



## Flavonoids from the Fruits of *Camellia oleifera*

LI SUN<sup>1,2</sup>, ZHEN LIAO<sup>2</sup>, XUESEN LI<sup>1</sup>, HUAIXUE MU<sup>1</sup>, ZHANGYU CHEN<sup>2,\*</sup> and QIUFEN HU<sup>1,\*</sup>

<sup>1</sup>Key Laboratory of Chemistry in Ethnic Medicine Resources, State Ethnic Affairs Commission & Ministry of Education, Yunnan University of Nationalities, Kunming 650031, P.R. China

<sup>2</sup>Key Laboratory of Tobacco Chemistry of Yunnan Province, Yunnan Academy of Tobacco Science, Kunming 650106, P.R. China

\*Corresponding authors: E-mail: huqiufena@yahoo.com.cn

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A new flavonoid, 8-formyl-3',4',5'-trihydroxy-6,7-dimethoxyflavonoid (**1**), together with eight flavonoids (**2-8**) were isolated from the fruits of *Camellia oleifera*. The structures of compounds **1-9** were elucidated on the basis of extensive NMR and MS means. The anti-tobacco mosaic virus and antioxidant activity of **1** were evaluated. It showed anti-tobacco mosaic virus activity with inhibition rates of 85.7 % and antioxidant activity with an IC<sub>50</sub> value of 2.78 mg/mL.

**Key Words:** Flavonoids, *Camellia oleifera*, Anti-tobacco mosaic virus activity, Antioxidant activity.

### INTRODUCTION

*Camellia oleifera* Abel., a theaceous evergreen tree, is widely distributed and long time cultivated in south China. The seeds are important oil material for producing a kind of cooking oil (tea oil), whose beneficial unsaturated fatty acids are comparable to those of olive oil<sup>1</sup>. To obtain one ton of tea oil, four times of residue composing of the remaining grounded fruit and shell will be produced, which is called 'seed cake of *C. oleifera*'. As a big amount of by-product, the seed cake of *C. oleifera* is normally used as detergent, animal feeds or organic fertilizer, due to the containing of rich polyphenols, saponins, protein, polysaccharide, etc.<sup>2-5</sup>. With the aim of continuing efforts to identify bioactive natural products from the plants, a chemical investigation on the fruits of *C. oleifera* indigenous to the Dehong Prefecture of Yunnan Province, P.R. China, was carried out and a new flavonoid (**1**), together with nine known flavonoids (**2-9**) were isolated from the fruits of this plant. In addition, the anti-tobacco mosaic virus (anti-TMV) and antioxidant activity of compound **1** were evaluated. Described in this paper are their structure elucidation and biological activities.

### EXPERIMENTAL

IR spectra were obtained in KBr disc on a Bio-Rad Wininfrared spectrophotometer. ESI-MS were measured on a VG Auto Spec-3000 MS spectrometer. <sup>1</sup>H, <sup>13</sup>C and <sup>2</sup>D NMR spectra were recorded on Bruker DRX-500 instruments with TMS as internal standard. Column chromatography was performed on silica gel (200-300 mesh), or on silica gel H

(10-40 mm, Qingdao Marine Chemical Inc., China). On second separate used Agilent 1100 HPLC equipped with ZORBAX-C<sub>18</sub> (9.4 × 250 nm, 5.0 μm) column and DAD detector.

The fruits of *Camellia oleifera* Abel. was collected in Dehong Prefecture, Yunnan Province, P.R. China, in October 2010 and was identified by Prof. N. Yuan. A voucher specimen (No. YNNi 09-2-02) was deposited in our laboratory.

**Extraction and isolation:** The air-dried and powdered fruits of *C. oleifera* (1.5 kg) were extracted with 70 % aqueous ethanol (3.0 L × 3, 24 h each) at room temperature and the extract was concentrated under vacuum condition. The dried extract (102 g) was applied to Si gel (200-300 mesh) column chromatography eluting with a CHCl<sub>3</sub>-Me<sub>2</sub>CO gradient system (9:1, 8:2, 7:3, 6:4, 5:5, 2:1) to give six fractions A-F. The separation of fraction C (CHCl<sub>3</sub>-Me<sub>2</sub>CO 7:3, 18.6 g) by Si gel column chromatography eluted with CHCl<sub>3</sub>-Me<sub>2</sub>OH (9:1-1:2) yielded mixtures C1-V5. Fraction C2 (1.87 g) was subjected to preparative HPLC (35 % MeOH-H<sub>2</sub>O or 22 % CH<sub>3</sub>CN-H<sub>2</sub>O, flow rate 12 mL/min) to give **1** (15.6 mg), **2** (54.5 mg) and **3** (32.5 mg). The separation of fraction D (CHCl<sub>3</sub>-Me<sub>2</sub>CO 1:1, 28.4 g) by Si gel column chromatography eluted with CHCl<sub>3</sub>-Me<sub>2</sub>OH (9:1-1:2) yielded mixtures D1-D5. Fraction D3 (3.76 g) was subjected to preparative HPLC (15 % MeOH-H<sub>2</sub>O or 10 % CH<sub>3</sub>CN-H<sub>2</sub>O, flow rate 12 mL/min) to give **4** (36.4 mg) **7** (11.6 mg) and **8** (60.8 mg). Fraction D4 (5.26 g) was subjected to preparative HPLC (10 % MeOH-H<sub>2</sub>O or 7.5 % CH<sub>3</sub>CN-H<sub>2</sub>O, flow rate 12 mL/min) to give **5** (36.4 mg), **6** (28.4 mg) and **9** (50.2 mg).

**Antioxidant activity assay:** Antioxidant activity was determined by the detection of the oxidative products with

the 2',7'-dichlorofluorescein diacetate (DCFH) method reported previously<sup>6</sup>.

**Anti-TMV assays:** The anti-tobacco mosaic virus actives were tested using the half-leaf method<sup>7</sup>. The inhibitory activities of the compound **1** against tobacco mosaic virus replication were tested using two approaches. First, the half-leaf method was used to test the antiviral activity in the local lesion host *N. glutinosa in vivo*. Then, the leaf-disk method was used to evaluate the antiviral activity of the compound in the systemic infection host *N. tabacum* cv. K326. Ningnanmycin, a commercial product for plant disease in China, was used as a positive control.

### Spectral data

**8-Formyl-3',4',5'-trihydroxy-6,7-dimethoxyflavonoid (1):** C<sub>18</sub>H<sub>14</sub>O<sub>8</sub>, orange-yellow gum; UV (CH<sub>3</sub>OH), λ<sub>max</sub> (log ε) 210 (4.95), 258 (4.22), 292 (3.80), 370 (4.15) nm; IR (KBr, ν<sub>max</sub>, cm<sup>-1</sup>) 3464, 1685, 1650, 1624, 1522, 1480, 1465, 1127, 1095, 952, 870; <sup>13</sup>C NMR and <sup>1</sup>H NMR data (C<sub>5</sub>D<sub>5</sub>N, 500 MHz) (Table-1); positive ESIMS *m/z* 381 [M+Ma]<sup>+</sup>; HRESIMS *m/z* 381.0586 [M+Na]<sup>+</sup> (calcd. for C<sub>18</sub>H<sub>14</sub>O<sub>8</sub>Na, 381.0580).

## RESULTS AND DISCUSSION

A 70 % aq. ethanol extract prepared from the fruits of *C. oleifera* was subjected repeatedly to column chromatography on Si gel, Sephadex LH-20, RP-18 and Preparative HPLC to afford 8-formyl-3',4',5'-trihydroxy-6,7-dimethoxyflavonoid (**1**), kaempferol (**2**)<sup>8</sup>, quercetin (**3**)<sup>8</sup>, quercetin 3-*O*-β-glucopyranoside (**4**)<sup>8</sup>, leucoside (**5**)<sup>9</sup>, kaempferol-3-*O*-β-D-glucopyranosyl-(1→2)-β-D-glucopyranoside (**6**)<sup>10</sup>, isovitexin (**7**)<sup>11</sup>, luteolin-6-*C*-β-D-glucopyranoside (**8**)<sup>12</sup>, isosaponarin (**9**)<sup>12</sup>. The structure of the **1-9** was shown in Fig. 1 and the <sup>1</sup>H and <sup>13</sup>C NMR spectroscopic data of **1** were listed in Table-1.

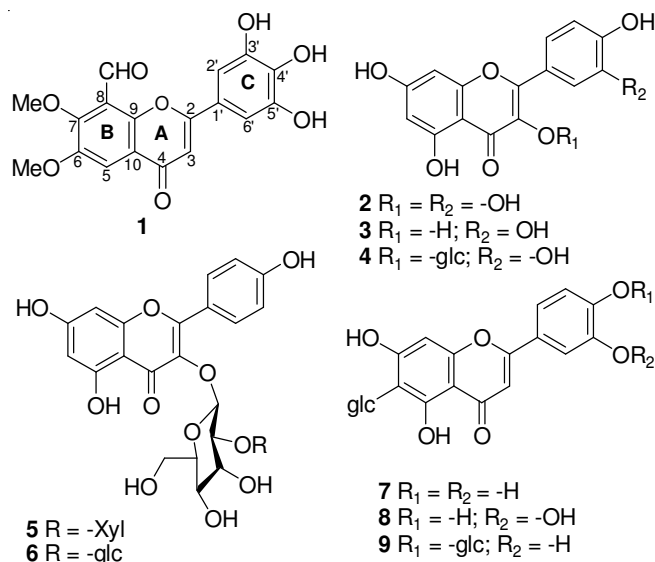


Fig. 1. Flavonoids from the fruits of *Camellia oleifera*

Compound **1** was obtained as an orange-yellow gum. It has the molecular formula C<sub>18</sub>H<sub>14</sub>O<sub>8</sub> from HRESIMS (*m/z*: 381.0580 [M+Na]<sup>+</sup>, calcd. 381.0586). The <sup>1</sup>H and <sup>13</sup>C NMR spectrum of **1** (Table-1) along with analysis of the DEPT spectra displayed 18 carbon signals and 14 proton signals, respec-

tively, corresponding to a flavonoid nucleus<sup>14</sup> (δ<sub>C</sub> 164.8, 106.2, 178.5, 122.8, 147.2, 159.1, 115.6, 152.1, 121.3, 123.1, 106.8, 147.5, 138.2, 147.5, 106.8), one aldehyde group (δ<sub>C</sub> 192.6; δ<sub>H</sub> 10.22), two methoxy groups (δ<sub>C</sub> 55.8, 61.8; δ<sub>H</sub> 3.80, 3.82) and two low-field hydroxyl protons (δ<sub>H</sub> 10.92 2H, 11.26 1H). The typical NMR signals at (δ<sub>H</sub> 7.06 s, 2H), (δ<sub>C</sub> 123.1 s, 106.8 d, 147.5 s, 138.2 s) observed in the <sup>1</sup>H NMR spectrum revealed that the aromatic ring C of **1** is a symmetrical 1,3,4,5-tetra-substituted phenolic moiety<sup>14</sup> and the HMBC of correlations (Fig. 2) of the hydroxyl proton signals, δ<sub>H</sub> 10.92 with C-2' (δ<sub>C</sub> 106.8), C-3' (δ<sub>C</sub> 147.5), C-4' (δ<sub>C</sub> 138.2) and δ<sub>H</sub> 11.26 with C-3' (δ<sub>C</sub> 147.5), C-4' (δ<sub>C</sub> 138.2), C-5' (δ<sub>C</sub> 147.5) suggested the attachment position of the three hydroxy groups at C-3', C-4' and C-5'. Since the substituents on ring C were evident, the surplus substituents (one aldehyde group, two methoxy groups) should be located at ring B. The HMBC correlations of aldehyde proton signal, δ<sub>H</sub> 10.22 with C-7 (δ<sub>C</sub> 159.1), C-8 (δ<sub>C</sub> 115.6), C-9 (δ<sub>C</sub> 152.1) suggested the placement of the formyl group at C-8. Two methoxy groups located at C-6 and C-7 was also supported by the HMBC correlation of the methoxy proton signals δ<sub>H</sub> 3.80 with C-6 (δ<sub>C</sub> 147.2) and δ<sub>H</sub> 3.82 with C-7 (δ<sub>C</sub> 159.1). Thus, the structure of **1** was established as shown in Fig. 2.

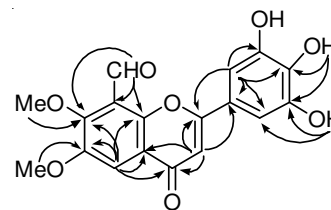


Fig. 2. Selected HMBC (↷) correlations of compound **1**

TABLE-1  
<sup>1</sup>H NMR AND <sup>13</sup>C NMR DATA (C<sub>5</sub>D<sub>5</sub>N) OF COMPOUNDS **1**

No.	Compound <b>1</b>	
	δ <sub>C</sub> (mult.)	δ <sub>H</sub> (mult, J, Hz)
2	164.8 s	
3	106.2 d	6.68 s
4	178.5 s	
5	122.8 d	
6	147.2 s	
7	159.1 s	
8	115.6 s	
9	152.1 s	
10	121.3 s	
1'	123.1 s	
2',6'	106.8 d	7.06, s
3',5'	147.5 s	
4'	138.2 s	
-CHO	192.6 s	10.22 s
-OMe-6	55.8 q	3.80 s
-OMe-7	61.8 q	3.82 s
-OH-3',5'		10.92 brs
Ar-OH-4'		11.26 brs

Since the flavonoids exhibited strong antioxidant activities<sup>15,16</sup> the antioxidant activity of compound **1** was determined by the detection of the oxidative products with the 2',7'-dichlorofluorescein diacetate (DCFH) method reported previously<sup>6</sup>. Compound **1** shows antioxidant activity with an

IC<sub>50</sub> value of 2.78 mg/mL. This indicated that compound **1** is high antioxidant activity.

Since some of the flavonoids exhibited anti virus activities<sup>15,17,18</sup> compound **1** was tested for their potencies in preventing anti-tobacco mosaic virus activity using the half-leaf method<sup>7</sup>.

The antiviral inhibition rates of the compound at the concentration of 20 μM tested by the half-leaf method. The results showed that the compound **1** exhibited inhibition activities against tobacco mosaic virus replication with inhibition rate of 85.7 %. This rate is higher than that of the positive control, ningnanmycin (34.8%).

*N. glutinosa* were pretreated with solutions of compounds or a solution of DMSO for 6 h before inoculation with tobacco mosaic virus. At the concentration of 20 μM, compound **1** showed potent protective effects to the host plants, with the inhibition rates ranging from 80.4-88.9 %. The results indicated that pretreatment with the compound **1** can increase the resistance of the host plant to tobacco mosaic virus infection.

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