



NOTE

A New Antiviral Phenolic Compounds from *Arundina grammifolia*

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Received: 29 August 2014;

Accepted: 30 January 2015;

Published online: 26 May 2015;

AJC-17292

A new phenolic compound, gramniphinol I (**1**), was isolated from the whole plant of *Arundina grammifolia*. Its structure was determined by means of HRESIMS, extensive 1D and 2D NMR spectroscopic studies and chemical evidence. Compound **1** was also tested for its antitobacco mosaic virus (anti-TMV) activity and it exhibited notable antitobacco mosaic virus activity with inhibition rate of 16.8 %.

Keywords: *Arundina grammifolia*, Phenolic compound, Antitobacco mosaic virus activity.

Arundina grammifolia (D. Don) Hochr. (bamboo orchid) belongs to the orchid family (Orchidaceae). The plant is used in Chinese folkloric medicine as a detoxifying and diuretic agent, as well as for the treatment of arthritis and inflammation¹. Previous phytochemical studies on *A. grammifolia* has revealed the presence of stilbenoids²⁻⁴, sterols^{5,6}, triterpenes^{7,8} and other phenolic compounds⁹⁻¹¹. Continuing the efforts to discover bioactive metabolites from local plants, we now investigated the chemical constituents of the whole plant of *A. grammifolia* growing in the Xishuangbanna Prefecture, leading to the isolation of a new phenolic compound (**1**). Its structure was determined by means of spectroscopic methods including 1D and 2D NMR techniques. Compound **1** was also tested for its antitobacco mosaic virus (anti-TMV) activity.

General procedures: UV spectra were obtained on a Shimadzu UV-2401A spectrophotometer; and CD spectra were measured on a JASCO J-810 spectropolarimeter. Tenor 27 spectrophotometer was used for scanning IR spectra (KBr pellets). 1D and 2D NMR spectra were recorded on a DRX-500 spectrometer with TMS as internal standard. Chemical shifts (δ) were expressed in ppm with reference to TMS. HRESIMS was performed on an API QSTAR spectrometer or a VG Autospec-3000 spectrometer. Preparative HPLC was performed on a Shimadzu LC-8A liquid chromatography equipped with ZORBAX PrepHT GF (21.2 mm \times 25 cm, 7 μ m) column or Venusil MP C₁₈ (20 mm \times 25 cm, 5 μ m) column. Column chromatography was performed using Si gel (200-300 mesh, Qing-dao Marine Chemical, Inc., Qingdao, China),

Lichroprep RP-18 gel (40-63 μ m, Merck, Darmstadt, Germany), Sephadex LH-20 (Sigma-Aldrich, Inc, USA), or MCI gel (75-150 μ m, Mitsubishi Chemical Corporation, Tokyo, Japan). Column fractions were monitored by TLC and the spots were visualized by heating the plates after spraying with 5 % H₂SO₄ in EtOH.

The whole plant of *Arundina grammifolia* (D. Don) Hochr. was collected in the Xishuangbanna prefecture of Yunnan Province in in September 2012. The identification of the plant material was verified by Prof. Wu SG (Xishuangbanna Botanical Garden). A voucher specimen (YMU-2012-9-18) has been deposited in our laboratory.

Extraction and isolation: The air-dried and powdered whole plant of *A. grammifolia* (2.0 kg) was extracted three times with 70 % aqueous acetone (3 \times 3.5 L) at room temperature and filtered to yield a filtrate. The filtrate was concentrated and partitioned between H₂O and EtOAc. The EtOAc fraction was dried under reduced pressure and then submitted. The extract was applied to silica gel (150-200 mesh) column chromatography, eluting with CHCl₃-MeOH gradients (20:1, 9:1, 8:2, 7:3, 6:4, 5:5) to afford fractions A-F. Further separation of fraction B (21.2 g) by silica gel (300-400 mesh), eluted with CHCl₃-CO(CH₃)₂ (9:1-1:2), yielded fractions B1-B7. Fraction B3 (2.62 g), upon further separation on silica gel using petroleum ether-CH₃COOC₂H₅ and semi-preparative HPLC (46 % MeOH-H₂O, flow rate 12 mL/min), afforded compound **1** (15.4 mg).

Gramniphinol I (1): C₁₇H₁₆O₅, yellow gum; UV (CH₃OH) λ_{\max} (log ϵ) 210 (3.92), 275 (3.26), 314 (3.52) nm; IR (KBr,

ν_{\max} , cm^{-1}): 3448, 1620, 1497, 1469, 1385, 1145, 878, 820; ^{13}C NMR and ^1H NMR data (DMSO- d_6 , 500 and 125 MHz) (Table-1); positive ESIMS m/z 323 $[\text{M}+\text{Na}]^+$; negative HRESIMS m/z 323.0888 $[\text{M}+\text{Na}]^+$ (calcd for $\text{C}_{17}\text{H}_{16}\text{O}_5$, 323.0895).

TABLE-1
 ^1H NMR AND ^{13}C NMR DATA OF
COMPOUND **1** (δ ppm, IN DMSO- d_6)

No.	δ_{C}	δ_{H} (J in Hz)	No.	δ_{C}	δ_{H} (J in Hz)
2	148.2 s		1'	110.3 s	
3	113.5 s		2'	156.9 s	
4	119.8 d	7.42 d (8.2)	3'	103.4 d	6.40 d (1.8)
5	108.2 d	6.79 dd (1.8, 8.2)	4'	158.6 s	
6	158.9 s	6.91 s	5'	105.8 d	6.31 dd (1.8, 8.2)
7	98.2 d	7.08 d (1.8)	6'	128.9 d	7.15 dd (8.2)
8	155.1 s		6-OMe	55.8 q	3.82 s
9	122.6 s		2'-OMe	56.2 q	3.79 s
10	58.4 t	4.68 s	Ar-OH		10.02 s

Powdered whole plants of *A. grammifolia* were extracted with 70 % aqueous acetone. The filtrate was concentrated and partitioned between H_2O and EtOAc. The EtOAc fraction was dried under reduced pressure and then submitted to silica gel, MCI, RP-18 gel column chromatography (CC) and semi-preparative HPLC to yield compound **1**. The structure of compound **1** was shown in Fig. 1 and the ^1H NMR and ^{13}C NMR spectroscopic data of compound **1** are listed in Table-1.

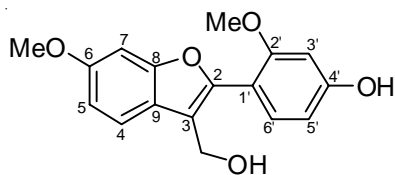


Fig. 1. Structure of compound **1**

Gramniphénol I (**1**) was obtained as yellow gum. It possessed the molecular formula $\text{C}_{17}\text{H}_{16}\text{O}_5$, as revealed by its HR-ESI-MS spectrum (m/z 323.0888, $[\text{M}+\text{Na}]^+$, calcd 323.0895). The IR absorptions at 3448 (OH) and 1620 (aromatic) cm^{-1} and UV absorptions at 314 and 275 nm were characteristics for 3-methylene benzofurans. On the basis of ^1H NMR and ^{13}C NMR spectral analysis (Table-1), compound **1** had a 14-carbon skeleton with one oxidated methylene group (δ_{H} 4.68/ δ_{C} 58.4), two methoxy group (δ_{H} 3.82/ δ_{C} 55.8 and δ_{H} 3.79/ δ_{C} 56.2) and one phenolic hydroxy group (δ_{H} 10.02). Six olefinic methane carbons (δ_{C} 98.2, 103.4, 105.8, 108.2, 119.8 and 128.9) and eight quaternary carbons [five oxygenated sp^2 carbons (δ_{H} 148.2, 158.9, 155.1, 156.9, 158.6)] were observed according to HSQC and DEPT spectra. In the ^1H NMR spectrum, two AMX systems (δ_{H} 7.08, d, $J = 1.8$ Hz; 7.42, d, $J = 8.2$ Hz; 6.79, dd, $J = 8.2, 1.8$ Hz and δ_{H} 6.40, d, $J = 1.8$ Hz; 7.15, d, $J = 8.2$ Hz; 6.31, dd, $J = 8.2, 1.8$ Hz) were observed. In the HMBC spectrum (Fig. 2), the $-\text{CH}_2\text{OH}$ protons at δ_{H} 4.68 were correlated with C-2 (148.2), C-3 (113.5) and C-9 (122.6), indicating that the $-\text{CH}_2\text{OH}$ was connected to C-3. The correlations

of the $-\text{OCH}_3$ protons at δ_{H} 3.82 and 3.79 with C-6 (158.9) and C-22 (156.9) showed that the $-\text{OCH}_3$ groups should be attached to C-6 and C-22, respectively. The $-\text{OH}$ proton (δ_{H} 10.02) showed HMBC correlations with C-32 (103.4), C-42 (158.6) and C-52 (105.8), indicating that it was attached to C-42. Consequently, compound **1** was identified as shown.

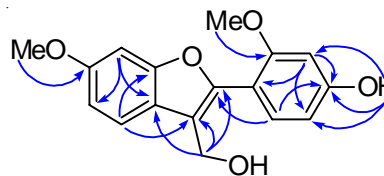


Fig. 2. Key HMBC (\curvearrowright) correlations of compound **1**

Since certain of the phenolic compounds exhibit potential antitobacco mosaic virus activity.¹²⁻¹⁴ Compound **1** was tested for its antitobacco mosaic virus activities. The inhibitory activities of compound **1** against tobacco mosaic virus replication were tested using the half-leaf method.¹⁵ Ningnanmycin, a commercial product for plant disease in China, was used as a positive control. The results showed that compound **1** exhibited antitobacco mosaic virus activity with inhibition rate of 16.8 %.

ACKNOWLEDGEMENTS

This research was supported by the Key Laboratory of Pharmacology for Natural Products (Kunming Medical University) (2014G005).

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