding and 5 g of analyze examples from the main example were weighed and mixed with 10 mL of water and 15 mL of acetonitrile with 1 % acetic acid and strongly shaken for 1 min. Afterwards, 6 g of anhydrous magnesium sulfate and 1.5 g of sodium acetate is added into falcon tubes and after being shaken for 1 min, centrifugated for 5 min at 4000 rpm. As the next step, 8 mL of examples from the previous examples' high phases were collected for the cleaning process and transported into 15 mL falcon tubes and mixed with 1.2 g of anhydrous MgSO₄ and 0.4 g of primary secondary amine (PSA) and centrifuged for 5 min at 4000 rpm rate, once again. Later, the high phase was transported into vials and kept in a freezer until the device evaluations. As the last injections into LC-MS/MS and GC-MS/MS devices were conducted and residue rates were determined.

RESULTS AND DISCUSSION

In the study, the obtained average residue limits are considered as each sample has three replications, according to "Turkish Food Codex (TGK) communique of allowed maximum pesticide residue limits on food stuffs (Official Gazette: 21.01.2011-27822; Communique no: 2011/2). Each pesticide residue limits of TGK on the each sample are given one by one on tables. As the pesticide residue limits, defined by highly analytic devices such as GC-MS and LC-MS/MS 100 active pesticide substances in LC-MS/MS and 103 active pesticide substances in GC-MS were found in Cumin (*Nigella orientalis* L.) seeds. In the study, conducted through 2012 to 2013, any analyzable pesticide residues were not found in neither years' samples.

Mohamadin *et al.* [20] investigated *Nigella sativa* L. oil's effects on reducing PPr (widely used broad spectrum carbamate; insecticides) toxicity. They found by applying free radical scavenging method that, *Nigella sativa* L. oil reduces the PPr toxicity and reduced the oxidative stress zone in rats' brains.

Nagahban *et al.* [21] in their study, using *Artemisia sieberi* oil, found that, this oil sort had a potential to be used under the control of *C. maculatus*, *S. oryzae* and *T. castaneum*.

Rajendran and Sriranjini [22] investigated the use of organic products as fumigants for insect control in stocked products. There are several organic products developed to form a toxic effect against stored grain pests [23-25]; especially the ones in vegetable oil formation [26,27]. Tunc *et al.* [28] claimed that, as an alternative for traditional fumigants, vegetable tallow extracted from aromatic plants have been investigated widely and toxic effects of insecticides for stored products are taken

care for last ten years. Most of these studies investigate the effects of vegetable oil against these livings, who are in larvae and adulthood periods. They studied vegetable oil from 5 different plant species, which have an effect on these pests during their egg phase.

Shaaya *et al.* [29] in their study, questioned the fumigant toxicity of vegetable oil from five different aromatic plant species, against pests, effecting stored products. As *Tribolium castaneum* (L.), *Sitophilus oryzae* L., *Rhyzopertha dominica* (F.) and *Oryzae phksurinamensis* (L.) compared, *Tribolium castaneum* (L.) was reported to be more stable than the others.

Abou-Arab [30] had a study on pesticide residues, heavy metals and aflatoxin compounds on medical plants (mint, camomille, anise, cumin), used for both babies and adults. Samples were gathered from different markets of Egypt. As a result, while malathion, dimethoate and profenofos were observed to be in high rates; aldrin, dieldrin, chlordane and lindane were observed to be in low rates. Chlorpyrifos, parathion, diazinon and endosulfan were not observed to be in analyzable rates.

REFERENCES

1. M. Zohary, *Plant Systematics and Evolution*, **142**, 71 (1983).
2. B.H. Ali and G. Blunden, *Phytother. Res.*, **17**, 299 (2003).
3. E. Agradi, G. Fico, F. Cillo, C. Francisci and F. Tome, *Planta Med.*, **67**, 553 (2001).
4. Anonymous (2007); <http://bitkilerinyararlari.com/2007/07/rek-otu-nigella-sativa.html>.
5. P.H. Davis, *Flora of Turkey*, **1**, 98 (1965).
6. Anonymous (2007); <http://www.ihya.org/forum/mc.php?t2=metin&mln=2&an=5519>.
7. L. Kokoska, J. Havlik, I. Valterova, A. Nepovim, V. Rada and T. Vanek, *Flavour and Fragrance J.*, **20**, 419 (2005).
8. M. Gun, *Lokman Hekim J.*, **2**, 43 (2012).
9. H. Wagner and N.R. Fransworth, *Economic and Medicinal Plant Research, Plants and Traditional Medicine*, Academic Press, vol. 4 (1990).
10. C. Nergiz and S. Otle, *Food Chem.*, **48**, 259 (1993).
11. M.A. Farag, H.A. Gad, A.G. Heiss and L.A. Wessjohann, *Food Chem.*, **151**, 333 (2014).
12. K. Baser, B. Demirci, A. Donmez and Z. Ugurlu, *Planta Med.*, **76**, 1231 (2010).
13. A. Meziti, H. Meziti, K. Boudiaf, B. Mustapha and H. Bouriche, *World Academy of Science, Eng. Technol.*, **6**, 24 (2012).
14. A. Wajs and D. Kalembe, *J. Essential Oil Res.*, **22**, 232 (2010).
15. M. Sirma and I. Kadioglu, *Ziraat Fakültesi Dergisi*, **27**, 27 (2010).
16. E. Tulukcu and O. Sagdic, *Erciyes Univ. Fen Bilimleri Enstitüsü Dergisi*, **27**, 304 (2011).
17. J. Kim, S. Seo, S. Lee, S. Shin and I. Park, *J. Agric. Food Chem.*, **56**, 7316 (2008).
18. N. Ersoy, O. Tatli, S. Ozcan, E. Evcil, L.S. Coskun, E. Erdogan and G. Keskin, *Selçuk Tarım Gıda Bilimleri Dergisi*, **25**, 70 (2011).
19. S.J. Lehotay, *J. AOAC Int.*, **90**, 485 (2007).
20. A.M. Mohamadin, B. Sheikh, A.A. Abd El-Aal, A.A. Elberry and F.A. Al-Abbasi, *Pestic. Biochem. Physiol.*, **98**, 128 (2010).
21. M. Negahban, S. Moharrampour and F. Sefidkon, *J. Stored Prod. Res.*, **43**, 123 (2007).
22. S. Rajendran and V. Sriranjini, *J. Stored Prod. Res.*, **44**, 126 (2008).
23. H.F.C. Su, *J. Entomol. Sci.*, **25**, 16 (1990).
24. S.N. Mukherjee and M. Joseph, *J. Med. Arom. Plant Sci.*, **22-23**, 154 (2000).
25. M.K. Chaubey, *African J. Agric. Res.*, **2**, 596 (2007).
26. E. Shaaya, U. Ravid, N. Paster, B. Juven, U. Zisman and V. Pissarev, *J. Chem. Ecol.*, **17**, 499 (1991).
27. T.L.S. Ngamo, A. Goudoum, M.B. Ngassoum, G.M. Lognay, F. Malaisse and T. Hance, *African J. Agric. Res.*, **2**, 164 (2007).
28. I. Tunc, B.M. Berger, F. Erler and F. Dagli, *J. Stored Prod. Res.*, **36**, 161 (2000).
29. E. Shaaya, M. Kostjukovski, J. Eilberg and C. Sukprakarn, *J. Stored Products Res.*, **33**, 7 (1997).
30. A.A.K. Abou-Arab, *Food Chem.*, **65**, 509 (1999).