



## A New Sesquiterpene from *Nicotiana tabacum* and Its Cytotoxicity

YONG-KUAN CHEN<sup>1</sup>, CHUNYANG MENG<sup>1,2</sup>, SU ZHONGBI<sup>1</sup>, LU WANG<sup>1</sup>, GUANGYU-YU YANG<sup>1</sup> and MING-MING MIAO<sup>1,\*</sup>

<sup>1</sup>Key Laboratory of Tobacco Chemistry of Yunnan Province, Yunnan Academy of Tobacco Science, Kunming 650106, P.R. China

<sup>2</sup>Key Laboratory of Ethnic Medicine Resource Chemistry (Yunnan University of Nationalities), State Ethnic Affairs Commission & Ministry of Education, Kunming 650031, P.R. China

\*Corresponding author: E-mail: mmmiao@cyats.com

Received: 24 April 2013;

Accepted: 17 August 2013;

Published online: 15 April 2014;

AJC-15001

A new sesquiterpene, tabsesquiterpene A (**1**) was isolated from the leaves of *Nicotiana tabacum*. Its structure was elucidated by spectroscopic methods, including extensive 1D and 2D NMR techniques. Compound **1** was tested for its cytotoxicity. The results showed that **1** exhibited modest cytotoxicity against SHSY5Y, PC3 and MCF7 with IC<sub>50</sub> values of 6.9, 4.5 and 8.8 μM, respectively.

**Keywords:** *Nicotiana tabacum*, Sesquiterpene, Cytotoxicity, Tabsesquiterpene A.

### INTRODUCTION

*Nicotiana tabacum* L. is one of the most commercially valued agricultural crops in the world.<sup>1,2</sup> In addition to being used in cigarette industry, *N. tabacum* is also used as insecticide, anesthetic, diaphoretic, sedative and emetic agents in Chinese folklore medicine because of containing many useful chemical compounds<sup>1,3-5</sup>.

In previous work, a number of bioactive compounds, such as terpenoids<sup>6-8</sup>, alkaloids<sup>9,10</sup>, lignans<sup>11,12</sup>, flavonoid<sup>13</sup>, phenylpropanoids<sup>14</sup> and the homologous, were isolated from this plant. Motivated by search for bioactive metabolites from this plant, the investigation on the chemical constituents of the leaves of *N. tabacum* was carried out. As a result, a new sesquiterpene, tabsesquiterpene A (**1**), was isolated from this plant. In addition, the cytotoxicity of **1** was evaluated. This article deals with the isolation, structural elucidation and biological activities of this new compound.

### EXPERIMENTAL

**General experimental procedures:** Optical rotation was measured in Horiba SEPA-300 High Sensitive Polarimeter. IR spectra were obtained in KBr disc on a Bio-Rad Wininfrared spectrophotometer. APCIMS were measured on a VG Auto Spec-3000 MS spectrometer. <sup>1</sup>H, <sup>13</sup>C and 2D NMR spectra were recorded on Bruker DRX-500 instrument with TMS as internal standard. Column chromatography was performed on silica gel (200-300 mesh), or on silica gel H (10-40 μm,

Qingdao Marine Chemical Inc., China). Second separate was used an Agilent 1100 HPLC equipped with ZORBAX-C<sub>18</sub> (21.2 nm × 250 nm, 7.0 μm) column and DAD detector.

The leaves of *nicotiana tabacum* L (tobacco leaves) was collected from Yuxi County, Yunnan Province, P. R. China, in September 2011.

**Extraction and isolation:** The air-dried and powdered leaves of *nicotiana tabacum* (2.8 kg) were extracted with 70 % aqueous ethanol (3.5 L × 3, 24 h each) at room temperature and the extract was concentrated under vacuum condition. The dried extract (98.5 g) was applied to Si gel (200-300 mesh) column chromatography eluting with a CHCl<sub>3</sub>-Me<sub>2</sub>CO gradient system (9:1, 8:2, 7:3, 6:4, 5:5, 2:1) to give six fractions A-F. Fraction A1 (9:1, 6.47 g) was subjected to silica gel column chromatography using CHCl<sub>3</sub>-MeOH and preparative HPLC (75 % MeOH-H<sub>2</sub>O, flow rate 12 mL/min) to give **1** (26.8 mg).

**Cytotoxicity assays:** The cytotoxicities was tested against five human tumor cell lines (NB4, A549, SHSY5Y, PC3 and MCF7) using the MTT method as reported previously<sup>16</sup>. Taxol was used as the positive control. The results shown that compounds **1** exhibited modest cytotoxicity against SHSY5Y, PC3 and MCF7 with IC<sub>50</sub> values of 6.9, 4.5 and 8.8 μM, respectively.

**Tabsesquiterpene A:** Obtained as obtained as a colourless oil; [α]<sub>D</sub><sup>24.5</sup> +32.1° (c = 0.1, MeOH); UV (MeOH), λ<sub>max</sub> (log ε) end absorbance; IR (KBr, ν<sub>max</sub>, cm<sup>-1</sup>): 3415, 2926, 1762, 1736, 1639, 1547, 1476, 1136, 1075, 976; <sup>1</sup>H and <sup>13</sup>C NMR data (CDCl<sub>3</sub>, 500 and 125 MHz), Table-1; APCIMS (negative ion mode) *m/z* 339; HRAPCIMS (negative ion mode) *m/z* 339.1449 [M-H]<sup>-</sup> (calcd. (%) 339.1444 for C<sub>17</sub>H<sub>23</sub>O<sub>7</sub>).

TABLE-1  
<sup>1</sup>H AND <sup>13</sup>C NMR DATA OF COMPOUND **1** IN CDCl<sub>3</sub>

No.	δ <sub>c</sub> (mult.)	δ <sub>H</sub> (mult., J, Hz)
1	43.8 d	7.76 (m)
2	38.5 t	2.08 (m); 1.67 (m)
3	78.0 d	3.58 (m)
4	48.2 d	1.75 (m)
5	53.0 d	1.92 (m)
6	83.5 d	4.08 (dd, J = 10.2, 9.6)
7	62.3d	2.75 (dd, J = 10.2, 9.6)
8	70.9 d	2.74 (m)
9	48.2 t	2.75 (dd, J = 12.0, 4.2), 2.09 (brd, 12.0)
10	145.8 s	–
11	75.8 s	–
12	181.2 s	–
13	66.8 t	4.55 (d, J = 10.8)
	–	4.22 (d, J = 10.8)
14	115.2 t	4.99 (s); 5.08 (s)
15	19.2 q	1.28 (d, J = 6.5)
1'	169.5 s	–
2'	20.8 q	2.12 (s)

## RESULTS AND DISCUSSION

A 70 % aq. methanol extract prepared from the leaves of *N. tabacum* was subjected repeatedly to column chromatography on silica gel, sephadex LH-20, RP-18 and preparative HPLC to afford compound **1**<sup>15</sup>. Its structure was shown in Fig. 1 and its <sup>1</sup>H and <sup>13</sup>C NMR data was listed in Table-1.

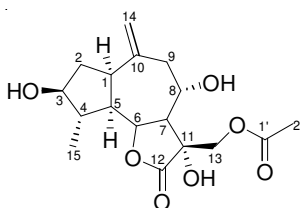


Fig. 1. Structure of new sesquiterpene

Compound **1** was obtained as colourless oil. The negative-ion APCIMS showed a quasi-molecular ion peak [M-H]<sup>-</sup> at *m/z* 339. The molecular formula, C<sub>17</sub>H<sub>24</sub>O<sub>7</sub>, was established by HRAPCIMS (*m/z* 339.1449 [M-H]<sup>-</sup>; calcd. (%) 339.1444), indicating six degrees of unsaturation. The IR spectrum of **1** exhibited absorption at 3415, 1762, 1736 and 1639 cm<sup>-1</sup> ascribable to hydroxyl, γ-lactone and C=C functional groups. The <sup>1</sup>H NMR spectrum (Table-1) displayed one secondary methyl at δ<sub>H</sub> 1.28 (3H, d, J = 6.5 Hz, H-15) and one acetyl methyl at δ<sub>H</sub> 2.12 (3H, s, H-2'). The <sup>13</sup>C NMR and DEPT spectra (Table-1) revealed 17 carbon resonances including two methyl, four methylenes, seven methines, four quaternary carbons. The lactone carbonyl resonances were located at δ<sub>c</sub> 83.5 (d, C-6), 181.2 (s, C-12), four oxygenated carbon resonances were observed at δ<sub>c</sub> 78.0 (d, C-3), 70.9 (d, C-8), 75.8 (s, C-11) and 66.8 (t, C-13), exocyclic methylene resonances at δ<sub>c</sub> 145.8 (s, C-10) and 115.2 (t, C-14), an acetoxy group at δ<sub>c</sub> 169.5 (s, C-1') and 20.8 (q, C-2'), respectively.

The <sup>1</sup>H and <sup>13</sup>C spectra data of **1** was very similar to these of 3β,8α,11α,13-tetrahydroxy-10 (14)-guaien-1α,4β,5α,6βH-6α,12-olide,<sup>15</sup> except for the additional acetoxy group signal [δ<sub>c</sub> 169.5 (s, C-1') and 20.8 (q, C-2'); δ<sub>H</sub> at 2.12 (s, H-2')] of compound **1**. The partial structural unit C-1 to C-9 was deduced

from the analysis of the HSQC and <sup>1</sup>H <sup>1</sup>H COSY spectra of **1**. The HMBC correlations (Fig. 2) between the AB system signals at δ<sub>H</sub> 4.22 (1H, d, J = 10.8 Hz, Hα-13) and δ<sub>H</sub> 4.55 (1H, d, J = 10.4 Hz, Hβ-13) and C-12 and the oxygenated quaternary carbon signal at δ<sub>c</sub> 75.8 (s, C-11) confirmed two hydroxyl groups attached to C-11 and C-13, respectively. The acetoxy group attached at C-13 was supported by the HMBC correlation of H-13 [δ<sub>H</sub> 4.55 (d, J=12.0 Hz), 4.22 (d, J = 12.0 Hz)] with the acetoxy carbonyl carbon δ<sub>c</sub> 169.5 (s, C-1'). The HMBC showed the cross-peaks of H-15 [δ<sub>H</sub> 1.28 (d, J = 6.5 Hz)] to C-3, C-4 and C-5, H-6 [δ<sub>H</sub> 4.08 (dd, J = 10.4, 10.1 Hz)] to C-12 (δ<sub>c</sub> 181.2), H-14 [δ<sub>H</sub> 4.99 (s), 5.08 (s)] to C-1, C-9 and C-10 and H-9 [δ<sub>H</sub> 2.75 (dd, J = 12.0, 4.2 Hz), 2.09 (brd, J = 12.0 Hz)] to C-1, C-10 and C-14, respectively. These informations confirmed the planar structure of **1**.

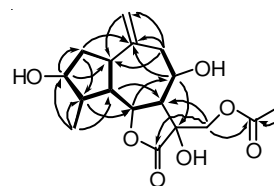


Fig. 2. <sup>1</sup>H <sup>1</sup>H COSY (---) and selected HMBC (—) correlations of **1**

The relative configuration of **1** was determined by comparison with 3β,8α,11α,13-tetrahydroxy-10 (14)-guaien-1α,4β,5α,6βH-6α,12-olide<sup>15</sup> and confirmed by a ROESY experiment. The ROESY correlations (Fig. 3) of Hα-1 with H-3 and Hα-5 with H-3, H-7 and Me-15 indicated that H-3, H-7 and Me-15 possessed α-orientations, respectively. The ROESY correlations of H-6 with H β-4 and H-8 suggested that H-6 and H-8 possessed β-orientations, respectively. On the basis of the evidence mentioned above, the structure of **1** was elucidated as shown and gives the trivial name of tabsesquiterpene A.

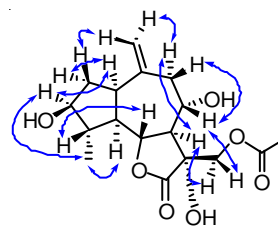


Fig. 3. Key ROESY (↔) correlations of **1**

## ACKNOWLEDGEMENTS

This project was supported financially by the Basic Research Foundation of Yunnan Tobacco Industry Co. Ltd (2012JC01), the National Natural Science Foundation of China (No. 31360081), the Excellent Scientific and Technological Team of Yunnan High School (2010CI08).

## REFERENCES

1. The Editorial Committee of the Administration Bureau of Flora of China, *Flora of China*, Beijing Science and Technology Press, Beijing, Vol. 67 (2005).
2. T.W. Hu and Z. Mao, *Tob. Control*, **15**(S), i37 (2006).
3. A. Rodgman and T.A. Perfetti, *The Chemical Components of Tobacco and Tobacco Smoke*. CRC Press, Taylor and Francis Group, Boca Raton, Florida (2008).

4. A.P. Cavender and M. Alban, *J. Ethnobiol. Ethnomed.*, **5**, 3 (2009).
5. A. Inta, P. Shengji, H. Balslev, P. Wangpakapattanawong and C. Trisonthi, *J. Ethnopharmacol.*, **116**, 134 (2008).
6. X. Feng, J.-S. Wang, J. Luo and L.-Y. Kong, *J. Asian Nat. Prod. Res.*, **12**, 252 (2010).
7. Y. Shinozaki, T. Tobita, M. Mizutani and T. Matsuzaki, *Biosci. Biotechnol. Biochem.*, **60**, 903 (1996).
8. T. Pettersson, A.M. Eklund and I. Wahlberg, *J. Agric. Food Chem.*, **41**, 2097 (1993).
9. X.C. Wei, S.C. Sumithran, A.G. Deaciuc, H.R. Burton, L.P. Bush, L.P. Dvoskin and P.A. Crooks, *Life Sci.*, **78**, 495 (2005).
10. T. Braumann, G. Nicolaus, W. Hahn and H. Elmenhorst, *Phytochemistry*, **29**, 3693 (1990).
11. Y.K. Chen, X.S. Li, G.Y. Yang, Z.Y. Chen, Q.F. Hu and M.M. Miao, *J. Asian Nat. Prod. Res.*, **14**, 450 (2012).
12. Q.F. Hu, G. Yang, X. Li, X. Yang, H. Mu, Y. Chen and X.M. Gao, *Heterocycles*, **85**, 147 (2012).
13. Z.Y. Chen, J.L. Tan, G.Y. Yang, M.M. Miao, Z.Y. Chen and T.F. Li, *Phytochem. Lett.*, **5**, 233 (2012).
14. J.L. Tan, Z.Y. Chen, G.Y. Yang, M.M. Miao, Y.K. Chen and T.F. Li, *Heterocycles*, **83**, 2381 (2011).
15. R. Liu, K.L. Hsieh and J.K. Liu, *Bot. Yunnan*, **31**, 383 (2005).
16. T. Mosmann, *J. Immunol. Methods*, **65**, 55 (1983).