

Design, Synthesis of Some New Scaffolds based on Pyrrolyl-Pyridines as Potential Anticancer Agents

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Received: 18 June 2022;	Accepted: 6 January 2023;	Published online: 30 January 2023;	AJC-21132
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Pyrrolyl-pyridine heterocyclic compounds have received considerable attention because of its unique bioisosteric properties and an unusually wide spectrum of biological activities. Thus, it is a perfect framework for the synthesis of novel C–N, C–C bond formations like 5-substituted-1-benzyl-1*H*-pyrrolo[2,3-*b*]pyridines (**3**), 2-(1*H*-pyrrolo[2,3-*b*]pyridin-5-yl)phenol (**5**) and screened for their anticancer activity. The synthesized compounds were characterized by ¹H NMR, ¹³C NMR, IR, mass spectral techniques and elemental analysis. The outcomes of these compounds 2,6-difluorobenzyl-pyrrolidin-1*H*-pyrrolo[2,3-*b*]pyridine, 2,6-difluorobenzyl-*N*,*N*-dimethyl-1*H*-pyrrolo[2,3-*b*]pyridin-5-amine had a signicant anticancer activity against human cervical cancer cell line (Hela) with IC₅₀ 4.8, 9.7 µg/mL and whereas pyrrolo[2,3-*b*]pyridin-5-phenol, pyrrolo[2,3-*b*]pyridin-5-vinylphenol are active against human breast cancer cell line (MCF-7) with IC₅₀ 8.1, 3.2 µg/mL, respectively.

Keywords: Azaindoles, Anticancer activity, C-C formation, C-N formation, Docking studies.

INTRODUCTION

Approximately 9 million deaths worldwide each year are attributed to cancer, which is defined as the uncontrolled, rapid, and abnormal growth of aberrant cells [1,2]. Despite significant advances in the discovery of anticancer medications, there are still numerous challenges to cancer treatment, such as high toxicity, low efficacy and drug resistance, all of which have had a significant impact on patients' normal lives [3]. As a result, current cancer research is focused on finding strong, safe anticancer agents with great selectivity [4]. The indole skeleton is one of the most attractive anti-tumor structures and is widely seen in active molecules and natural chemicals such as koumine, vinblastine, vincristine and topsentine [5].

Azaindoles, which have a pyrrole ring fused to a pyridine ring, are frequently employed as bioisoteres of indoles to increase anticancer drug discovery activities. The azaindole core, rather than other bicyclic fused heterocycles, can be employed to modify and finely adjust Lipinski's rule of five, solubility, pK_a and lipophilicity, target binding and ADME-toxic characteristics [6-9]. Azaindoles have been identified as favoured structures in the control of biological processes, medicinal chemistry and drug discovery programmes [10-12].

Improvements in C-C bond and C-heteroatom (C-N, C-O and C-P) condensed and coupling reactions could be a hot topic in modern organic and medicinal chemistry [13]. Agrochemicals, prescription medications and compounds of interest in material sciences are all synthesized via these processes. The pyrrolo-pyridine nucleus is an important class of organic chemistry that encompasses a number of pharmacologically active chemicals that can be generated in the lab or obtained naturally [14]. Pyridine ring is present in many natural products including vitamins such as niacins and vitamin B₆, alkaloids e.g. trigonelline and coenzymes e.g. nicotinamide adenine dinucleotide [15]. The pyrrolo-pryridines are present in the molecular structure of various biologically active compounds such as antalarmin, vemurafenib, peficitinib, pexidartinib, plexxikon, genentech famitinib, etc. [16]. They are displaying interesting biological profiles such as antitumor [17,18], antibacterial [19], antifungal [20], anti-TB [21], anti-inflammatory [22], antiproliferative [23], anti-Parkinson's [24], antiviral [25], muscarinic antagonist [26]. Considering this panorama, herein

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we described the optimization of the synthetic route to obtain 1*H*-pyrrolo[2,3-*b*]pyridin-5-yl derivatives under microwave assisted synthesis as well conventional technique and evaluation of anticancer activity.

EXPERIMENTAL

All the reactants, reagents and solvents were obtained from commercially available sources and were of analytical grade. IR (KBr) spectrum was recorded on a Perkin-Elmer 400 FT-IR spectrometer (v_{max} , cm⁻¹) or a Varian 670-IR in the frequency range of 4000-600 cm⁻¹. Melting points were determined by the open capillary method and are uncorrected. ¹H NMR was recorded by using DMSO- d_6 solvent in the ranging 300-400 MHz and ${}^{13}C$ NMR was recorded by using DMSO- d_6 & CDCl₃ solvent in the ranging 100-125 MHz and TMS as an internal standard. The mass spectrum was recorded by using VG micro mass 70-70H instrument. The purity of these compounds was checked by TLC on silica gel and using *n*-hexane and ethyl acetate solvents. A monowave 300 mass 24 (Anton par) microwave was used to carry out the microwave reactions in 30 mL microwave process vials with temperature control by the infrared detection temperature sensor.

General procedure for the synthesis of 1-substituted benzyl-1*H*-pyrrolo[2,3-*b*]pyridine (3a-e): Under nitrogen atmosphere, 5-chloro-1*H*-pyrrolo[2,3-*b*]pyridine (1) (2 mmol), corresponding amines (2 mmol), K₂CO₃ (0.5 mmol) were added to acetonitrile (20 mL) and the reaction mixture was heated to reflux for 8-10 h. The reaction was carefully monitored using TLC and quenched with ice at 0 °C and the reaction was then extracted using ethyl acetate as solvent. The organic layer was removed using water followed by brine solution and solvent evaporation, yielding the final azaindole derivatives (2). Then compound 2 (2 mmol), substituted halides (2.4 mmol) and K₂CO₃ (1.5 mmol) were added in DMF (10 mL) and then the reaction mixture was stirred at room temperature for 12-14 h. After completion of reaction, it was diluted with water and extracted with ethyl acetate (2 × 25 mL), the crude product 3 was purified by column chromatography petroleum ether:ethyl acetate (9:1).

General procedure for the synthesis of substituted 2-(1H-pyrrolo[2,3-b]pyridin-5-yl)phenol (5a-e): Compound 5-chloro-1*H*-pyrrolo[2,3-*b*]pyridine (1, 1 mmol) was reacted with 2-substituted phenol (1 mmol) using Pd(tetrakis)triphenyl phosphene (0.2 mmol) as catalyst in the presence of Cu(I) and K₃PO₄ in 1,4-dioxane (10 mL) under refluxing conditions in seal-tube for 10-12 h to get 5-(2-methoxyphenyl)-1H-pyrrolo-[2,3-b]pyridine (4). Key intermediate 4 (10 mmol), tribromoborane (5 mmol) was dissolved in dichloromethane (15 mL) was added to the magnetically stirred solution at -78 °C to room temperature for 8-9 h (Scheme-I). The progress of the reaction was monitored by TLC and after completion of the reaction, the mixture was poured into crushed ice and extracted with DCM (2×50 mL). Finally, 5-substituted azaindoles was purified by column chromatography using eluent petroleum ether:ethyl acetate (9:1).

Microwave irradiation: A mixture of substituted azaindoles (1 mmol) (2), aryl halides (1.2 mmol) were taken in 10 mL seamless pressure vial in a Microwave 300 operating system and heated at 100-120 °C for 15-20 min. After completion of reaction, it was diluted with water and extracted with ethyl acetate (2×25 mL), the crude product was purified by column chromatography by using petroleum ether:ethyl acetate (9:1).

4-(1-(2,6-Difluorobenzyl)-1*H***-pyrrolo[2,3-***b***]pyridin-5yl)morpholine (3a):** Pale brown solid, m.p.: 176-178 °C. ¹H NMR (400 MHz, CDCl₃) δ ppm: 3.55 (t, 4H, J = 7.3 Hz, morpholine CH₂), 3.72 (t, 4H, J = 7.3 Hz, morpholine CH₂), 4.76 (s, 2H, NCH₂), 6.18 (d, 1H, J = 7.0 Hz, ArH), 7.04 (s, 1H, ArH), 7.23 (d, 2H, J = 7.0 Hz, ArH), 7.50 (d, 1H, J = 7.0Hz, ArH), 7.85 (t, 1H, J = 7.5 Hz, ArH), 9.69 (s, 1H, pyridine-H). ¹³C NMR (100 MHz, CDCl₃) δ ppm: 42.60 (NCH₂), 49.70 (NCH₂), 61.79 (OCH₂), 113.10 (pyrrole C), 114.08 (ArC), 118.88 (ArC), 119.10 (ArC), 119.62 (ArC), 119.77 (pyrrole C), 127.05 (ArC), 127.14 (pyridine C), 131.37 (pyridine C), 133.75 (pyridine C), 143.01 (ArC), 168.55 (ArC). IR (KBr, ν_{max}, cm⁻¹): 3077.76 (=CH), 2918.48, 2847.19 (CH), 1564.24



Scheme-I: Designed strategy of azaindole derivatives

(C=C), 1443.22 (C=N), 1350.44 (C-N), 1105.27 (COC), 967.26 (CF). ESI (m/z) calcd. for $C_{18}H_{17}F_2N_3O [M + 1]^+$: 330.21, found: 330.14.

4-(1-(4-Methoxybenzyl)-1H-pyrrolo[2,3-b]pyridin-5yl)morpholine (3b): White crystalline solid, m.p.: 195-197 °C. ¹H NMR (400 MHz, CDCl₃) δ ppm: 3.53 (t, 4H, J = 7.3 Hz, morpholine CH₂), 3.71 (t, 4H, J = 7.3 Hz, morpholine CH₂), 3.74 (s, 3H, OCH₃), 4.61 (s, 2H, NCH₂), 7.53 (d, 2H, J = 7.1Hz, ArH), 7.64 d (2H, J = 7.2 Hz, ArH), 7.81 d (1H, J = 7.2 Hz, ArH), 7.92 (d, 1H, J = 7.1 Hz, ArH), 8.52 (s, 1H, ArH), 10.23 (s, 1H, pyridine-H). ¹³C NMR (100 MHz, CDCl₃) δ ppm: 42.63 (morpholine C), 49.77 (OCH₃), 52.23 (NCH₂), 63.86 (morpholine C), 114.33 (pyrrole C), 115.02 (ArC), 121.10 (ArC), 122.98 (pyridine C), 124.10 (pyridine C), 126.27 (pyrrole C), 128.67 (ArC), 131.95 (ArC), 132.96 (pyridine C), 137.72 (pyridine C), 144.98 (ArC), 157.67 (ArC). IR (KBr, v_{max}, cm⁻¹): 3092.94 (=CH), 2923.62, 2856.06 (CH), 1563.05 (C=C), 1419.96 (C=N), 1250.19 (C-N), 1162.78 (COC), 1105.60 (COC). ESI (m/z) calcd. for C₁₉H₂₁N₃O₂ [M + 1]⁺: 324.21, found: 324.06.

1-(2,6-Difluorobenzyl)-5-(pyrrolidin-1-yl)-1*H*-pyrrolo-[**2,3-***b*]pyridine (3c): Pale brown solid, m.p.: 174-176 °C. ¹H NMR (400 MHz, CDCl₃) δ ppm: 2.75 (t, 4H, *J* = 7.5 Hz, CH₂), 3.09 (t, 4H, *J* = 7.3 Hz, CH₂), 4.65 (s, 2H, NCH₂), 6.51 (d, 1H, *J* = 6.8 Hz, ArH), 7.08 (d, 1H, *J* = 7.5 Hz, ArH), 7.40 (d, 1H, *J* = 7.5 Hz, ArH), 7.68 (s, 1H, ArH), 7.75 (t, 1H, *J* = 7.0 Hz, ArH), 8.10 (d, 1H, *J* = 7.0 Hz, ArH), 8.70 (s, 1H, pyridine-H). ¹³C NMR (100 MHz, CDCl₃) δ ppm: 23.56 (pyrrolidine C), 50.31 (NCH₂), 61.67 (pyrrolidine C), 122.71 (pyrrole C), 115.81 (ArC), 118.40 (ArC), 121.60 (ArC), 122.71 (pyridine C), 124.49 (pyridine C), 125.30 (pyrrole C), 134.37 (ArC), 138.37 (pyridine C), 154.63 (pyridine C), 161.64 (ArC). IR (KBr, v_{max}, cm⁻¹): 3074.24 (=CH), 2978.43, 2844.92 (CH), 1535.80 (C=C), 1448.23 (C=N), 1312.82 (C-N), 966.68 (CF). ESI (*m/z*) calcd. for C₁₈H₁₇F₂N₃ [M + 1]⁺: 314.18, found: 314.21.

1-(4-Methoxybenzyl)-5-(pyrrolidin-1-yl)-1H-pyrrolo-[2,3-b]pyridine (3d): White crystalline solid, m.p.: 155-157 °C. ¹H NMR (400 MHz, CDCl₃) δ ppm: 2.61 (t, 4H, J = 7.5 Hz, CH_2), 3.01 (t, 4H, J = 7.3 Hz, CH_2), 3.46 (s, 3H, OCH_3), 4.74 $(s, 2H, NCH_2), 6.74 (d, 1H, J = 7.0 Hz, ArH), 7.16 (d, 2H, J =$ 7.2 Hz, ArH), 7.46 (d, 2H, J = 7.2 Hz, ArH), 7.73 (s, 1H, ArH), 7.75 (d, 2H, J = 7.2 Hz, ArH), 8.09 (d, 1H, J = 7.0 Hz, ArH), 8.75 (s, 1H, pyridine-H). ¹³C NMR (100 MHz, CDCl₃) δ ppm: 23.56 (pyrrolidine C), 50.96 (pyrrolidine C), 52.80 (OCH₃), 60.51 (NCH₂), 115.02 (pyrrole C), 119.94 (ArC), 120.07 (pyridine C), 121.59 (ArC), 122.11 (pyridine C), 125.96 (ArC), 125.04 (ArC), 135.17 (pyridine C), 139.29 (pyridine C), 156.83 (pyridine C), 162.47 (ArC). IR (KBr, v_{max} , cm⁻¹): 3052.34 (=CH), 2961.10, 2897.49 (CH), 1571.08 (C=C), 1438.46 (C=N), 1318.31 (C-N), 957.69 (CF). ESI (*m/z*) calcd. for C₁₉H₂₁N₃O $[M + 1]^+$: 308.16, found: 308.09.

1-(2,6-Difluorobenzyl)-*N*,*N*-dimethyl-1*H*-pyrrolo[2,3-*b*]pyridin-5-amine (3e): Pale brown solid, m.p.: 130-132 °C. ¹H NMR (400 MHz, CDCl₃) δ ppm: 2.78 (s, 6H, 2 × CH₃), 4.61 (s, 2H, CH₂), 6.77 (d, 1H, *J* = 7.2 Hz, ArH), 7.47 (s, 1H, ArH), 7.59 (m, 2H, ArH), 7.72 (s, 1H, ArH), 8.04 (d, 1H, *J* = 7.2 Hz, ArH), 8.49 (s, 1H, pyridine-H). ¹³C NMR (100 MHz, CDCl₃) δ ppm: 43.45 (NCH₃), 50.82 (NCH₂), 114.70 (pyrrole C), 115.13 (ArC), 120.19 (ArC), 123.35 (pyridine C), 125.30 (ArC), 125.37 (pyrrole C), 127.37 (pyridine C), 130.18 (ArC), 136.05 (pyridine C), 138.94 (pyridine C), 148.03 (pyridine C), 162.40 (ArC). IR (KBr, v_{max} , cm⁻¹): 3045.41 (=CH), 2964.70 (CH), 1555.37 (C=C), 1430.18 (C=N), 1267.34 (C-N), 972.63 (CF). ESI (*m/z*) calcd. for C₁₆H₁₅F₂N₃ [M + 1]⁺: 288.17, found: 288.07.

5-Methyl-2-(1*H***-pyrrolo[2,3-***b***]pyridin-5-yl)phenol (5a):** White crystalline solid, m.p.: 110-112 °C. ¹H NMR (400 MHz, CDCl₃) δ ppm: 2.73 (s, 3H, CH₃), 5.64 (s, 1H, OH), 6.65 (d, 1H, *J* = 7.0 Hz, ArH), 7.34 (s, 1H, ArH), 7.60 (d, 1H, *J* = 7.3 Hz, ArH), 7.70 (s, 1H, ArH), 8.06 (d, 1H, *J* = 7.3 Hz, ArH), 8.17 (d, 1H, *J* = 7.0 Hz, ArH), 8.40 (s, 1H, pyridine-H). ¹³C NMR (100 MHz, CDCl₃) δ ppm: 23.04 (ArCH₃), 110.43 (pyrrole C), 114.27 (ArC), 120.15 (pyridine C), 122.31 (pyridine C), 124.36 (ArC), 125.45 (ArC), 126.38 (pyrrole C), 130.45 (pyridine C), 135.05 (ArC), 138.30 (ArC), 149.76 (pyridine C), 160.20 (ArC). IR (KBr, v_{max}, cm⁻¹): 3420.15 (OH), 3228.35 (NH), 3046.22 (=CH), 2925.17 (CH), 1544.15 (C=C), 1432.79 (C=N). ESI (*m*/*z*) calcd. for C₁₄H₁₂N₂O [M + 1]⁺: 225.18, found: 225.10.

5-Ethyl-2-(1*H***-pyrrolo[2,3-***b***]pyridin-5-yl)phenol (5b):** White crystalline solid, m.p.: 115-117 °C. ¹H NMR (400 MHz, CDCl₃) δ ppm: 2.73 (t, 3H, *J* = 7.8 Hz, CH₃), 3.10 (q, 2H, *J* = 9.2 Hz, CH₂), 5.10 (s, 2H, CH₂), 6.60 (d, 1H, *J* = 7.0 Hz, ArH), 7.30 (s, 1H, ArH), 7.61 (d, 1H, *J* = 7.2 Hz, ArH), 7.68 (s, 1H, ArH), 8.04 (d, 1H, *J* = 7.2 Hz, ArH), 8.14 (d, 1H, *J* = 7.0 Hz, ArH), 8.41 (s, 1H, pyridine-H). ¹³C NMR (100 MHz, CDCl₃) δ ppm: 23.04 (CH₃), 27.31 (CH₂), 111.34 (pyrrole C), 114.10 (pyridine C), 120.16 (pyridine C), 122.25 (ArC), 124.37 (ArC), 125.49 (ArC), 126.17 (pyrrole C), 131.33 (ArC), 134.22 (pyridine C), 137.35 (pyridine C), 148.35 (ArC), 160.88 (ArC). IR (KBr, v_{max} , cm⁻¹): 3410.21 (OH), 3298.16 (NH), 3055.21 (=CH), 2925.30 (CH), 1605.01, 1536.30 (C=C), 1398.22 (C=N). ESI (*m/z*) calcd. for C₁₅H₁₄N₂O [M + 1]⁺: 239.15, found: 239.19.

2-(1H-Pyrrolo[2,3-*b***]pyridin-5-yl)-5-vinylphenol (5c):** Light yellow solid, m.p.: 120-122 °C. ¹H NMR (400 MHz, CDCl₃) δ ppm: 5.09 (d, 2H, *J* = 7.5 Hz, CH₂), 5.84 (d, 1H, *J* = 7.5 Hz, CH), 6.71 (s, 1H, OH), 7.24 (d, 3H, *J* = 6.5 Hz, ArH), 8.01 (d, 2H, *J* = 8.6 Hz, ArH), 8.32 (s, 1H, ArH), 9.06 (s, 1H, pyridine-H), 10.04 (brs, 1H, NH). ¹³C NMR (100 MHz, CDCl₃) δ ppm: 114.70 (pyrrole C), 115.13 (=CH₂), 120.17 (ArC), 120.29 (pyridine C), 121.16 (ArC), 122.15 (ArC), 125.94 (pyrrole C), 126.03 (ArC), 129.52 (ArC), 138.07 (=CH), 145.04 (pyridine C), 146.77 (pyridine C), 155.93 (pyridine C), 160.43 (ArC). IR (KBr, v_{max}, cm⁻¹): 3383.50 (OH), 3218.24 (NH), 3043.18 (=CH), 2923.54 (CH), 1507.18 (C=C), 1348.57 (C=N). ESI (*m/z*) calcd. for C₁₅H₁₂N₂O [M + 1]⁺: 237.16, found: 237.09.

2-(1*H***-Pyrrolo[2,3-***b***]pyridin-5-yl)phenol (5d):** Pale brown solid, m.p.: 140-142 °C. ¹H NMR (400 MHz, CDCl₃) δ ppm: 5.38 (s, 1H, OH), 6.44 (d, 1H, *J* = 7.0 Hz, ArH), 6.70 (d, 1H, *J* = 7.2 Hz, ArH), 7.22 (dd, 1H, *J* = 7.5, 3.2 Hz, ArH), 7.56 (dd, 1H, *J* = 7.8, 3.0 Hz, ArH), 8.01 (d, 2H, *J* = 8.0 Hz, ArH), 8.25 (s, 1H, ArH), 9.01 (s, 1H, pyridine-H), 9.69 (brs, 1H, NH). ¹³C NMR (100 MHz, CDCl₃) δ ppm: 114.37 (pyrrole C), 120.44 (ArC), 121.64 (pyridine C), 122.20 (pyridine C), 123.44 (ArC), 125.99 (pyrrole C), 126.25 (ArC), 129.33 (ArC), 137.10 (ArC), 144.37 (pyridine C), 154.66 (pyridine C), 160.28 (ArC). IR (KBr, v_{max}, cm⁻¹): 3408.40 (OH), 3326.20 (NH), 2934.24 (CH),

1545.05 (C=C), 1368.28 (C=N). ESI (m/z) calcd. for C₁₃H₁₀N₂O [M + 1]⁺: 211.12, found: 211.04.

4-(1*H***-Pyrrolo[2,3-***b***]pyridin-5-yl)benzene-1,3-diol (5e):** Pale brown solid, m.p.: 133-135 °C. ¹H NMR (400 MHz, CDCl₃) δ ppm: 5.32 (s, 1H, OH), 6.25 (d, 1H, *J* = 6.8 Hz, ArH), 6.68 (s, 1H, ArH), 7.12 (d, 1H, *J* = 6.8 Hz, ArH), 7.42 (d, 1H, *J* = 6.5 Hz, ArH), 7.42 (d, 1H, *J* = 6.5 Hz, ArH), 7.57 (d, 1H, *J* = 6.5 Hz, ArH), 8.03 (s, 1H, ArH), 8.68 (s, 1H, ArH), 9.72 (s, 1H, pyridine-H). ¹³C NMR (100 MHz, CDCl₃) δ ppm: 115.72 (pyrrole C), 119.33 (ArC), 120.14 (ArC), 121.30 (pyridine C), 122.16 (pyridine C), 125.95 (pyrrole C), 128.15 (ArC), 130.40 (ArC), 137.86 (ArC), 146.75 (pyridine C), 148.47 (pyridine C), 160.39 (ArC), 162.43 (ArC). IR (KBr, v_{max}, cm⁻¹): 3420.14 (OH), 2346.11 (NH), 2925.43 (CH), 1605.21 (C=C), 1376.17 (C=N). ESI (*m/z*) calcd. for C₁₃H₁₀N₂O₂ [M + 1]⁺: 227.24, found 227.12.

Anticancer activity: The anticancer activity of the final compounds was determined using the ATCC procedure [27] and the MTT colorimetric assay. The most common human breast cancer cell line (MCF-7) as well as cervical cancer cell line (Hela), was obtained from the National Centre for Cell Science, Pune, India. The P388 leukaemia cancer cell line was grown in DMEM media with 10% neonatal calf serum (NBCS), 1% non-essential amino acids, 1% sodium pyruvate, 0.2% NaHCO₃ and 1% antibiotic combination (10,000 U penicillin and 10 mg streptomycin per mL). The MCF-7 breast cancer cell line was grown in RPMI-1640 media with 10% NBCS, 100 IU/mL penicillin, 100 mg/mL streptomycin and 2 mM glutamine. Cell lines were kept at 37 °C in a humidified 5% CO₂ incubator and processed by first trypsinizing adherent cells and then centrifuging to obtain cell pellets. Fresh media was added to pellets for cell counting on a haemocyto-meter and 100 µL of media was dispensed in a 96-well plate with cells ranging from 5,000 to 6,000 per well. The plate was placed in a CO₂ incubator overnight to allow the cells to adhere and restore their shape. Cells were treated with 25 μ M solutions of the test compounds in the medium after 24 h. The cells were cultured for 48 h to see if the test chemicals had any effect on the cell lines. To deduct even more from 48 h data, a zero-hour reading with untreated cells and a control with 1% DMSO were taken. The cells were treated with MTT 3-(4,5-dimethylthiazol-2yl)-2,5-diphenyltetrazolium bromide diluted in PBS (5 mg/ mL) and incubated for 3-4 h at 37 °C after a 48 h incubation period. The formazan crystals were then dissolved in 100 µL of DMSO and the viability was determined using a Spectra Max multimode reader at 540 nm.

RESULTS AND DISCUSSION

A reaction between 5-chloro-1*H*-pyrrolo[2,3-*b*]pyridine (1), corresponding amines and K_2CO_3 in acetonitrile were heated for 8-10 h to give 5-(pyrrolidine/morpholine)-1*H*-pyrrolo[2,3-*b*]-pyridines (2) in good yields. Further to obtain compound 2, substituted halides were added in presence of K_2CO_3 in DMF at room temperature for 12-14 h in excellent yields and the derived analogues **3** was purified by column chromatography using petroleum ether:ethyl acetate (9:1) as solvent. Whereas another series of azaindoles have been synthesized by using

the reaction between 5-chloro-1*H*-pyrrolo[2,3-*b*]pyridine (1) with substituted phenol in the presence of Pd(PPh₃)₄, Cu(I) and K_3PO_4 in 1,4-dioxane under refluxing conditions for 10-12 h to get 5-(2-methoxyphenyl)-1*H*-pyrrolo[2,3-*b*]pyridine (4). Moreover, key intermediate 4 was treated with tribromoborane in dichloromethane as a solvent under room temperature for 8-9 h to desired final 5-substituted azaindoles **5** (Scheme-I).

The microwave irradiation technique has been offering a standard shift in the target synthesis due to their peculiar direct 'in core' heating of the reactions thus decreasing the reaction times [28]. Moreover, it offered myriad advantages such as acceleration of sluggish transformation, higher yields and environmentally greener conditions. Under the non classical reaction rates are faster than classical heating conditions, for example, the synthesis of azaindoles (5) were obtained in 12 h by conventional method whereas, under microwave conditions, it was acquired in 15 min. Reaction between 5-substituted azaindoles 2 and aromatic halides, K₂CO₃ in DMF, whereas another series of substituted azaindoles with BBr₃ in DCM were taken in a closed vessel and irradiated under the microwave at 70-120 °C for 15-20 min. All the reactions were performed in the presence of 80-90 watts of microwave irradiation.

Moreover, we exploited the developed methodology to synthesize a novel series of simplied heterocyclic analogues designed by changing the azaindoles attached substituted rings in order to investigate the stereo-electronic and the lipophilic inuences at this position on the sedative role of pyrrolo-pyridine scaffolds. On the other hand, the exploitation of successive molecular simplications on the structure of sedative prototype (3a) led us to the design of the morpholine azaindole 2a and 2,6-difluorobenzene as shown in Table-1.

TABLE-1 OPTIMIZATION OF REACTION CONDITIONS FOR THE SYNTHESIS OF AZAINDOLE DERIVATIVE 3a							
Entry	Solvent	Base	Mol (%)	Temp./time	Yield (%) ^a		
1	ACN	K_2CO_3	1	RT/11 h	66		
2	t-BuOH	K_2CO_3	1	RT/12 h	72		
3	DMSO	K_2CO_3	1	RT/11 h	67		
4	EtOH	K_2CO_3	1	RT/12 h	76		
5	DMF	K_2CO_3	1	RT/12 h	84		
6	-	K_2CO_3	1	70 °C/15 min	70 ^b		
7	-	K_2CO_3	1	80 °C/15 min	74 ^b		
8	-	K_2CO_3	1	90 °C/15 min	82 ^b		
9	-	K_2CO_3	1	100 °C/15 min	95 ^b		
10	-	K_2CO_3	1	120 °C/15 min	89 ^b		

^aIsolated yield, ^bUnder microwave

However, adding 1 mol% K_2CO_3 to the reaction under room temperature conditions in acetonitrile after 11 h yielded the desired product in 66% yield (Table-1, entry 1). Another model reaction produced the required product **3a** in 72% yields, under 1 mol% K_2CO_3 , *tert*.-butanol at ambient temperature (Table-1, entry 2). In order to reduce reaction time, an attempt was made to implement the framework reaction conditions in DMSO at room temperature, which resulted in a 67% decrease in product yield (Table-1, entry 3). However, further increment

TABLE-2 ANTICANCER ACTIVITY RESULTS (μ g/mL) OF THE SYNTHESIZED PYRROLYL-PYRIDINES							
Entry	Products	Hela (IC ₅₀)	MCF-7 (IC ₅₀)	Convention time/yield	Microwave time/yield		
3a		14.6	3.8	12 h/84%	15 min/95%		
3b		ND	23.1	12 h/86%	15 min/97%		
3c		4.8	10.7	12.5 h/75%	17 min/86%		
3d		19.2	5.0	11.5 h/70%	15 min/81%		
3e		9.7	18.3	11 h/65%	15 min/74%		
5a	OH N H	3.8	8.1	9.5 h/80%	17 min/89%		
5b	OH N H	6.9	ND	9 h/82%	20 min/92%		
5c	OH N H	8.0	3.2	10 h/82%	17 min/94%		
5d		ND	24.8	9.5 h/70%	16 min/81%		
5e		15.7	ND	9 h/75%	15 min/93%		
DXN	-	5.0	5.0	-	-		

ND = $IC_{50} > 50 \mu g/mL$.

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of time from 11 h to 12 h led to increasing in yields (76%) in the presence of 1 mol% K₂CO₃, ethanol employing as a solvent (Table-1, entry 4). Despite the superior yield, product 3a was synthesized by 1 mol% K₂CO₃ in DMF to obtained 84% yield (Table-1, entry 5). In order to minimize the reaction time, further an attempt to carryout out the model reaction in the microwave reactor using sealed vessels under fast conventional heating at 70 °C was successful, resulting in the desired product 3a in 70% yield of within 15 min (Table-1, entry 6). Further optimizing of the reaction conditions, it was found that under neat reaction the conditions at 80 °C the product yield improved up to 74% (Table-1, entry 7). Whereas the reaction was found in the reaction conditions at 100 °C, the desired compound give excellent yields up to 95% (Table-1, entry 9). However, further increment of temperature from 100 °C to 120 °C led to decrease in yield (Table-1, entries 10). Despite the good yield, product 3a was isolated without using column chromatography purification process as sole product. After completion of the reaction, water, ethyl acetate was added to the reaction mixture and the precipitated solid was filtered and washed with ethanol to give the pure 4-(1-(2,6-difluorobenzyl)-1H-pyrrolo[2,3-b]pyridin-5-yl)morpholine (3a).

Anticancer activity: The anticancer activity of the synthesized compounds was assessed in vitro by 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) assay against Hela and MCF-7 cell lines. The percentage of cell death was measured for the novel indole linked to oxadiazoles (3) and 5) exhibited higher anticancer activity against at various concentrations along with IC₅₀ values (Table-2). Compounds 3c, 3e, 5a and 5c showed excellent anticancer activity against both Hela, MCF-7 cell lines. From screening results, the synthesized compounds displayed promising activity, whereas 3c, 3e exhibited the highest cytotoxicity against Hela cell line with IC_{50} 4.8, 9.7 µg/mL and active against MCF-7 cell line with IC₅₀10.7, 18.3 µg/mL values respectively. Compounds **3a**, **3d** with difluorobenzyl indole morpholine, methoxybenzyl indole elicited good potent activity against MCF-7 cell line with IC₅₀ value 14.6, 19.2 µg/mL, respectively. Compounds 5a and 5c displayed excellent anticancer activity against MCF-7 cell line with IC₅₀ value of 8.1, $3.2 \,\mu$ g/mL where as these indoles revelas the superiar anticancer activity against Hela cell line along with IC₅₀ value of 3.8, 8.0 µg/mL, respectively. Morover, compounds 5b, 5d and 5e show the moderate to good anticancer activity against both the cell lines with IC₅₀ values range 6.9-24.8 µg/mL, respectively.

Conclusion

Carbon-carbon, carbon-heteroatom forms the 'spine' of the organic molecule and is essentially the critical transformation in organic synthesis to line up the carbon backbone of organic particles. Final azaindoles **3c**, **3e**, **5a** and **5c** displayed promising anticancer activity against Hela, MCF-7 cell lines and prepared scaffolds are most active against both the cell lines. These synthetic methodologies provide a controlled, modular and facile access to azaindole scaolds with high eciency, broad substrate scope and excellent functional group compatibility.

ACKNOWLEDGEMENTS

One of the authors, NM are thankful to GITAM Deemed to be University & Symphony Pharma Pvt. Ltd., Hyderabad, India for providing the laboratory facility to carry out this work.

CONFLICT OF INTEREST

The authors declare that there is no conflict of interests regarding the publication of this article.

REFERENCES

- R.L. Siegel, K.D. Miller, S.A. Fedewa, D.J. Ahnen, R.G. Meester, A. Barzi and A. Jemal, *CA Cancer J. Clin.*, 67, 177 (2017); <u>https://doi.org/10.3322/caac.21395</u>
- F. Bray, J. Ferlay, I. Soerjomataram, R.L. Siegel, L.A. Torre and A. Jemal, *CA Cancer J. Clin.*, 68, 394 (2018); https://doi.org/10.3322/caac.21492
- 3. J. Akhtar, A.A. Khan, Z. Ali, R. Haider and M. Shahar Yar, *Eur. J. Med. Chem.*, **125**, 143 (2017);
- https://doi.org/10.1016/j.ejmech.2016.09.023
 4. O.O. Fadeyi, S.T. Adamson, E.L. Myles and C.O. Okoro, *Bioorg. Med. Chem. Lett.*, 18, 4172 (2008);
- https://doi.org/10.1016/j.bmc1.2008.05.078 M Greenwell and PK S M Rahman Int J P
- M. Greenwell and P.K.S.M. Rahman, *Int. J. Pharm. Sci. Res.*, 6, 4103 (2015).
- 6. J.Y. Merour, F. Buron, K. Ple, P. Bonnet and S. Routier, *Molecules*, **19**, 19935 (2014);
- https://doi.org/10.3390/molecules191219935
- M. Prudhomme, Eur. J. Med. Chem., 38, 123 (2003); https://doi.org/10.1016/S0223-5234(03)00011-4
- H.C. Zhang, H. Ye, B.R. Conway, C.K. Derian, M.F. Addo, G.H. Kuo, L.R. Hecker, D.R. Croll, J. Li, L. Westover, J.Z. Xu, R. Look, K.T. Demarest, P. Andrade-Gordon, B.P. Damiano and B.E. Maryanoff, *Bioorg. Med. Chem. Lett.*, 14, 3245 (2004); https://doi.org/10.1016/j.bmcl.2004.03.090
- E.R. Grant, M.A. Errico, S.L. Emanuel, D. Benjamin, M.K. McMillian, S.A. Wadsworth, R.A. Zivin and Z. Zhong, *Biochem. Pharmacol.*, 62, 283 (2001);
 - https://doi.org/10.1016/S0006-2952(01)00665-7
- D.A. Horton, G.T. Bourne and M.L. Smythe, *Chem. Rev.*, **103**, 893 (2003); https://doi.org/10.1021/cr020033s
- 11. D.R. Motati, R. Amaradhi and T. Ganesh, Org. Chem. Front., 8, 466 (2021);
- https://doi.org/10.1039/D0Q001079K

 12.
 V. Sharma, P. Kumar and D. Pathak, J. Heterocycl. Chem., 47, 491 (2010);
- <u>https://doi.org/10.1002/jhet.349</u> 13. H.J. Schäfer, *C.R. Chim.*, **14**, 745 (2011);
- https://doi.org/10.1016/j.crci.2011.01.002 14. J.Y. Merour and B. Joseph, *Curr. Org. Chem.*, **5**, 471 (2001);
- https://doi.org/10.2174/1385272013375427
- A.A. Altaf, A. Shahzad, Z. Gul, A. Shahzad, Z. Gul, N. Rasool, A. Badshah, B. Lal and E. Khan, *J. Drug Design Med. Chem.*, 1, 1 (2015); https://doi.org/10.11648/j.jddmc.20150101.11
- K.T. Flaherty, I. Puzanov, K.B. Kim, A. Ribas, G.A. McArthur, J.A. Sosman, P.J. O'Dwyer, R.J. Lee, J.F. Grippo, K. Nolop and P.B. Chapman, N. Engl. J. Med., 363, 809 (2010); https://doi.org/10.1056/NEJMoa1002011
- R. Cincinelli, L. Musso, L. Merlini, G. Giannini, L. Vesci, F.M. Milazzo, N. Carenini, P. Perego, S. Penco, R. Artali, F. Zunino, C. Pisano and S. Dallavalle, *Bioorg. Med. Chem.*, 22, 1089 (2014); <u>https://doi.org/10.1016/j.bmc.2013.12.031</u>
- Sh. Ahmad, O. Alam, M.J. Naim, M. Shaquiquzzaman, M.M. Alam and M. Iqbal, *Eur. J. Med. Chem.*, **157**, 527 (2018); <u>https://doi.org/10.1016/j.ejmech.2018.08.002</u>
- S.D. Paget, C.M. Boggs, B.D. Foleno, R.M. Goldschmidt, D.J. Hlasta, M.A. Weidner-Wells, H.M. Werblood, K. Bush and M.J. Macielag, *Bioorg. Med. Chem. Lett.*, 16, 4537 (2006); <u>https://doi.org/10.1016/j.bmcl.2006.06.023</u>

- W.B. Han, A.H. Zhang, X.Z. Deng, X. Lei and R.X. Tan, Org. Lett., 18, 1816 (2016); <u>https://doi.org/10.1021/acs.orglett.6b00549</u>
- A.D. Khoje, C. Charnock, B. Wan, S. Franzblau and L.-L. Gundersen, Bioorg. Med. Chem., 19, 3483 (2011); https://doi.org/10.1016/j.bmc.2011.04.023
- S.S. Fatahala, S. Hasabelnaby, A. Goudah, G. Mahmoud and R.H. Abd El Hameed, *Molecules*, 22, 461 (2017); https://doi.org/10.3390/molecules22030461
- S. Narva, S. Chitti, B.R. Bala, M. Alvala, N. Jain and V.G.C.S. Kondapalli, *Eur. J. Med. Chem.*, **114**, 220 (2016); https://doi.org/10.1016/j.ejmech.2016.02.059
- V.S. Goodfellow, C.J. Loweth, S.B. Ravula, T. Wiemann, T. Nguyen, Y. Xu, D.E. Todd, D. Sheppard, S. Pollack, O. Polesskaya, D.F. Marker, S. Dewhurst and H.A. Gelbard, *J. Med. Chem.*, 56, 8032 (2013); <u>https://doi.org/10.1021/jm401094t</u>

- S. Hong, S. Lee, B. Kim, H. Lee, S.-S. Hong and S. Hong, *Bioorg. Med. Chem. Lett.*, **20**, 7212 (2010); https://doi.org/10.1016/j.bmcl.2010.10.108
- 26. B. Blass, ACS Med. Chem. Lett., 6, 726 (2015); https://doi.org/10.1021/acsmedchemlett.5b00185
- M.F. Abu Bakar, M. Mohamad, A. Rahmat, S.A. Burr and J.R. Fry, *Food Chem. Toxicol.*, 48, 1688 (2010); <u>https://doi.org/10.1016/j.fct.2010.03.046</u>
- Y.H. Zhao, M.H. Abraham, J. Le, A. Hersey, C.N. Luscombe, G. Beck, B. Sherborne and I. Cooper, *Pharm. Res.*, **19**, 1446 (2002); <u>https://doi.org/10.1023/A:1020444330011</u>