

Air Pollution Tolerance Index to Assess the Pollution Tolerance Level of Plant Species in Industrial Areas

SUVENDU KUMAR DASH^{1,*} and ADITYA KISHORE DASH²

¹Department of Environmental Engineering, Institute of Technical Education and Research, Siksha 'O' Anusandhan University, Bhubaneswar-751030, India

²Biofuel and Bioprocessing Research Centre, Institute of Technical Education and Research, Siksha 'O' Anusandhan University, Bhubaneswar-751030, India

*Corresponding author: E-mail: suvendudash84@gmail.com

Received: 21 August 2017;	Accepted: 14 October 2017;	Published online: 30 November 2017;	AJC-18679

The objective of the present research was to examine the air pollution tolerance index (APTI) of 20 plant species near an industrial complex. The highest APTI was calculated for plant *Dalbergia sisoo* (25.75) and the lowest was for *Artocarpus* sp. (11.40). Relative water content for different plant species varied between 52.45 mg/g (*Artocarpus* sp.) to 89.56 mg/g (*Ailanthus excels*) and pH between 3.49 (*Pongamia pinnata*) to 7.01 (*Dalbergia sisoo*). The chlorophyll content was found maximum in *Mangifera indica* (14.23 mg/g) and minimum in *Eucalyptus* sp. (5.34 mg/g). The highest ascorbic acid was recorded in *Dalbergia sisoo* (9.05 mg/g) and lowest in *Gmelina arborea* (4.36 mg/g). Further, both total chlorophyll and relative water contents were significantly correlated with APTI (r = 0.7356 and r = 0.8302, respectively). It was calculated that there is no significant correlation of pH with APTI (r = 0.1407), whereas ascorbic acid was significantly correlated with APTI (r = 0.8522). Present study provides a basic idea for screening and selecting plant species for plantation in industrial areas.

Keywords: Air pollution tolerance index (APTI), Ascorbic acid, Chlorophyll, Relative water content.

INTRODUCTION

Air pollution in urban areas is gradually becoming a great challenge. Now a day's industrialization and globalization are the key factor which causes serious air pollution problems [1]. Continuous increase in human population, increasing in road transportation, vehicular traffic and industrialization have resulted an increase in the concentration of gaseous and particulate matter in the atmosphere [2-6]. The overall physiology of plants has been affected due to the particulate matters and gaseous pollutants presence in the atmosphere beyond the threshold limit [7,8]. A number of publication are reported to evaluate the response of traffic load on plants [9]. The ability of plants to remove pollutants from the air and act as sink for air contaminates were explored [10]. Green plants has an important role to mitigate the air pollution is also well known [11] but urban developmental activities is replacing maximum vegetation areas with concrete buildings [12,13]. Gaseous pollutants such as sulfur dioxide, nitrogen dioxide, carbon monoxide and particulate matters released from combustion activities along with toxic metals, organic materials and radioactive isotopes goes to the atmosphere [14]. Plant absorbs NO₂ which is assimilated into nitrogenous compounds and utilizes as a major nutrient for its growth [15].

The leaf area of different plant provides a platform for absorption and accumulation of air pollutants which helps to minimize the pollution level in the environment [16,17] which varied from species to species. In the present study air pollution tolerance index (APTI) has been calculated for 20 different plant species by synthesizing the values of different biochemical parameters such as ascorbic acid, relative water, chlorophyll and leaf extract pH. Various studies on the APTI of plants for development of green belt have been conducted by many workers [18-20].

Rather than individual parameter the combination of all biochemical parameters gave a more reliable value to predict the tolerance of plant species towards different pollutants. By adopting this technique of choosing suitable species as a mitigation measure for air pollution, industrial areas could able to achieve the air quality standard [21-23].

EXPERIMENTAL

Leaf samples were collected from 20 different tree species from the study areas which were adjacent to the industrial and mining activities. For further analysis, leaf samples were collected, cleaned and then refrigerated at 22 °C under suitable condition as per the standard procedures. The list of plants selected for the study with their scientific name and family are given in Table-1.

TABLE-1

LIST OF PLANTS STUDIED				
S. No.	Common name	Scientific name	Family	
1	Mango	Mangifera indica	Anacardiaceae	
2	Teak	Tectona grandis	Verbenaceae	
3	Peepal	Ficus religiosa	Moraceae	
4	Bela	Aegle marmelos	Rutaceae	
5	Kusum	Sghleichera oleosa	Sapindaceae	
6	Sal	Shorea robusta	Diplterocarpaceae	
7	Bamboo	Bambusa bamboos	Poaceae	
8	Panasa	Artocarpus	Moraceae	
		hetterophyllus		
9	Margosa	Azadirachta indica	Meliaceae	
10	Krushnachuda	Delonix regia	Ceasal Pinaceae	
11	Guava	Psidium guajava	Myrtaceae	
12	Gumhari	Gmelina arborea	Verbenaceae	
13	Kendu	Diospyros melanoxylon	Ebenaceae	
14	Sishoo	Dalbergia sisoo	Fabeceae	
15	Karanja	Pongamia pinnata	Fabeceal	
16	Mahalimb	Ailanthus excels	Simarubaceae	
17	Gulmohar	Ceasal pinia	Ceasalpiniaceae	
18	Ashoka	Polyalthia longifolia	Annonaceae	
19	Jack fruit	Artocarpus sp.	Moraceae	
20	Eucalyptus	Eucalyptus sp.	Myrtaceae	

Analysis of different leaf parameters

Ascorbic acid: Ascorbic acid also acts as an antioxidant which influences the plant's resistivity towards air pollution [24]. Ascorbic acid content (mg/g) was measured using the standard formula [25].

Total chlorophyll: Higher chlorophyll content has the potential to tolerate the concentrations of different pollutants.

Total chlorophyll content of the leaf samples were estimated as per the standard [26].

Leaf extract pH: It has been observed that the increasing level of leaf extract pH in plant leads to increase the tolerance level towards the air pollutants [27]. For pH measurement, 1 g of leaf sample was taken into a mortar and pestle and then homogenized with water. Finally, 20 mL of deionized water was added and allowed to settle for 15-20 min and then the pH was measured.

Relative water content: Relative water content is an indicator of water balances of a plant, because it expresses the absolute amount of water, which the plant requires to reach artificial full saturation [28]. Leaf samples were collected from the field and transported immediately to the laboratory. Leaf material (10 g) was taken and dried at 60-70 °C in a properly ventilated hot air oven for a period of 3 h. The weight of the leaf material was measured and the percentage of water content in the leaf material was calculated [29].

Air pollution tolerance index (APTI): The APTI was determined by calculating pH (P), ascorbic acid (A), total chlorophyll (T) and relative water content (R) of the leaf samples. The APTI was computed by using the given formula [30]:

$$APTI = A (T + P)/10$$

RESULTS AND DISCUSSION

The relative water content, pH, chlorophyll, ascorbic acid and APTI values for different plant species are given in Table-2.The relative water content varied between 52.45 mg/g (*Artocarpus* sp.) to 89.56 mg/g (*Ailanthus excels*). Higher amount of water is maintained the physiological balance in a plant body against the pollutants where the rate of transpiration iscomparatively high. The variation of relative water content is based on the atmospheric conditions, humidity and availability of moisture content in soil.

Leaf extract pH varied between 3.49 (*Pongamia pinnata*) to 7.01 (*Dalbergia sisoo*). pH plays a significant role in regulating

BIOCHEMICAL PARAMETERS AND APTI VALUES FOR DIFFERENT PLANT SPECIES IN THE STUDY AREA						
S. No.	Scientific name	Relative water content (%)	pH of leaf extracts	Chlorophyll content (mg/g)	Ascorbic acid content (mg/g)	APTI
1	Mangifera indica	78.65	6.85	14.23	7.69	24.08
2	Tectona grandis	64.61	5.92	12.56	6.43	18.34
3	Ficus religiosa	79.69	5.12	11.53	8.92	22.82
4	Aegle marmelos	81.25	4.98	10.96	6.13	17.90
5	Sghleichera oleosa	79.21	6.06	13.41	6.99	21.53
6	Shorea robusta	73.98	4.93	12.98	7.98	21.69
7	Bambusa bamboos	64.76	6.21	8.69	7.56	17.74
8	Artocarpus hetterophyllus	85.61	5.23	12.98	8.65	24.31
9	Azadirachta indica	75.62	5.23	11.96	8.56	22.28
10	Delonix regia	72.26	5.36	13.36	5.98	18.42
11	Psidium guajava	80.23	6.01	12.58	9.01	24.77
12	Gmelina arborea	69.75	3.68	11.69	4.36	13.68
13	Diospyros melanoxylon	73.24	5.89	11.57	8.72	22.55
14	Dalbergia sisoo	83.56	7.01	12.21	9.05	25.75
15	Pongamia pinnata	79.23	3.49	12.06	8.23	20.72
16	Ailanthus excels	89.56	6.02	13.97	7.89	24.73
17	Ceasal pinia	76.35	4.92	10.83	5.74	16.68
18	Polyalthia longifolia	62.34	6.02	7.24	7.64	16.36
19	Artocarpus sp.	52.45	6.32	6.92	4.65	11.40
20	Eucalyptus sp.	57.83	6.89	5.34	6.53	13.77

TABLE-2 BIOCHEMICAL PARAMETERS AND APTI VALUES FOR DIFFERENT PLANT SPECIES IN THE STUDY AREA

 SO_2 sensitivity of plants. The plant's tolerance level is directly proportional to the higher level of leaf extract pH which helps a plant to sustain in polluted conditions. The importance of pH in mediating physiological responses to stress was another reason for including it in APTI component [31].

Chlorophyll content depends on the rate of photosynthesis and the amount of nutrient available in the soil [32]. Chlorophyll was found to be maximum in *Mangifera indica* (14.23 mg/g) and minimum in *Eucalyptus* sp. (5.34 mg/g). Leaf chlorophyll changes can serve as relative indicators of environmental quality [33].

Ascorbic acid acts as a strong reluctant and it activates several physiological and defense characteristics in the plant [34]. The highest ascorbic acid was recorded in *Dalbergia sisoo* (9.05 mg/g) and lowest in *Gmelina arborea* (4.36 mg/g). The APTI values were recorded highest in *Dalbergia sisoo* (25.75) and lowest in *Artocarpus* sp. (11.40). Low APTI values were generally sensitive to air pollutants and *vice-versa*. Air pollution tolerance index provides an idea for screening or selecting of plants for their efficiency as well as susceptibility to different air pollutants. From the above results, it was observed that out of 20 plant species, 5 species belongs to sensitivity category and 15 species belongs to intermediate category, when compared with the standard APTI value.

Further, a correlation matrix among different parameters along with APTI has been calculated and shown in Table-3. There is a significant correlation between chlorophyll content and relative water content with APTI value (r = 0.7356 and r = 0.8302). There is no significant correlation of pH with APTI, whereas ascorbic acid was significantly correlated with APTI (r = 0.8522). Hence, APTI value has a strong and positive correlation with total chlorophyll, relative water content and ascorbic acid. This indicates that concentrations of different parameters like total chlorophyll, relative water content, pH and ascorbic acid influence the APTI value [35,36].

TABLE-3 CORRELATIONCOEFFICIENT OF PARAMETERS STUDIED					
	Total chlorophyll	RWC	pН	AA	APTI
Total					
chlorophyll	1				
RWC	0.7885**	1			
pН	-0.2392	-0.2068	1		
AA	0.3290	0.5463	0.19006	1	
APTI	0.7356**	0.8302**	0.14078	0.8522**	1
**Correlation (r) is strong and significant; RWC = Relative water					

Figs. 1-3 show the correlation graph of APTI with total

Conclusion

Determination of air pollution tolerance index (APTI) value is an useful technique to trace the tolerance and susceptibility of different species towards air pollution. The present study revealed that different plants respond differently to air pollution. Therefore biomonitoring is an important tool to find out the impact of air pollution on different plant species particularly in industrial areas. To fulfill the concept of green-belt

chlorophyll, relative water content and ascorbic acid content.



development programme, plants with high APTI values are recommended which are potential to grow under air pollution condition. The present research revealed the efficiency of some selected plant species for plantation near industrial areas to minimize the ambient air pollution load.

REFERENCES

- L.S. Vailshery, M. Jaganmohan and H. Nagendra, *Urban Forest Urban Green.*, **12**, 408 (2013);
- <u>https://doi.org/10.1016/j.ufug.2013.03.002</u>.
 P.O. Agbaire and E. Esiefarienrhe, *J. Appl. Sci. Environ. Manage.*, 13, 11 (2009);

https://doi.org/10.4314/jasem.v13i1.55251.

- D. Sahoo, A.K. Dash and S.K. Sahu, Int. J. Eng. Sci. Res. Technol., 6, 429 (2017).
- 4. S.K. Dash and A.K. Dash, Pollut. Res., 34, 181 (2015).
- S.K. Dash and A.K. Dash, *Indian J. Sci. Technol.*, 8, 1 (2015); https://doi.org/10.17485/ijst/2015/v8i35/81468.
- A.K. Dash, S.K. Sahu, A. Pradhan, S.K. Dash and R.N. Kolli, *Asian J. Chem.*, 29, 1150 (2017);
- https://doi.org/10.14233/ajchem.2017.20477.
 T.W. Ashenden and I.A.D. Williams, *Environ. Pollut.*, 21, 131 (1980); https://doi.org/10.1016/0143-1471(80)90041-0.
- V. Mejstrik, *Environ. Pollut.*, **21**, 73 (1980); https://doi.org/10.1016/0143-1471(80)90034-3.
- 9. P.G. Angold, *J. Appl. Ecol.*, **34**, 409 (1997); https://doi.org/10.2307/2404886.
- 10. A.K. Tripathi and M. Gautam, J. Environ. Biol., 28, 127 (2007).
- 11. M.N. Islam, K.S. Rahman, H.M. Bahar, M.A. Habib, K. Ando and N. Hattori, *Urban Forest Urban Green.*, **11**, 460 (2012);
- <u>https://doi.org/10.1016/j.ufug.2012.06.004</u>.
 12. A. Onishi, X. Cao, T. Ito, F. Shi and H. Imura, *Urban Forest Urban Green.*, 9, 323 (2010);
- https://doi.org/10.1016/j.ufug.2010.06.002. 13. H. Sugawara and T. Takamura, *Boundary-Layer Meteorol.*, **153**, 539 (2014); https://doi.org/10.1007/s10546-014-9952-0.
- T. Bhattacharya, L. Kriplani and S. Chakraborty, Univ. J. Environ. Res. Technol., 3, 199 (2013).
- Sulistijorini, Z.A. Mas'ud, N. Nasrullah, A. Bey and S. Tjitrosemito, *Hayati J. Biosci.*, **15**, 123 (2008); <u>https://doi.org/10.4308/hjb.15.3.123</u>.
- F.J. Escobedo, J.E. Wagner, D.J. Nowak, C.L. De la Maza, M. Rodriguez and D.E. Crane, *J. Environ. Manage.*, 86, 148 (2008); <u>https://doi.org/10.1016/j.jenvman.2006.11.029</u>.
- 17. A. Garg, P. Saxena and C. Ghosh, Int. J. Sci. Technol. Res., 4, 199 (2015).
- M. Govindaraju, R.S. Ganesh Kumar, V.R. Muthukumaran and P. Visvanathan, *Environ. Sci. Pollut. Res. Int.*, **19**, 1210 (2012); https://doi.org/10.1007/s11356-011-0637-7.
- 19. A. Gholami, A. Mojiri and H. Amini, J. Anim. Plant Sci., 26, 475 (2016).

- 20. K. Navjot, S. Simpy and N.C. Gupta, Int. J. Curr. Sci., 19, 181 (2016).
- K. De Ridder, V. Adamec, A. Bañuelos, M. Bruse, M. Bürger, O. Damsgaard, J. Dufek, J. Hirsch, F. Lefebre, J.M. Pérez-Lacorzana, A. Thierry and C. Weber, *Sci. Total Environ.*, **334-335**, 489 (2004); https://doi.org/10.1016/j.scitotenv.2004.04.054.
- 22. A. Lee and R. Maheswaran, J. Public Health (Bangkok), 33, 212 (2011); https://doi.org/10.1093/pubmed/fdq068.
- R.N. Lohe, B. Tyagi, V. Singh, P. Kumar Tyagi, D.R. Khanna and R.A. Bhutiani, *Global J. Environ. Sci. Manage.*, 1, 315 (2015).
- Y.J. Liu and H. Ding, WSEAS Trans. Environ. Dev., 4, 24 (2008).
 A. Begum and S. Harikrishna, E-J. Chem., 7(S1), S151 (2010);
- https://doi.org/10.1155/2010/398382.
- 26. D.I. Arnon, *Plant Physiol.*, **24**, 1 (1949); <u>https://doi.org/10.1104/pp.24.1.1</u>.
- 27. C.S. Chaudhary and D.N. Rao, *Proc. Indian Natl. Sci. Acad.*, **43**, 236 (1977).
- D.N. Sen and M.C. Bhandari, ed.: A.M. Althawadi, Ecological and Water Relation to Two *Citrullus* spp., In: Indian Arid Zone, Environmental Physiology and Ecology of Plants, pp. 203-228 (1978).
- 29. G. Klumpp, C.M. Furlan, M. Domingos and A. Klumpp, *Sci. Total Environ.*, **246**, 79 (2000);
- https://doi.org/10.1016/S0048-9697(99)00453-2.
 30. S.K. Singh and D.N. Rao, Proceedings of International Symposium on Air Pollution Control, IIT, Delhi, India, pp. 218-224 (1983).
- K. Arunpandiyan, S. Malathy and F. Mujeera, *Int. J. Educ. Sci. Res.*, 5, 33 (2016).
- J.N.B. Bell and C.H. Mudd, ed.: T.A. Mansfield, Effects of Air Pollutants on Plants, Cambridge University Press, London, pp. 87-103 (1976).
- G.A. Carter and A.K. Knapp, Am. J. Bot., 88, 677 (2001); https://doi.org/10.2307/2657068.
- 34. T. Keller and H. Schwager, *Eur. J. Forest Pathol.*, **7**, 338 (1977); https://doi.org/10.1111/j.1439-0329.1977.tb00603.x.
- M.U. Beg, M. Farooq, S.K. Bhargava, M.M. Kidwai and M.M. Lal, *Environ. Ecol.*, 8, 791 (1990).
- A. Chauhan, New York Sci. J., 3, 45 (2010); <u>https://doi.org/10.7537/marsnys030210.08</u>.