



Isolation and Identification of Insecticidal Component from Roots of *Stellera chamaejasme* L. Against *Locusta migratoria manilensis*

LANG WU¹, TAO PU², MANLU GAO², LONG CHEN², KE TAO², KUN LIU² and TAIPING HOU^{1*}

¹Key Laboratory of Bio-Resource and Eco-Environment of Ministry of Education, Sichuan University, Chengdu 610064, Sichuan, P.R. China

²College of Life Science, Sichuan University, Chengdu 610064, Sichuan Province, P.R. China

*Corresponding author: Fax: +86 28 85214048; Tel: +86 28 88846045; E-mail: houtplab@scu.edu.cn

Received: 11 February 2013;

Accepted: 12 April 2013;

Published online: 26 December 2013;

AJC-14468

In order to find the effective bio-pesticide to against locusts, we tested the bioactivity of the different extracts from roots of *Stellera chamaejasme* L., respectively. The results of bioassay indicated that petroleum ether extracts had obviously insecticidal activity towards locusts. With silica column chromatography method, we isolated one compound from petroleum ether extracts. Spectra data of ¹NMR and ¹³NMR showed the insecticidal compound in the roots of *Stellera chamaejasme*.L against *Locusta migratoria manilensis* was dibutyl phthalate. The LC₅₀ of dibutyl phthalate was found to be 292.68 ppm.

Keywords: *Stellera chamaejasme* L., *Locusta migratoria manilensis*, Dibutyl phthalate, Insecticidal activity.

INTRODUCTION

Stellera chamaejasme L. is a species of *Stellera* L. There are about 90 species *Stellera* L. and widely distributed in Tibet, Qinghai and Sichuan province of China¹. Compared with other grassland plants, *Stellera chamaejasme* L. was viewed as a harmful and destructive plant for its strong viability and fertility. Not only it brought the troubles to the development of animal husbandry, but also threatened the protection of grassland. On the other side, *Stellera chamaejasme* L. has long been used as the good botanical pesticides by local people. Extracts of *Stellera chamaejasme* L. have been reported to have a wide spectrum of biological activities including anti-fungal², antibacterial³, antitumor⁴ and insecticidal activity^{5,6}, the research of Li Jie found that the petroleum ether extracts of *Stellera chamaejasme* L. had a strong insecticidal activity against *Tetranychus viennensis*, the LC₅₀ was 180 mg/L⁷. Consequently, *Stellera chamaejasme* L. was regarded as new generation insecticide candidates in recent years.

Locust is a kind of worldwide harmful insects. More than 800 species are known in China and about 50 species are harmful to agriculture, forestry and animal husbandry⁸. Every year the locusts causes tremendous losses. Chemical pesticide was efficient to control locust, while it caused a series side effects, such as resistance and environment pollution⁹. It is desirable to find a new class of insecticide which active against locust and friendly to environment.

EXPERIMENTAL

Root of *Stellera chamaejasme* L. was collected from Ruoergai plateau, Sichuan Province of China in 2010. The roots was naturally air dried and grinded with electric mill. All the 3 instars locusts were obtained from Mushan locust farm, Chengdu, Sichuan Province of China and feed on fresh leaves of ryegrass. Gas chromatograph/mass spectrometer (Shimadzu, Japan) was used to qualitative analysis of compound. Bruker-AV II -600MHz type nuclear magnetic resonance was used to collect the ¹H and ¹³C NMR spectrum data to identify the compound structure.

Extraction and isolation: The air-dried and ground plant material (100 g) was extracted exhaustively three times with 1 L of solvent mixture ethanol/water (95:5, v/v) for 24 h at 45 °C. The solvents was evaporated in *vacuo* to yield the total extracts (19 g). This extract was then suspended in ethanol and extracted with petroleum ether, methylene chloride, acetic ether and methyl alcohol, respectively. The insecticidal activity of each extracts were applied to the locusts.

The petroleum ether extracts (1 g) was subjected to silica gel column chromatography(CC) and eluted with a gradient of *n*-hexane and ethyl acetate (10:1,5:1,2:1,1:1, ethyl acetate) to give five fractions (R1-R5). The bioassay showed that fraction of R5 had significant insecticidal activity than others. The fraction R5 (100 mg) was further separated by a silica gel column chromatography with petroleum ether:acetic ether

(10:1, 7:1, 5:1, 3:1 and ethyl acetate) to obtain three fractions (R51-R53). All fractions were evaluated for insecticidal activity to locust (Table-1). Fraction R52 showed obviously insecticidal activity. The structures of R52 was elucidated by spectroscopic analyses, notably GC-MS and NMR whose structure was presented in Fig. 1.

Fractions	Time (h)	Regression equation	Value of regression	LC ₅₀ (ppm)
R51	120	y = 0.484x + 2.774	R ² = 0.92	39126.84
	144	y = 0.452x + 3.089	R ² = 0.86	16705.69
	168	y = 0.655x + 2.578	R ² = 0.86	4952.73
R52	120	y = 1.512x + 1.127	R ² = 0.98	362.91
	144	y = 1.512x + 1.666	R ² = 0.98	342.01
	168	y = 1.431x + 1.470	R ² = 0.95	292.68
R53	120	y = 1.261x + 1.555	R ² = 0.99	539.15
	144	y = 1.344x + 1.413	R ² = 0.99	465.13
	168	y = 1.476x + 1.228	R ² = 0.97	357.84

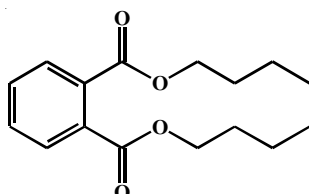


Fig. 1. Structure of compound R52

Compound R52 (17 mg) was obtained as a colourless oil and showed a blue spot under the 254 nm UV light on TLC. ¹H and ¹³C NMR data of compound R52 was as follows: ¹H NMR (600 MHz, CDCl₃) δ_H: 7.71(2H, dd, J = 3.5, 5.5 Hz, H-3,6), 7.53 (2H, dd, J = 3.5, 5.5 Hz, H-1,2), 4.31 (4H, t, J = 6.6 Hz, H = 8,8'), 1.72 (4H, m, J = 6.6 Hz, H = 9,9'), 1.44 (4H, m, J = 7.3 Hz, H = 10,10'), 0.97 (6H, t, J = 7.3 Hz, H = 11, 11'). ¹³C NMR (600 MHz, CDCl₃) δ_C: 167.73 (C-7,7), 132.33 (C-1,2), 130.92 (C-4,5), 128.85 (C-3,6), 65.58 (C-8,8'), 32.13 (C-9,9'), 19.19 (C-10,10'), 13.73 (C-11,11'). Meanwhile, the assignment was reconfirmed by HMQC and HMBC experiments.

Insecticidal activity assay: Toxicity of various solvent extracts was screened against 3 instars locusts using WHO method¹⁰. Five concentrations of the extracts were designed as 20000, 10000, 5000, 2500 and 1250 ppm. The sample was dissolved in 1 mL of acetone and dispersed in 10 Twain water. Thirty locusts (3 instars) were collected and transferred into the different concentration extracts for 1 s. Control were treated with acetone and 10 Twain water under similar conditions. Each test were replicated three times and kept in a room at 26 ± 1 °C. Effects of treatments on the mortality were checked each 24 h. Insecticidal activity was calculated by using of Abbott's formula¹¹:

$$\text{Mortality (\%)} = \frac{N_1}{N} \times 100$$

N₁: the number of death insects, N: the number of total insects.

$$\text{Corrected mortality (\%)} = \frac{M_1 - M_2}{1 - M_2} \times 100$$

M₁: the rate of mortality in control group, M₂: the rate of mortality in treatment group.

The mean mortality data of three replicates per dose were used to calculate the LC₅₀¹².

RESULTS AND DISCUSSION

Preliminary screening showed that the petroleum ether extract was most toxic against locust, followed by methylene chloride, acetic ether and methyl alcohol. The insecticidal activity of the fractions R1-R5 in controlling the locusts were tested at the concentration of 5 mg/mL (Fig. 2). After 24 h, the corrected mortality of those fractions were not much higher as expected. Nevertheless, with the concentration increased and time extended, there were significant differences in contact toxicity among the five fractions. Fractions R5 and R3 were dramatically increased within 48 h and the insecticidal activities were 89.65 and 44.82 %, respectively. After 96 h, the activity of R5 was 100 %. Meanwhile, we can see that the activity of R4 was the weakest fraction which the mean mortality only 13.33 % at 96 h. R5 showed the remarkably toxicity towards locust compared with other fractions from 24 to 96 h. Then R5 was further separated.

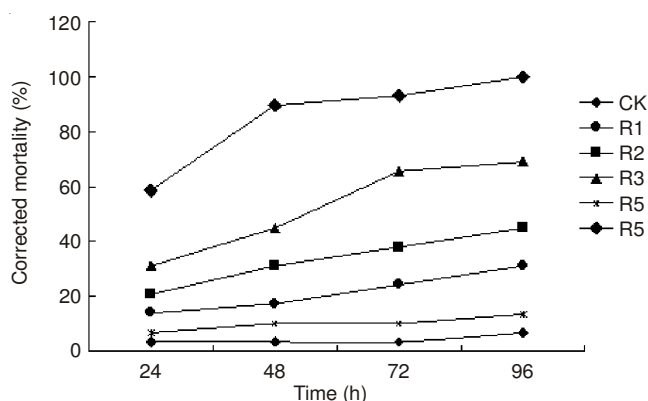


Fig. 2. Insecticidal activity of R-1 to R-5

Insecticidal activity of fraction R51, R52, R53 were noted and presented in Table-1. In our experiments, both R52 and R53 proved to be active against locust, the LC₅₀ were found to be 292.68 and 357.84 ppm, respectively. The proportion of R52 showed a good correlation (R² = 0.95) with their corresponding LC₅₀. This suggested that R52 is one parts of the primary active component in roots of *Stellera chamaejasme* L.

Bioactivity guided fractionation of petroleum ether extract ultimately led to the isolation of R52. Spectral data of R52 were well consistent with the literature¹³⁻¹⁵, the compound was identified as dibutyl phthalate. Dibutyl phthalate is a high production chemical and it have a wide spectrum of industrial and commercial applications, including plasticizers, solvents and in flexible plastics¹⁶. Xing *et al.*, first isolated dibutyl phthalate from roots of *Stellera chamaejasme* L¹⁷. Bioactivity of dibutyl phthalate has already been observed to be toxic to a variety of aquatic organisms¹⁸. Antimicrobial efficacy of dibutyl phthalate has also been reported from *Streptomyces*^{14,19} and it was used as peroxisome proliferator which was an effective compound against demodicidosis²⁰, as well as an endocrine disruptor with estrogenic activity²¹. While the insecticidal

activity of dibutyl phthalate were rarely reported. In this study, we had firstly tested the insecticidal activity of dibutyl phthalate against locust, although the toxicity were not very significant than other reported insecticide, while it would be assistance to investigators to design and preparation of useful and effective insecticide by structural modification. However, there have a lot of works to do, the exact role of dibutyl phthalate in pathogenesis were not well understood and we could not determine whether the observed insecticidal effect resulted from the action of both components of the mixtures or only of dibutyl phthalate. Further research on the application of dibutyl phthalate and the isolation of bioactive constituents from *Stellera chamaejasmev* L. are undergoing in our laboratory.

Conclusion

Different polar solvent extracts were evaluated for their insecticidal activities. By the bioactivity assay, it is concluded that the petroleum ether extracts from roots of *Stellera chamaejasmev* L. has the obviously insecticidal activity against locusts. With the further purification, we discovered that dibutyl phthalate was one of the active ingredients. It was the first time to report that dibutyl phthalate has the insecticidal activity towards locust and also it suggested that the class of phthalate can be used as the insecticide. Meanwhile, R53 also showed a very strong contact toxicity, it is necessary to further separation.

ACKNOWLEDGEMENTS

This study was financially supported by the National Natural Science Foundation of China (No. 31272068/C140501) and Hi-tech Research and Development Program of China (863 Program, No. 2011AA10A202-3) and National Key Technology Research and Development Program of the Ministry of Science and Technology of China (2011BAE06B-01-23).

REFERENCES

1. S. Zhicheng, Important poisonous Plants of China Grassland, Chinese Agriculture Publication, China, p. 322 (1997).
2. G. Xiaoxia, L. Wenjuan and O. Yangqiu, *J. Sichuan Univ.*, **43**, 687 (2006).
3. L. Wenjuan, G. Xiaoxia and H. Wang, *Acta Bot. Boreal.-Occident. Sin.*, **25**, 1661 (2006).
4. F. Weijian, I.K. Tetsuro and Y. Mitsuzi, *Chin. J. Cancer Res.*, **8**, 101 (1996).
5. T. Renjun, Z. Li, M. Zhou and Z. Chunguang, *J. Sichuan Univ.*, **42**, 1266 (2005).
6. Z. Guozhou, W. Yamei and X. Hanhong, *J. Anhui Agric. Univ.*, **29**, 163 (2002).
7. J. Li, F. Zhao and K. Weina, *J. Plant Resour. Environ.*, **16**, 31 (2007).
8. G. Shujin, L. Aiping and X. Linbo, *Modern AGC*, **9**, 44 (2010).
9. W. Wenjuan and R. Bingzong, *J. Beihua Univ.*, **6**, 481 (2002).
10. World Health Organization (WHO), Report of the WHO Informal Consultation on the Evaluation on the Testing of Insecticides, Geneva, Switzerland, p. 96 (1996).
11. W.S. Abbott, *J. Econ. Entomol.*, **18**, 265 (1925).
12. Z. Zhixiang, X. Hanhong and C. Dongmei, *Entomol. Know.*, **39**, 67 (2002).
13. S. Wei, H. Liang, Y.Y. Zhao and Y.Y. Zhang, *J. Chin. Mater. Med.*, **22**, 293 (1997).
14. X.-Y. Qu, Q.-Q. Gu, C.-B. Cui, Y.-C. Fang, H.-B. Liu, T.-J. Zhu and W.-M. Zhu, *Chin. J. Mar. Drugs*, Issue No. 6, 1 (2004).
15. M.E. Savard, J.D. Miller, L.A. Blais, K.A. Seifert and R.A. Samson, *Mycopathologia*, **127**, 19 (1994).
16. O. Bajt, G. Mailhot and M. Bolte, *Appl. Catal. B: Environ.*, **33**, 239 (2001).
17. X. Youquan, L. Fengqin and C. Nianhai, *J. Heilongjiang Univ.*, **7**, 74 (1990).
18. M. Muneer, J. Theurich and D. Bahnemann, *J. Photochem. Photobiol. A: Chem.*, **143**, 213 (2001).
19. M.Y.M. El-Naggar, *Biomed. Lett.*, **55**, 125 (1997).
20. F.S Yuan, S.-L. Guo, Z.-X. Qiu, S.-H. Deng and G.-H. Huang, *Chin. J. Parasitol. Parasit. Dis.*, **19**, 160 (2001).
21. H. Ohtani, I. Miura and Y. Ichikawa, *Environ. Health Perspect.*, **108**, 1189 (2000).