

Synthesis, Characterization and Pharmacological Activity of Phenoxy Acetic Acid and Pyrazinium Chlorochromate

K. ANBARASU and K.K. ILAVENIL*

Department of Chemistry, Arignar Anna Government Arts College, Musiri-621 211, India

*Corresponding author: E-mail: illavenil81@gmail.com

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This study describes the synthesis and antibacterial, antifungal activity of phenoxy acetic acid and pyrazinium chlorochromate. The compounds were characterized by infrared and ultra violet visible spectral data. These compounds were reviewed for antibacterial and antifungal activity against *Streptococcus*, *Entrococcus*, *Bacillus cereus*, *Proteus vulgaris*, *Mycobacterium tuberculosis*, *Azotobacter*, *E. coli*, *Pseudomonas aeruginosa*, *Candida albicans*, *A. niger*, *Fusarium* and *Trichoderma* by disc diffusion method.

Keywords: Phenoxy acetic acid, Pyrazinium chlorochromate, Biological activities.

INTRODUCTION

The basic molecular structures of the heterocyclic system exhibit a broad range of biological and pharmacological activities [1]. Many new drugs are synthesized using these moieties. The over practice of antimicrobial drugs have produced more impedance to the bacteria and fungi. As a result many infectious diseases have come to light. In order to prevent the spreading of these diseases, the pharmacists are in the need to develop a unique, wide-ranging of antimicrobial agents [2-4]. The heterocyclic ring consisting of nitrogen or sulphur atoms are researched for their physicochemical properties. The sensitivity of these heterocyclic compounds towards the biological properties plays a key role in the kinetic study [5,6].

The literature survey reveals that the pyrazine derivatives displays numerous properties like antibacterial [7], anti-inflammatory [8], analgesic [9], ulcerogenic potential [10], antituberculosis [11], antimicrobial [12], antifungal, antioxidant, anticonvulsants [13], antiproliferative assay [14], serotonin activity [15], cytotoxic properties [16,17], antipolytic activity [18], antidiabetic and antihistamines [19]. Several phenoxy derivatives illustrate antimycobacterial [20], antiviral [21], antituberculosis activity [22], hypolipidemic activity [23], antinociceptive [24], anti-inflammatory activity, antifungal activities [25], antioxidant activity [27], antidiabetic property [28].

Aspergillus niger and *Fusarium* species are plant pathogens and they affect the plant growth. The mycotoxins [29] produced by these are prone to cause lung defilement, ear infections and has been disclosed in HIV patients [30]. Phenoxy acetic acid, pyrazinium chlorochromate compounds are screened for their biological activities towards few Gram-positive, Gram-negative bacteria and fungi.

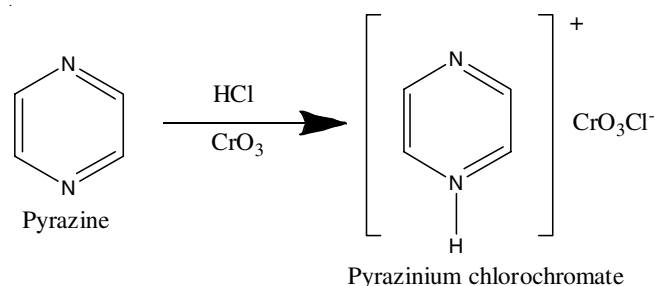
EXPERIMENTAL

AnalaR grade reagents are used for the synthesis of phenoxy acetic acid and pyrazinium chlorochromate. The melting points of these two compounds were determined by Thomas Hoover capillary melting point instrument. The IR spectra were recorded on FTIR Bruker Alpha and UV-visible spectra were reported using PerkinElmer Lambda 365 spectrophotometer.

Synthesis of pyrazinium chlorochromate ($C_4N_2H_5CrO_3Cl$):

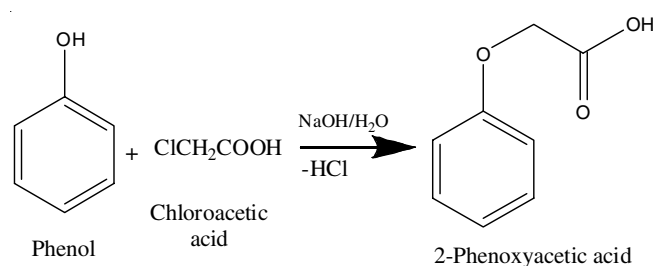
An orange coloured solid was prepared by adding 1.30 mmol of chromium trioxide and 2.4 mmol of 12 M HCl in 2 mL of doubly distilled water with constant stirring to a solution of 2.24 mmol of pyrazine and 2.28 mmol of 12 M HCl in 4 mL of water, in ice bath. After constant stirring for 2 h at 0 °C an orange coloured solid [31] obtained was filtered and then dried. The recrystallized solid (**Scheme-I**) from acidified water was used throughout the work (m.p. 148 to 150 °C).

Preparation of phenoxy acetic acid ($C_8H_8O_3$): To 11.47 mmol of sodium hydroxide 5 mmol of phenol was added and



Scheme-I: Synthesis of pyrazinium chlorochromate (PzCC)

diluted. 6 mmol of chloroacetic acid was dissolved in warm water and added to the above mixture and stirred constantly for 2 h at 60 to 80 °C. 35 % Hydrochloric acid was added and vacuum fractionated using benzene as solvent. The white coloured solid obtained (**Scheme-II**) was recrystallized using water and ethanol mixture (m.p. 98.2 to 98.9 °C) [32].



Scheme-II: Synthesis of phenoxy acetic acid

Phenoxy acetic acid: m.f. $C_8H_8O_3$, m.p. 98.2-99.0 °C, m.w. 152.15, Yield 98.5 %. IR (KBr, ν_{max} , cm^{-1}) 3300, 2467.70, 859.16, 3000, 1441.72, 3120, 1607.11, 1172.25, 1348.95, 1235.27 1043.36. The UV-visible spectrum showed absorbance peaks at 216.85 nm (s), 268.70 nm (w), transmittance peaks at 216.85 nm (s), 268.70 nm (s).

Pyrazinium chlorochromate: m.f. $C_4N_2H_5CrO_3Cl$, m.p. 148-154 °C, Yield 61%. IR (KBr, ν_{max} , cm^{-1}) 3113, 3049, 2955, 2768, 2700, 2085, 1614, 1490, 1373, 608, 1160, 770. The UV-visible spectrum showed peaks at 312 nm (m,b), 267 nm (s), transmittance peaks at 260.05 nm (s), 350.45 nm (b), 772.80 nm (w).

Determination of biological potential: The solution of the phenoxy acetic acid and pyrazinium chlorochromate were

prepared in various concentrations like 25, 50, 75 and 100 μL by dissolving in dimethyl sulphoxide solvent.

Disc diffusion antibiotic sensitivity testing: The synthesized compounds were screened for antibacterial activity against *Streptococcus*, *Enterococcus*, *Bacillus cereus*, *Proteus vulgaris*, *Mycobacterium tuberculosis*, *Azotobacter*, *E. coli*, *Pseudomonas aeruginosa*, *Candida albicans*, *A. niger*, *Fusarium* and *Trichoderma* using the agar disc diffusion method [33-36].

Bacterial inoculums containing approximately 10^5 - 10^6 colony forming units (CFU)/mL were used at 37 °C. The culture medium (Agar medium) was composed of peptone 5 g/L, NaCl-5 g/L, Agar-15 g/L, Beef extract-3 g/L, Yeast extract-1.5 g/L. The pH was maintained to be neutral at 7. All the materials were diluted in doubly distilled water and the medium was sterilized at 121 °C for 20 min at pressure 15 psi. The solutions of the chemical compounds were prepared at the concentration of 25, 50, 75 and 100 μL in DMSO.

The agar plate was incubated for 20 to 28 h at 37-38 °C for bacteria and 25 °C for fungi by following the standard procedure. Ciprofloxacin was selected as the standard and the zone of inhibition were measured and compared with the controls. Bacterial cultures were obtained from Eumic analytical Lab and Research Institute, Tiruchirappalli. Bacterial strains were maintained on Nutrient agar slants (Hi media) at 4 °C.

RESULTS AND DISCUSSION

As the concentration of phenoxy acetic acid and pyrazinium chlorochromate increased from 25 to 100 μL . The lethal zone of the bacteria and fungi also increased (Tables 1 and 2). The inhibition for the organic compound phenoxy acetic acid was high at 100 μL for (Gram-positive bacteria) *Mycobacterium tuberculosis*, *Proteus vulgaris* (Gram-negative bacteria) and fungi, *Aspergillus niger*, *Fusarium* and *Trichoderma* than the control of standard drug ciprofloxacin. Likewise for heterocyclic pyrazinium chlorochromate compound the impedance was more for (Gram-positive bacteria) *Mycobacterium tuberculosis*, (Gram-negative bacteria) *Proteus vulgaris* and fungi as discussed above.

Conclusion

At higher concentration (100 μL) the phenoxy acetic acid exhibited good resistant towards the organisms like *Bacillus*

TABLE-1
INHIBITION ZONE FOR PHENOXY ACETIC ACID

Organisms	Gram stain	Control 100 μL added and zone of inhibition (mm/mL)				
		25 μL	75 μL	50 μL	100 μL	Control
<i>Streptococcus</i>	Gram-positive bacteria	10	12	14	16	30
<i>Bacillus cereus</i>	Gram-positive bacteria	12	14	16	18	20
<i>Enterococcus</i>	Gram-positive bacteria	10	12	14	16	20
<i>Proteus vulgaris</i>	Gram-negative bacteria	10	12	14	16	15
<i>Micrococcus</i>	Gram-positive bacteria	10	11	13	16	30
<i>Mycobacterium tuberculosis</i>	Gram-positive bacteria	12	15	18	20	22
<i>Azotobacter</i>	Gram-negative bacteria	12	14	16	18	25
<i>E. coli</i>	Gram-negative bacteria	10	12	14	16	25
<i>Pseudomonas aeruginosa</i>	Gram-negative bacteria	12	15	18	20	25
<i>Candida albicans</i>	Fungi	12	14	16	19	32
<i>A. niger</i>	Fungi	—	10	11	12	12
<i>Fusarium</i>	Fungi	11	13	16	18	12
<i>Trichoderma</i>	Fungi	10	11	12	14	12

TABLE-2
INHIBITION ZONE FOR PYRAZINIUM CHLOROCHROMATE

Organisms	Gram stain	Control 100 µL added and zone of inhibition (mm/mL)				
		25 µL	75 µL	50 µL	100 µL	Control
<i>Streptococcus</i>	Gram-positive bacteria	10	12	14	16	28
<i>Bacillus cereus</i>	Gram-positive bacteria	12	14	16	18	22
<i>Enterococcus</i>	Gram-positive bacteria	12	15	18	20	22
<i>Proteus vulgaris</i>	Gram-negative bacteria	14	17	21	25	15
<i>Micrococcus</i>	Gram-positive bacteria	–	10	11	12	20
<i>Mycobacterium tuberculosis</i>	Gram-positive bacteria	14	16	18	20	20
<i>Azotobacter</i>	Gram-negative bacteria	10	12	14	16	25
<i>E. coli</i>	Gram-negative bacteria	12	14	16	18	25
<i>Pseudomonas aeruginosa</i>	Gram-negative bacteria	10	11	13	15	23
<i>Candida albicans</i>	Fungi	12	15	18	21	30
<i>A. niger</i>	Fungi	10	11	12	14	12
<i>Fusarium</i>	Fungi	–	10	11	12	12
<i>Trichoderma</i>	Fungi	10	12	14	16	14

cereus, *Proteus vulgaris*, *Mycobacterium tuberculosis*, *Aspergillus niger*, *Fusarium*, *Trichoderma* and pyrazinium chlorochromate were resistant towards *Enterococcus*, *Proteus vulgaris*, *Mycobacterium tuberculosis*, *Aspergillus niger* and *Fusarium*, *Trichoderma*.

CONFLICT OF INTEREST

The authors declare that there is no conflict of interests regarding the publication of this article.

REFERENCES

- A.P. Taylor, R.P. Robinson, Y.M. Fobian, D.C. Blakemore, L.H. Jones and O. Fadeyi, *Org. Biomol. Chem.*, **14**, 6611 (2016); <https://doi.org/10.1039/C6OB00936K>.
- R. Chawla, U. Sahoo, A. Arora, P.C. Sharma and V. Radhakrishnan, *Acta Pol. Pharm. Drug Res.*, **67**, 55 (2010).
- A.R. Katritzky, *Chem. Rev.*, **104**, 2125 (2004); <https://doi.org/10.1021/cr0406413>.
- M.E. Azab, M.M. Youssef and E.A. El-Bordany, *Molecules*, **18**, 832 (2013); <https://doi.org/10.3390/molecules18010832>.
- K.G. Sekar and G. Manikandan, *Oxid. Commun.*, **35**, 577 (2012).
- B.H. Asghar, *Arab. J. Chem.*; <https://doi.org/10.1016/j.arabjc.2015.04.006>.
- H. Foks, D. Pancechowska-Ksepko, A. Kedzia, Z. Zwolska, M. Janowiec and E. Augustynowicz-Kopec, *II Farmaco*, **60**, 513 (2005); <https://doi.org/10.1016/j.farmac.2005.05.002>.
- E. Abignente, *Actual Chim. Ther.*, **18**, 193 (1991).
- P. Gund and T.Y. Shen, *J. Med. Chem.*, **20**, 1146 (1977); <https://doi.org/10.1021/jm00219a007>.
- E. Abignente, P. de Caprariis, M.G. Rimoli, L. Avallone, L. Gomez Paloma, F. Rossi, M. D'Amico, V. Calderaro and C. Parrillo, *Eur. J. Med. Chem.*, **28**, 337 (1993); [https://doi.org/10.1016/0223-5234\(93\)90150-D](https://doi.org/10.1016/0223-5234(93)90150-D).
- M.V. Baldwin, D.A. Sonia, T.J. Sindhu, C. Meena, A.R. Bhar and K. Krishnakumar, *World J. Pharm. Pharm. Sci.*, **3**, 1124 (2014).
- M. Gür, N. Sener, H. Muglu, M.S. Çavus, O.E. Özkan, F. Kandemirli and I. Sener, *J. Mol. Struct.*, **1139**, 111 (2017); <https://doi.org/10.1016/j.molstruc.2017.03.019>.
- K. Nishie, A.C. Weiss Jr. and A.C. Keyl, *Toxicol. Appl. Pharmacol.*, **17**, 244 (1970); [https://doi.org/10.1016/0041-008X\(70\)90149-3](https://doi.org/10.1016/0041-008X(70)90149-3).
- L. Racané, S. Kraljevic Pavelic, I. Ratkaj, V. Stepanic, K. Pavelic, V. Tralic-Kulenovic and G. Karminski-Zamola, *Eur. J. Med. Chem.*, **55**, 108 (2012); <https://doi.org/10.1016/j.ejmech.2012.07.005>.
- B.V. Clineschmidt, J.C. McGuffin and A.B. Pflueger, *Eur. J. Pharmacol.*, **44**, 65 (1977); [https://doi.org/10.1016/0014-2999\(77\)90117-0](https://doi.org/10.1016/0014-2999(77)90117-0).
- N. Dias, U. Jacquemard, B. Baldeyrou, A. Lansiaux, J.-F. Goossens, C. Bailly, S. Routier and J.-Y. Mèroux, *Eur. J. Med. Chem.*, **40**, 1206 (2005); <https://doi.org/10.1016/j.ejmech.2005.07.003>.
- R.S. Kumar, I.A. Arif, A. Ahamed and A. Idhayadhulla, *Saudi J. Biol. Sci.*, **23**, 614 (2016); <https://doi.org/10.1016/j.sjbs.2015.07.005>.
- P.P. Lovisolo, G. Briatico-Vangosa, G. Orsini, R. Ronchi, R. Angelucci and G. Valzelli, *Pharmacol. Res. Commun.*, **13**, 151 (1981); [https://doi.org/10.1016/S0031-6989\(81\)80016-1](https://doi.org/10.1016/S0031-6989(81)80016-1).
- C.P. Meher, A.M. Rao and Md. Omar, *Asian J. Pharm. Sci. Res.*, **3**, 43 (2013).
- M. Shahar Yar, A.A. Siddiqui and M.A. Ali, *Bioorg. Med. Chem. Lett.*, **16**, 4571 (2006); <https://doi.org/10.1016/j.bmcl.2006.06.021>.
- M. Shahar Yar, M.A. Bakht, A.A. Siddiqui, M.M. Abdullah and E. De Clercq, *J. Enzym. Inhib. Med. Chem.*, **24**, 876 (2009); <https://doi.org/10.1080/14756360802447917>.
- M.A. Ali, M.S. Yar, M. Kumar and G.S. Pandian, *Nat. Prod. Res.*, **21**, 575 (2007); <https://doi.org/10.1080/14786410701369367>.
- S.N. Mokale, M.C. Nevase, N.S. Sakle, P.N. Dube, V.R. Shelke, S.A. Bhavale and A. Begum, *Bioorg. Med. Chem. Lett.*, **24**, 2155 (2014); <https://doi.org/10.1016/j.bmcl.2014.03.030>.
- G. Turan-Zitouni, L. Yurttas, Z.A. Kaplancikli, Ö.D. Can and Ü. Demir Özkay, *Med. Chem. Res.*, **24**, 2406 (2015); <https://doi.org/10.1007/s00044-014-1309-1>.
- A. Iqbal, H. Siddiqui, C. Ashraf, M. Ahmad and G. Weaver, *Molecules*, **12**, 245 (2007); <https://doi.org/10.3390/12020245>.
- M. Urban, J. Sarek, M. Kvasnica, I. Tislerova and M. Hajduch, *J. Nat. Prod.*, **70**, 526 (2007); <https://doi.org/10.1021/np060436d>.
- M. Kucerova-Chlupacova, M. Dosedel, J. Kunes, M. Soltesova-Prnova, M. Majekova and M. Stefek, *Monatsh. Chem.*, **149**, 921 (2018); <https://doi.org/10.1007/s00706-018-2146-6>.
- X. Wang, T. Zhao, B. Yang, Z. Li, J. Cui, Y. Dai, Q. Qiu, H. Qiang, W. Huang and H. Qian, *Bioorg. Med. Chem. Lett.*, **23**, 132 (2015); <https://doi.org/10.1016/j.bmc.2014.11.016>.
- A. Ghafoor and S.A.J. Khan, List of Diseases of Economic Plants in Pakistan, Ministry of Food and Agriculture, Islamabad, Pakistan, pp. 26 (1976).
- I. Rzewnicki, E. Olszewska and D. Rogowska-Szadkowska, *Med. Sci. Monit.*, **18**, RA17 (2012); <https://doi.org/10.12659/MSM.882505>.
- G.L. Gard, H.B. Davis, R.M. Sheets, J.M. Brannfors and W.W. Paudler, *Heterocycles*, **20**, 2029 (1983); <https://doi.org/10.3987/R-1983-10-2029>.
- C.F. Koelsch, *J. Am. Chem. Soc.*, **53**, 304 (1931); <https://doi.org/10.1021/ja01352a042>.
- Atta-ur-Rahman, M.I. Choudhary and W.J. Thomsen, Bioassay Techniques for Drug Development, Harwood Academic Publishers, The Netherlands, p. 16 (2001).
- K.M. Khan, Z.S. Saify, M.Z. Khan, Zia-Ullah, M.I. Choudhary, Atta-ur-Rahman, S. Perveen, Z.H. Chohan and C.T. Supuran, *J. Enzym. Inhib. Med. Chem.*, **19**, 373 (2004); <https://doi.org/10.1080/14756360409162453>.