



NOTE

A New Isoflavonoid from the Stems of *Nicotiana tabacum* and its Anti-tobacco Mosaic Virus Activity

YANQING YE¹, QUNFANG ZHANG^{1,2}, WEI LIU², YONGKUAN CHEN², GUANG-YU YANG^{2,*} and QIUJEN HU^{1,*}

¹Key Laboratory of Chemistry in Ethnic Medicine Resources, State Ethnic Affairs Commission & Ministry of Education, Yunnan University of Nationalities, Kunming 650031, P.R. China

²Key Laboratory of Tobacco Chemistry of Yunnan Province, Yunnan Academy of Tobacco Science, Kunming 650106, P.R. China

*Corresponding author: E-mail: huqiufena@yahoo.com.cn

(Received: 20 June 2011;

Accepted: 21 June 2012)

AJC-11645

A new isoflavonoid (3',4',5'-trihydroxyl-5,7-dimethoxyl-isoflavonoid) was isolated from the stems of *Nicotiana tabacum*. Its structure was determined by means of HRESIMS, extensive ¹D and ²D NMR spectroscopic studies and chemical evidences. The anti-tobacco mosaic virus activity of the new isoflavonoid was also evaluated and it shows moderate anti-tobacco mosaic virus activity.

Key Words: Isoflavonoid, *Nicotiana tabacum*, Anti-tobacco mosaic virus activity.

Nicotiana tabacum L. belongs to solanaceae family. It is a perennial herbaceous plant originating from south America and it is one of the most commercially valued agricultural crops in the world^{1,2}. In addition to cigarette industry use, *N. tabacum* also contains many useful chemical compounds, such as sesquiterpenes^{3,4}, diterpenoids⁵⁻⁷, alkaloids^{8,9}, phenols¹⁰ and the like. The utilizations of these active compounds in the leaves and stems of *N. tabacum* were received more and more attentions.

Motivated by search for bioactive metabolites from this plant, an investigation on the chemical constituents of the stems of *N. tabacum* was carried out. As a result, a new isoflavonoid was isolated from this plant. In addition, the anti-tobacco mosaic virus (Anti-TMV) activity of the new isoflavonoid was evaluated. This article deals with the isolation, structural elucidation and biological activities of the new isoflavonoid.

IR spectra were obtained in KBr disc on a Bio-Rad Wininfrared spectrophotometer. ESI-MS were measured on a VG Auto Spec-3000 MS spectrometer. ¹H, ¹³C and ²D NMR spectra were recorded on Bruker DRX-500 instrument with TMS as internal standard. Column chromatography was performed on silica gel (200-300 mesh), or on silica gel H (10-40 mm, Qingdao Marine Chemical Inc., China). Second separate was used an Agilent 1100 HPLC equipped with ZORBAX-C₁₈ (21.2 × 250 nm, 7.0 μm) column and DAD detector.

The stems of *Nicotiana tabacum* L (tobacco stems) was collected from Yuxi County, Yunnan Province, P.R. China, in September, 2008.

Extraction and Isolation: The air-dried and powdered stems *N. tabacum* (2 kg) were extracted with 70 % aqueous ethanol (3 L × 3, 24 h each) at room temperature and the extract

was concentrated under vacuum condition. The dried extract (46.8 g) was applied to Si gel (200-300 mesh) column chromatography eluting with a CHCl₃-Me₂CO gradient system (9:1, 8:2, 7:3, 6:4, 5:5, 2:1) to give six fractions A-F. The separation of fraction C(CHCl₃- Me₂CO 7:3, 18.6 g) by Si gel column chromatography eluted with CHCl₃-Me₂OH (9:1 - 1:2) and preparative HPLC (35 % MeOH-H₂O, flow rate 12 mL/min) can obtained the new isoflavonoid. The structure of the compound was shown in Fig. 1 and its ¹H and ¹³C NMR spectroscopic data were listed in Table-1.

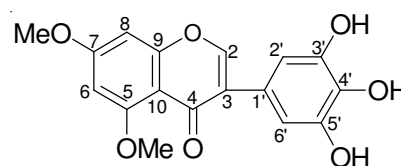


Fig. 1. Structure of the new isoflavonoid

Anti-tobacco mosaic virus assays: The anti-tobacco mosaic virus actives were tested using the half-leaf method¹¹. The inhibitory activities of new isoflavonoid against tobacco mosaic virus replication were tested using two approaches. First, the half-leaf method was used to test the antiviral activity in the local lesion host *N. glutinosa in vivo*. Then, the leaf-disk method was used to evaluate the antiviral activity of the compound in the systemic infection host *N. tabacum* cv. K326. Ningnanmycin, a commercial product for plant disease in China, was used as a positive control.

3',4',5'-Trihydroxyl-5,7-dimethoxyl-isoflavonoid was obtained as orange-yellow gum. The ESIMS spectrum

presented a molecular ion peak at m/z 329.0667 $[M-H]^-$ (calcd. 329.0661). The 1H NMR spectrum at δ_H 7.94 (s, 1H), 6.58 (d, $J = 2.2$, 1H), 6.49 (d, $J = 2.2$, 1H) and 7.02 (s, 2H) showed the presence of 3',4',5',5,7-substituted isoflavonoid nucleus¹² and this was supported by the ^{13}C NMR data (Table-1). In addition to isoflavonoid nucleus, two methoxyl signals (δ_H 3.75, s, 3.73 s; δ_C 56.7 q, 56.0 q) were also observed in its 1H and ^{13}C NMR spectrum. In the HMBC spectrum, the presence of cross peaks at δ_H 3.75 with δ_C 160.1 (C-5) and δ_H 3.73 with δ_C 162.0 (C-7) evidenced the attachment position of the two methoxyl groups at C-5 and C-7. Since no further aromatic carbon signals were evident, the substituents on a aromatic ring B should be three hydroxyl with 3,4,5-substituted¹² and the ^{13}C NMR spectrum of ring B (δ_C 122.7 s, 105.3 d, 146.8 s, 137.5 s) also supported this. Thus, the structure of the compound was established as shown (Fig. 2).

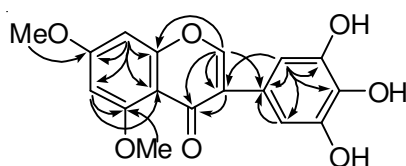


Fig. 2. Selected HMBC (\curvearrowright) correlations of new isoflavonoid

TABLE-1
 1H NMR AND ^{13}C NMR DATA OF THE COMPOUND IN $CDCl_3$

No.	δ_C (mult)	δ_H (mult, J , Hz)
2	154.1 d	7.94 s
3	123.1 s	
4	181.7 s	
5	160.1 s	
6	97.5 d	6.58 d, $J = 2.2$ Hz
7	162.0 s	
8	96.0 d	6.49 d, $J = 2.2$ Hz
9	158.2 s	
10	106.2 s	
1'	122.7 s	
2',6'	105.3 d	7.02, s
3',5'	146.8 s	
4'	137.5 s	
OMe-5	56.7 q	3.75, s
OMe-7	56.0 q	3.73, s

Since some of the flavonoids exhibited anti-virus activities^{13,14} 3',4',5'-trihydroxy-5,7-dimethoxyisoflavonoid was tested for its potencies in preventing anti-tobacco mosaic virus activity using the half-leaf method¹¹.

The antiviral inhibition rates of the compound at the concentration of 20 μM tested by the half-leaf method. The results showed that the compounds exhibited inhibition

activities against tobacco mosaic virus replication with inhibition rates of 92.8 %. The rates higher than that of the positive control, ningnanmycin (28.9 %).

N. glutinosa were pretreated with solutions of compounds or a solution of DMSO for 6.0 h before inoculation with tobacco mosaic virus. At the concentration of 20 μM , the compound showed potent protective effects to the host plants, with the inhibition rates ranging from 74.5-86.5 %. The results indicated that pretreatment with compound increase the resistance of the host plant to tobacco mosaic virus infection.

3',4',5'-trihydroxy-5,7-dimethoxyisoflavonoid: $C_{17}H_{14}O_7$, orange-yellow gum; UV (MeOH), λ_{max} (log ϵ) 356 (3.87), 310 (3.67), 246 (4.18), 210 (4.99) nm; IR (KBr, ν_{max} , cm^{-1}) 3384, 1658, 1615, 1548, 1508, 1426, 1125, 1047, 956, 863; 1H NMR and ^{13}C NMR data ($CDCl_3$, 500 MHz), Table-1; ESIMS m/z 329; HRESIMS (positive ion mode) m/z 329.0667 $[M-H]^-$ (calcd. 329.0661 for $C_{17}H_{13}O_7$).

ACKNOWLEDGEMENTS

This work was supported by program for Innovative Research Team (in Science and Technology) in University of Yunnan Province (IRTSTYN) and Green Chemistry and Functional Materials Research for Yunnan Research Team (2011HC008).

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