

NOTE

A New Biphenyl from the Fruits of Schisandra lancifolia and Its Cytotoxicity

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A new biphenyl *i.e.*, lancibiphenyl A (1) was isolated from the fruits of *Schisandra lancifolia*. The structure of 1 was elucidated by spectroscopic methods including extensive 1D- and 2D-NMR techniques. Compound 1 was evaluated for its cytotoxicity and it exhibited moderate cytotoxicity against A549 and PC3 cell with IC_{50} values of 6.4 and 5.7 μ M, respectively.

Keywords: Biphenyl, Schisandra lancifolia, Cytotoxicity.

The fruits of Schisandra plants are used as herb medicine as the treat of sedative and tonic agents^{1,2}. Previous studies³⁻⁵ have shown that the plants of this genus are rich in lignans and triperpenoids, which have been found to possess some beneficial activities. Schisandra lancifolia belongs to the genus Schisandra of the family Schisandraceae. It is a climbing plant mainly distributed in Yunnan, Sichuan and Shanxi Province in mainland China⁶. In previous studies, some new dibenzocyclooctadiene lignans were isolated from the fruits of S. lancifolia from of Erlang Mountain area of Sichuan Province⁷. In our continuing efforts to identify bioactive natural products from the medicinal plants of the Schisandraceae, a chemical investigation on the fruits of S. lancifolia from Lijiang Country, Yunnan Province was carried out. As a result, a new biphenyl (1) was separated from this plant and the cytotoxicity of 1 was evaluated. In this paper, the structure elucidation and biological activities of the compound are discussed.

General procedures: Optical rotations were measured in a Horiba SEPA-300 polarimeter. UV spectra were obtained on a Shimadzu UV-2401A spectrophotometer and CD spectra were measured on a JASCO J-810 spectropolarimeter. A Tenor 27 spectrophotometer was used for scanning IR spectra (KBr pellets). 1D- and 2D- NMR spectra were recorded on a DRX-500 spectrometer with TMS as internal standard. Chemical shifts (δ) are expressed in ppm with reference to TMS. HRESIMS was performed on an API QSTAR spectrometer or a VG Autospec-3000 spectrometer. Preparative HPLC was performed on a Shimadzu LC-8A liquid chromatograph equipped with Zorbax PrepHT GF (21.2 mm × 25 cm, 7 µm) column or Venusil MP C₁₈ (20 mm × 25 cm, 5 µm) column. Column chromatography was performed using Si gel (200-300 mesh, Qing-dao Marine Chemical, Inc., Qingdao, People's Republic of China), Lichroprep RP-18 gel (40-63 μ m, Merck, Darmstadt, Germany), Sephadex LH-20 (Sigma-Aldrich Corp. -St Louis, USA), or MCI gel (75-150 μ m, Mitsubishi Chemical Corporation, Tokyo, Japan). Column fractions were monitored by TLC and the spots were visualized by heating the plates after spraying with 5 % H₂SO₄ in EtOH.

The fruits of *S. lancifolia* were collected in Lijiang Country of Yunnan Province, People's Republic of China, in September 2011. The identification of the plant material was verified by Prof. Xi-Wen Li of Kunming Institute of Botany, Chinese Academy of Sciences. A voucher specimen (KIB 11-9-58) has been deposited in our Laboratory.

Extraction and isolation: The air-dried and powdered fruits of *S. lancifolia* (2.2 kg) were extracted four times with 70 % (CH₃)₂CO (4 × 3 L) at room temperature and filtered, with the filtrate evaporated under the reduced pressure and partitioned with EtOAc (3 × 1 L). The EtOAc partition (152 g) was applied to silica gel (200-300 mesh) column chromatography, eluting with a CHCl₃-Me₂CO gradient system (20:1, 9:1, 8:2, 7:3, 6:4, 5:5), to give five fractions, A-E. The further separation of fraction D (7:3, 18.6 g) by silica gel column chromatography, eluted with petroleum ether-acetone and semi-preparative HPLC (40 % MeOH-H₂O, flow rate 12 mL/min) to give **1** (15.2 mg).

Lancibiphenyl A (1): White powder; UV (MeOH) λ_{max} (log ϵ): 210 (4.18), 282 (3.75), 332 (3.08) nm; IR (KBr, ν_{max} , cm⁻¹) 3418, 2914, 1718, 1605, 1538, 1425, 1386, 1325, 1259, 1168, 1074, 867, 748; ¹H and ¹³C NMR: see Table-1; ESIMS

TABLE-1 ¹ H NMR AND ¹³ C NMR DATA OF COMPOUND 1 (IN C ₅ ND ₅ ; 500 AND 125 MHz)							
No.	$\delta_{\rm C}({\rm m})$	$\delta_{\rm H}$ (m, J, Hz)	No.	$\delta_{\rm C}({\rm m})$	$\delta_{\rm H}$ (m, J, Hz)		
1	133.6 s	-	5'	116.7 d	6.97, d, <i>J</i> = 8.6		
2, 6	104.7 d	6.65 s	6'	134.6 d	6.97, dd, J = 8.6, 2.0		
3, 5	148.9 s	-	7'	168.2 s	-		
4	134.5 s	-	3-OMe	56.1 q	3.86 s		
1′	133.0 s	-	7'-OMe	52.2 q	4.12 s		
2'	135.6 d	7.88, d, <i>J</i> = 2.2	4-OH	11.8 s	-		
3'	118.3 s	-	4'-OH	12.2 s	-		
4′	155.6 s	-		-	-		

(positive ion mode): m/z 327 [M + Na]⁺; HRESIMS (positive ion mode): m/z 327.0839 [M + Na]⁺ (calcd C₁₆H₁₆NaO₆ for 327.0845).

The fruits of *S. lancifolia* were extracted with 70 % acetone. The extract produced was subjected repeatedly to column chromatography on silica gel, Sephadex LH-20, RP-18 and RP-HPLC, to afford the new biphenyl, lancibiphenyl A. Its structure was shown in Fig. 1 and its ¹H and ¹³C NMR spectroscopic data were listed in Table-1.

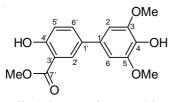


Fig. 1. Structure of compound 1

Compound 1 was obtained as yellow gum and was assigned the molecular formula of $C_{16}H_{16}O_6$, by HRESIMS at m/z 327.0839 $[M + Na]^+$ (calcd *m/z* 327.0845). The ¹H and ¹³C NMR spectrum showed signals characteristic of one 1,3,4,5-tetrasubstituted benzene [$\delta_{\rm C}$ 133.6 s, 104.7 d (2C), 148.9 s (2C), 134.5 s; $\delta_{\rm H}$ 6.65 s], 2.0 Hz), one 1,3,4-trisubstituted benzene [δ_c 133.0 s, 135.6 d, 118.3 s, 155.6 s, 116.7 d, 134.6 d; $\delta_{\rm H}$ 7.88 d J = 2.2, 6.97 d J = 8.6, 6.97 d d J = 8.6, 2.0], one methoxycarbonyl group ($\delta_{\rm C}$ 168.2, 52.2; $\delta_{\rm H}$ 4.12 s), one singlet signal of two identical methoxyl groups (δ_C 56.1 q; δ_H 3.86 s) and two phenolic hydroxyl groups ($\delta_{\rm H}$ 11.8 s, 12.2 s). Strong absorption bands accounting for hydroxy (3418 cm⁻¹), carbonyl (1718) and aromatic groups (1605, 1538, 1425 cm⁻¹) could be observed in its IR spectrum. The UV absorptions at 210, 282 and 330 also suggested the presence of a conjugated aromatic ring system. The HMBC correlations (Fig. 2) of H-2,6 ($\delta_{\rm H}$ 6.65) with C-1' (δ_{C} 133.0) and of H-2' (δ_{H} 7.88) and H-6' (δ_{H} 6.97) with C-1 (133.6) suggested that compounds 1 should be a biphenyl derivative⁸. In the HMBC experiment, the methoxy protons affected the signal of two equivalent aromatic protons (H-2 and H-6), suggesting that two identical methoxyl groups were located at C-3 and C-5 (δ_c 148.9) of the tetrasubstituted benzene ring. HMBC cross-peaks between H-2,6 with C-3,4 and C-5 also confirmed their location. The methoxycarbonyl group at C-6 was supported by HMBC correlations of H-2' with the ester carbonyl carbon ($\delta_{\rm C}$ 168.2) and no correlation was observed between H-5' and the ester carbonyl. Tow phenolic hydroxy groups located at C-4 and C-4' were supported by the HMBC correlations of one phenolic hydroxy proton

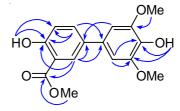


Fig. 2. Selected HMBC (
) correlation of 1

signal ($\delta_{\rm H}$ 11.8) with C-4 ($\delta_{\rm C}$ 134.5) and C-3,5 ($\delta_{\rm C}$ 148.9); and of another phenolic hydroxy proton signal ($\delta_{\rm H}$ 12.2) with C-3' ($\delta_{\rm C}$ 118.3), C-4' ($\delta_{\rm C}$ 155.6) and C-5' ($\delta_{\rm C}$ 116.7), respectively.

Compound **1** was tested for its cytotoxicity against five human tumor cell lines (NB4, A549, SHSY5Y, PC3 and MCF7) using the MTT method as reported previously⁹. The cytotoxicity tests for compound was performed against NB4, A549, SHSY5Y, PC3 and MCF7 tumor cell lines by MTTassay with paclitaxel as the positive control. The results shown that compound **1** exhibited moderate cytotoxicity against A549 and PC3 cell with IC₅₀ values of 6.4 and 5.7 μ M, respectively.

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