



Mechanical Studies on Chitosan Functionalized Multiwalled Carbon Nanotube Reinforced Epoxy Bionanocomposites

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The chitosan functionalized multiwalled carbon nanotubes reinforced epoxy bionanocomposite (CS-g-MWCNTs/EP) was developed. Chitosan (CS) was grafted to multiwalled carbon nanotubes (MWCNTs), first by reacting the oxidized carbon nanotubes (MWCNT-COOH) with thionyl chloride to form acyl-chlorinated multiwalled carbon nanotubes (MWCNT-COCl) followed by subsequent dispersion in chitosan. The chitosan grafted multiwalled carbon nanotubes (CS-g-MWCNTs) were reinforced in the epoxy matrix using high speed stirrer and then cured. The TEM of CS-g-MWCNTs shows a visible coating of chitosan on the surface of MWCNTs. FT-IR spectra of CS-g-MWCNTs indicate an overlap of the amide band and the free amino groups of the chitosan. The mechanical properties such as tensile and impact strength of epoxy matrix increases to 16 and 52 %, respectively by the reinforcement of 1.5 wt. % CS-g-MWCNTs, which clearly reveals that the free amino groups of CS-g-MWCNTs acted as a curing agent and was covalently attached into epoxy matrix. Morphologies of pure chitosan, MWCNT, MWCNT-COOH and CS-g-MWCNTs/EP bionanocomposites were studied by scanning electron microscope.

Keywords: Chitosan, Multiwalled carbon nanotubes, Bionanocomposites, Mechanical properties, Morphology.

INTRODUCTION

Organic light emitting diodes and organic photovoltaics are susceptible to chemical degradation in the presence of oxygen and moisture. The sensitivity of these materials towards oxygen and moisture makes it imperative to protect them by encapsulation. High quality epoxy resins, having exceptional combination of properties such as toughness, adhesion and chemical resistance¹, are used for effective fixture and encapsulation of the electronic components. The epoxy encapsulated electronic parts are protected from adverse environmental, mechanical, thermal and electrical stresses. Carbon nanotubes (CNTs) have unique molecular structures and outstanding thermal, electronic, optical and mechanical properties²⁻⁶. Furthermore, they are potentially useful fillers in nanocomposites with polymers, metals and ceramics⁷⁻¹⁰. Addition of conductive fillers in an insulating matrix dramatically increases the electrical properties when a network was developed throughout the matrix. The increase in electrical properties of the composites depends on the property of the conducting fillers, dispersion of conducting fillers in polymeric matrix and interaction between the fillers and the polymer¹¹⁻¹⁴. The incorporation of CNTs into polymer bulk materials will surely enhance their thermal and electrical properties¹⁵. An enhancement of

the compatibility between the CNTs and resin matrix can improve the electrical and mechanical properties of the composite materials. Chitosan is a natural polymer generally obtained by extensive deacetylation of chitin and has many industrial and pharmaceutical applications¹⁶. Decorating CNTs with non-conducting natural polymers will decrease its electrical conductance. Further, reinforcement of biopolymer grafted CNTs onto polymer matrix will undergo degradation easily.

In this work chitosan grafted MWCNTs (CS-g-MWCNTs) and CS-g-MWCNTs/EP bionanocomposites were developed and their structural, mechanical, thermal and morphological properties were investigated.

EXPERIMENTAL

Commercially available diglycidyl ether of bisphenol-A based epoxy resin (DGEBA), LY556, having epoxy equivalent = 180-190, with viscosity = 10,000 cP and 4,4'-diamino-diphenyl-methane (DDM), epoxy curing agent were obtained from Ciba-Geigy Ltd., India. Multiwalled carbon nanotubes (MWCNTs) with surface area = 200 m²/g, diameter = 10-20 nm, length > 10 μ and purity > 98 % was purchased from Chemapol Industries Ltd, India. Chitosan having molecular mass (Mv) = 1,80,000 Da, moisture content = 9.46 %, viscosity = 229 cP, DD = 86.39 was provided by Indian sea foods, Cochin, India.

Purification of chitosan: 10 g of chitosan was dissolved in 200 mL of 1 % acetic acid solution, then filtered, using glass wool to remove insoluble sample. 200 mL of 2 M sodium hydroxide was added to the filtered sample with continuous stirring. After removal of excess NaOH solution, the filtrate was washed with distilled water until a pH 7 was reached. The structure of chitosan is shown in Fig. 1.

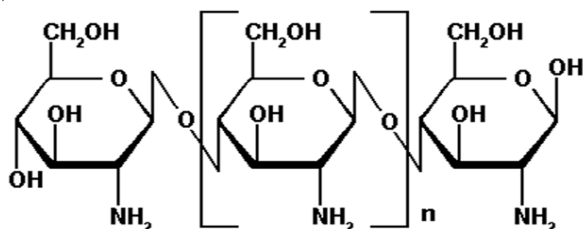


Fig. 1. Structure of chitosan

Preparation of CS-g-MWCNTs composites: 2 g of MWCNTs was dissolved in 100 mL conc. HCl for the removal of metal impurities. Then it was washed with distilled water until become neutral. Multiwalled carbon nanotubes were functionalized with carboxyl group by sonication at 40 °C for 2 h with a 3:1 (v/v) acid mixture (98 % H_2SO_4 and 65 % HNO_3). The oxidized carbon nanotubes (MWCNT-COOH) were suspended in a solution of thionyl chloride and refluxed for 24 h at 75 °C to convert carboxyl groups to acyl chloride groups. The acyl chloride functionalized carbon nanotubes (MWCNT-COCl) were separated by membrane filtration, washed with THF and dried in an oven. Chitosan was mixed with MWCNT-COCl in DMF and refluxed at 120 °C for 96 h under nitrogen atmosphere. After reaction, the mixture was filtered through a 0.22 μm polyether sulphone membrane and dried in a vacuum oven to obtain black chitosan grafted MWCNT (CS-g-MWCNTs) composite powder¹⁷. Fig. 2 represents the formation of CS-g-MWCNTs.

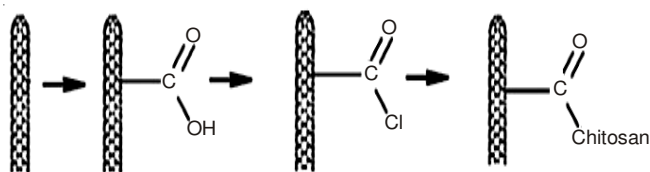


Fig. 2. Schematic representation of chitosan grafted MWCNTs

Preparation of neat epoxy matrix: 100 g of epoxy resin (DGEBA) was heated to 70 °C in an oil bath for the removal of trapped air bubbles and moisture. The amine hardener (DDM) (27 % in accordance with the epoxy resin) was also heated to 90 °C to melt completely. Both the resin and hardener were mixed with continuous stirring and then poured into mould, cured in an air oven at 100 °C for 4 h and post cured at 140 °C for 3 h.

Preparation of CS-g-MWCNTs/epoxy bionanocomposites: Calculated amount of CS-g-MWCNTs (0.1, 0.5, 1.0, 1.5, 2.0 g) were added to the epoxy resin and the mixture was sonicated for 2 h followed by continuous stirring for 20 min. at 12000 rpm. The resin mixture, benzoyl peroxide and the hardener (DDM) were mixed with continuous stirring and then cured and post cured in an air oven.

Analytical methods: FT-IR spectra of MWCNTs, MWCNT-COOH, chitosan and CS-g-MWCNTs were recorded on a Perkin-Elmer Spectrometer in the range of 4000-400 cm^{-1} at room temperature. The thermal analysis was carried out by a Du Pont Differential Scanning Calorimeter with a heating rate of 10 °C/min. Tensile strength was determined as per ASTM-D3039 using a universal testing machine (Instron, Model 6025 UK), at a cross-head speed of 10 mm/min. with specimens of width 25 mm, length 200 mm and thickness 3 mm. Unnotched Impact strength was investigated as per ASTM-D256-88 using Izod impact tester with specimens of 3 mm thickness, 10 mm width and 90 mm length. For both studies five specimens were tested for each sample. Transmission Electron Microscopy was recorded using TEM 100CX (JEOL) with an accelerating voltage of 100 keV to find the size and shape of MWCNTs and CS-g-MWCNTs. Surface morphology of fractured surface of the composites were recorded using scanning electron microscope (SEM; JEOL JSM Model 6360).

RESULTS AND DISCUSSION

FT-IR spectroscopy: FT-IR spectroscopy was used to characterize the chemical components on the MWCNTs surface and the result shown in Fig. 3. Although there were no characteristics peaks in the FT-IR spectra of MWCNTs [Fig. 3(a)], the MWCNT-COOH shows several vibrational peaks, carboxylic C=O stretching vibration at 1635 cm^{-1} , O-H stretching vibration at 3440 cm^{-1} and C-H stretching and bending vibrations at 2889, 1209 and 1149 cm^{-1} ¹⁸. These results indicate that carboxylic acid groups have been covalently attached to the surface of the MWCNTs [Fig. 3(b)].

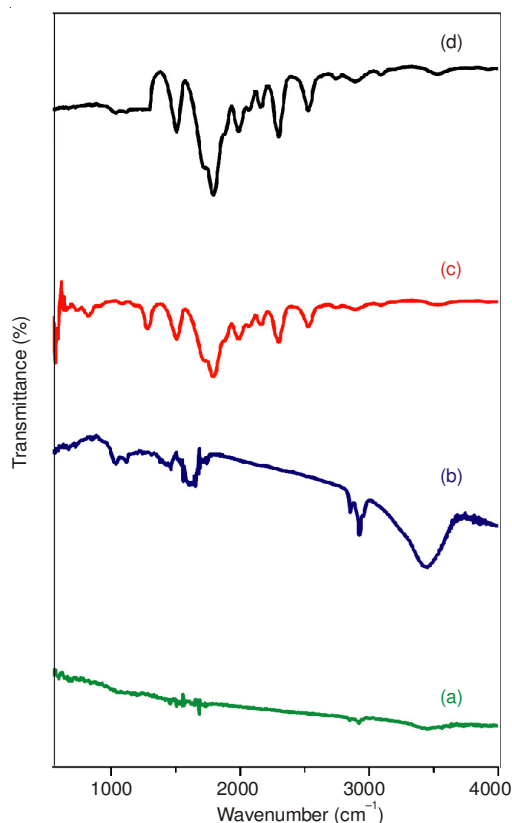


Fig. 3. FT-IR of (a) MWCNTs, (b) MWCNT-COOH, (c) Chitosan, (d) CS-g-MWCNT

FT-IR spectra of chitosan: Fig. 3(c) represents the following: C=O stretching vibration of acetyl unit at 1647 cm^{-1} , C-O-C stretching vibration at 1091 cm^{-1} , N-H bending vibration at 1585 cm^{-1} and β (1, 4) glycosidic bond vibration at 1151 cm^{-1} . The broad peak at 3431 cm^{-1} due to -OH stretching vibration is superimposed on -NH stretching band and broaden due to inter hydrogen bonds of polysaccharides.

The FT-IR of CS-g-MWCNTs is shown in Fig. 3(d). When compared to MWCNT-COOH Fig. 3(b) a new peak due to stretching vibrations of -OH and -NH is obtained at 3430 cm^{-1} . Additionally a new absorption peak at 1586 cm^{-1} indicates an overlap of the amide band and the amino group of the chitosan^{19,20}. Similarly, peaks due to β (1, 4) glycosidic bonds and C-O-C stretching vibration are shifted to 1123 and 1041 cm^{-1} , respectively. The above results confirm that chitosan is grafted on the MWCNTs surfaces.

Mechanical properties: The observed values for tensile and unnotched impact properties of the CS-g-MWCNTs reinforced epoxy bionanocomposites are shown in Figs. 4 and 5. Fig. 4 shows that the tensile strength of the bionanocomposites increases from 72.5 M.Pa (neat epoxy) to 87.54 M.Pa (1.5 wt. % CS-g-MWCNTs). Fig. 5 shows the unnotched impact

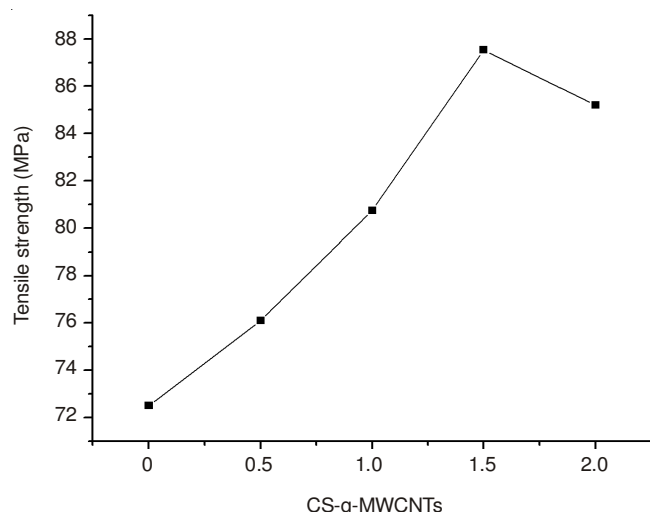


Fig. 4. Effect of CS-g-MWCNTs on the tensile strength of CS-g-MWCNTs/EP bionanocomposites

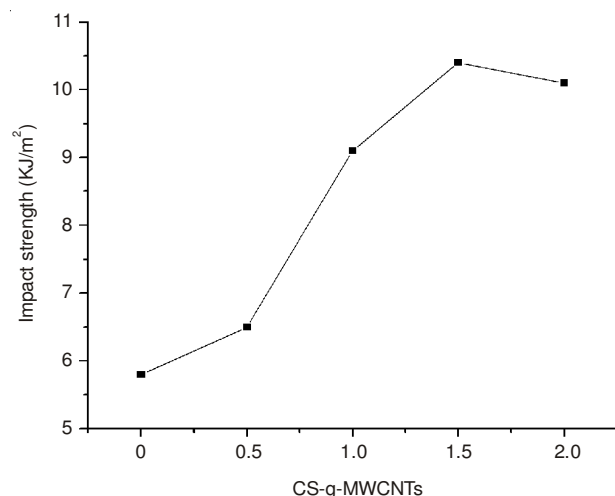


Fig. 5. Effect of CS-g-MWCNTs on the impact strength of CS-g-MWCNTs/EP bionanocomposites

strength of bionanocomposites increases from 5.6 KJ/m^2 (neat epoxy) to 10.5 KJ/m^2 (1.5 wt. % CS-g MWCNTs). The significant improvement in mechanical properties is due to the formation of covalent bond between free amine group of CS-g-MWCNTs and epoxy matrix which leads to effect stress transfer. However, as CS-g-MWCNTs content increases to 2 wt. %, the tensile strength and impact strength decreases slightly due to agglomeration.

TEM analysis: TEM image of CS-g-MWCNTs (Fig. 6b) shows the immobilization of chitosan on the surface of MWCNTs and is quite visible when compared to TEM of pure MWCNTs (Fig. 6a). This is the clear indication of functionalization of chitosan on the surface of MWCNTs.

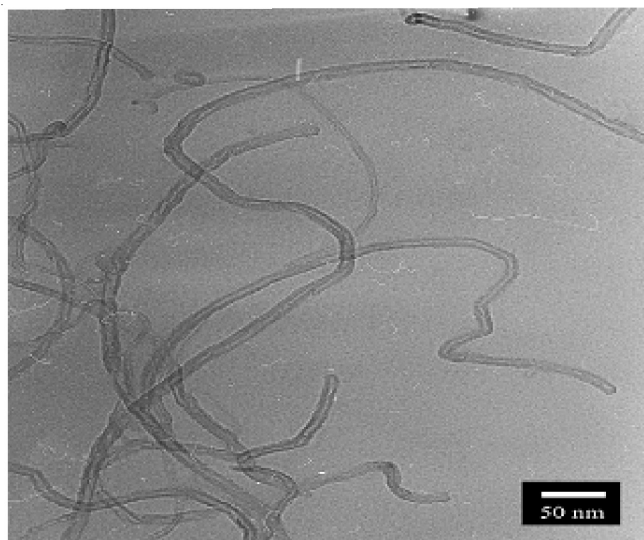


Fig. 6. (a) TEM image of MWCNTs

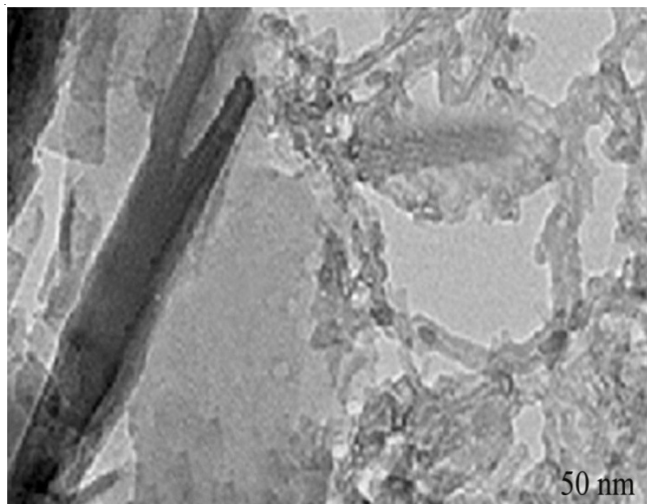


Fig. 6. (b) TEM image of CS-g-MWCNTs

SEM analysis: SEM micrograph of unmodified MWCNTs (Fig. 7a) reveals a clean and smooth surface with bundles. In contrast, the SEM of MWCNT-COOH (Fig. 7b) shows separation of bundles.

Fig. 7c and 7d shows the SEM images of tensile fracture surfaces for MWCNT-COOH/EP composites and CS-g-MWCNTs/EP bionanocomposites, respectively. The fracture surface of MWCNT-COOH reinforced epoxy nanocomposites indicates

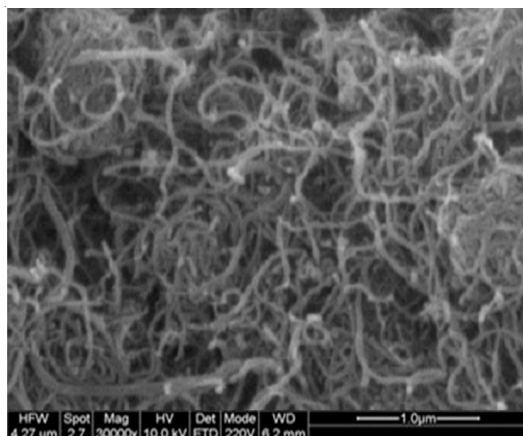


Fig. 7. (a) SEM of unmodified MWCNTs

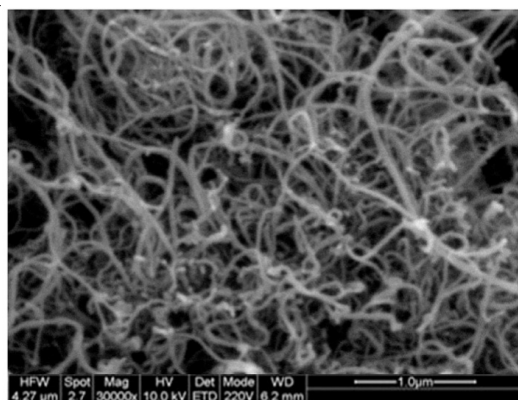


Fig. 7. (b) SEM of MWCNT-COOH

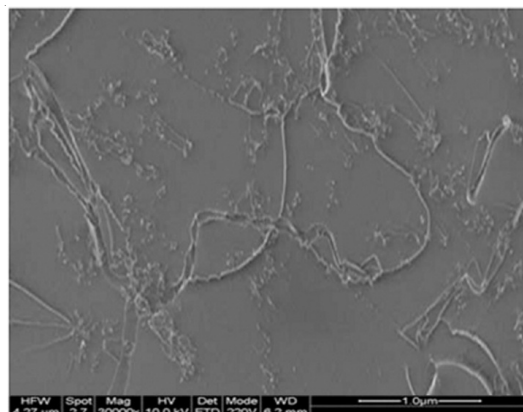


Fig. 7. (c) SEM of MWCNT-COOH/EP

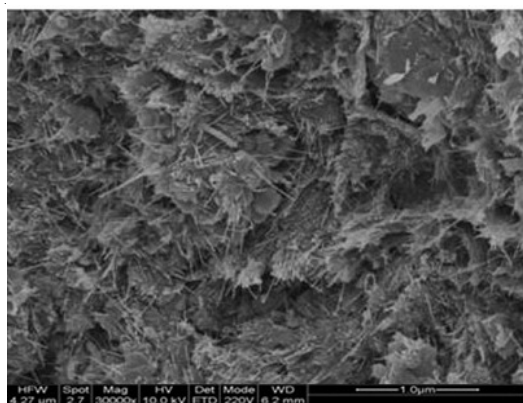


Fig. 7. (d) SEM of CS-g-MWCNTs/EP

the well dispersion of nanoparticles in epoxy matrix²¹. CS-g-MWCNTs filled epoxy bionanocomposites shows the reinforcement of CS-g-MWCNTs flakes in epoxy matrix.

Conclusion

The chitosan was successfully functionalized onto MWCNTs a surface which was confirmed by FT-IR and TEM. Increase in mechanical and thermal properties of CS-g-MWCNTs/EP bionanocomposites is due to complicated reaction between free amino groups of CS-g-MWCNTs with epoxy matrix. The fracture surfaces of CS-g-MWCNTs/EP bionanocomposites indicate the dispersion of CS-g-MWCNTs onto epoxy matrix. This bionanocomposites can be used for the encapsulation of electronic devices such as organic light emitting diodes and organic photovoltaics which are susceptible to chemical degradation in the presence of oxygen and moisture. Further, CS-g-MWCNTs/EP bionanocomposites are degradable and environment friendly.

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