

## A New Dihydroxanthone from the Fermentation Products of Endophytic Fungus of *Phomopsis amygdali*

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A new dihydroxanthone, (1R,2R)-2-hydroxy-6-(hydroxymethyl)-8-methoxy-9-oxo-2,9-dihydro-1H-xanthene-1-carboxylic acid (**1**) was isolated from the fermentation products of a *Phomopsis* fungus. Its structure was elucidated by spectroscopic methods. Compound **1** was tested for its cytotoxicity and it showed high cytotoxicity against NB4 and PC3 cell with IC<sub>50</sub> values of 4.5 and 3.8 μM.

**Keywords:** Dihydroxanthone, *Phomopsis* fungus, Fermentation products, Cytotoxicity.

### INTRODUCTION

The *Phomopsis* species known as an important phytopathogenic genus contains more than 900 species named from a wide range of hosts<sup>1</sup>. These microorganisms produce a number of secondary metabolites with various biological activities, including antimicrobial<sup>2,3</sup>, antifungal<sup>4,5</sup>, antimalarial<sup>6,7</sup>, antitumor<sup>7,8</sup>, etc. The xanthone derivatives are important metabolites isolated from the *Phomopsis* genus and they appeals to medicinal chemists because of their pronounced pharmacological effects<sup>9,10</sup>.

With the aim of multipurpose utilization endophytic fungus of isolated from the rhizome of *Paris polyphylla* var. *yunnanensis* and identify bioactive natural products, the phytochemical investigation on fermentation products of the endophytic fungus *Phomopsis amygdali*. was carried out. As a result, a new dihydroxanthone (**1**) (Fig. 1) was isolated. In addition, the cytotoxicity of compound **1** was evaluated. This article reported the isolation, structure elucidation and cytotoxicity of the new compound.

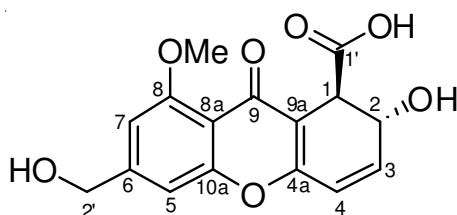


Fig. 1. Structure of **1**

### EXPERIMENTAL

Optical rotations were measured with a Horiba SEPA-300 polarimeter. UV spectra were obtained using a Shimadzu UV-2401A spectrophotometer. IR spectra were obtained in KBr disc on a Bio-Rad Wininfrared spectrophotometer. ESI-MS were measured on a VG Auto Spec-3000 MS spectrometer. <sup>1</sup>H, <sup>13</sup>C and 2D NMR spectra were recorded on Bruker DRX-500 instrument with TMS as internal standard. Column chromatography was performed on silica gel (200-300 mesh), or on silica gel H (10-40 mm, Qingdao Marine Chemical Inc., China). Second separate was used an Agilent 1100 HPLC equipped with ZORBAX-C18 (21.2 mm × 250 mm, 7 mm) column and DAD detector.

The culture of *Phomopsis* sp. was isolated from the rhizome of *Paris polyphylla* var. *yunnanensis* collected from Shizhong, Yunnan, People's Republic of China, in 2011. The strain was identified by Gang Du based on the analysis of the ITS sequence. It was cultivated at room temperature for 7 days on potato dextrose agar at 28 °C. Agar plugs were inoculated into 250 mL Erlenmeyer flasks each containing 100 mL potato dextrose broth and cultured at 28 °C on a rotary shaker at 180 rpm for 5 days. Large scale fermentation was carried out in 50 Fernbach flasks (500 mL) each containing 100 g of rice and 120 mL of distilled H<sub>2</sub>O. Each flask was inoculated with 5 mL of cultured broth and incubated at 25 °C for 45 days.

**Extraction and isolation:** The fermentation products were extracted four times with 70 % acetone (4 × 5 L) at room temperature and filtered. The crude extract (42.5 g) was applied

TABLE-1  
<sup>1</sup>H- AND <sup>13</sup>C-NMR DATA OF COMPOUND **1** (500 AND 125 MHz, IN CD<sub>3</sub>OD)

No.	δ <sub>c</sub> (m)	δ <sub>H</sub> (m, J, Hz)	No.	δ <sub>c</sub> (m)	δ <sub>H</sub> (m, J, Hz)
1	47.5 d	4.22 d (3.8)	9	183.2 s	
2	65.2 d	4.68 dd (3.8, 4.8)	4a	158.4 s	
3	137.6 d	6.45 d (4.8, 9.9)	8a	110.9 s	
4	123.8 d	6.68 dd (9.9)	9a	112.2 s	
5	106.2 d	6.95 s	10a	156.4 s	
6	149.5 s		1'	176.4 s	10.24 brs
7	108.5 d	6.74 s	2'	65.1 t	4.65 s
8	162.8 s		8-OMe	55.9 q	3.81 s

to silica gel (200-300 mesh) column chromatography, eluting with a CHCl<sub>3</sub>-acetone gradient system (20:1, 9:1, 8:2, 7:3, 6:4, 5:5), to give six fractions A-F. The further separation of fraction D (7:3, 6.53 g) by silica gel column chromatography and preparative HPLC (32 % MeOH, flow rate 12 mL/min) to give **1** (5.4 mg).

**(1R,2R)-2-hydroxy-6-(hydroxymethyl)-8-methoxy-9-oxo-2,9-dihydro-1H-xanthene-1-carboxylic acid (1)**: yellow gum; [α]<sub>D</sub><sup>24.5</sup> -8.21 (c 0.20, MeOH); UV (CH<sub>3</sub>OH), λ<sub>max</sub> (log ε) 340 (3.82), 272 (4.14), 210 (4.38) nm; IR (KBr, ν<sub>max</sub>, cm<sup>-1</sup>): 3420, 2945, 2850, 1752, 1654, 1602, 1538, 1455, 1416, 1274, 1158, 1072, 966, 857; <sup>13</sup>C NMR and <sup>1</sup>H NMR data (500 and 125 MHz, in CD<sub>3</sub>OD) see Table-1; positive ESIMS *m/z* 341 [M + Na]<sup>+</sup>; positive HRESIMS *m/z* 341.0632 [M + Na]<sup>+</sup> (calcd for C<sub>16</sub>H<sub>14</sub>NaO<sub>7</sub>, 341.0637).

## RESULTS AND DISCUSSION

A 70 % aq. acetone extract prepared from fermentation products of the endophytic fungus *Phomopsis amygdali* was subjected repeatedly to column chromatography on Silica gel, Sephadex LH-20, RP-18 and Preparative HPLC to afford compound **1**. The structure of **1** was shown in Fig. 1 and its <sup>1</sup>H and <sup>13</sup>C NMR data were listed in Table-1.

**Compound 1 was obtained as a yellow powder**: The molecular formula was established as C<sub>16</sub>H<sub>14</sub>NaO<sub>7</sub> *m/z* 341.0632 [M + Na]<sup>+</sup> (calcd. for 341.0637), which indicated 10 degrees of unsaturation. Its UV spectrum showed the maximum absorption at 340, 272 and 210 nm. Strong absorption bands accounting for hydroxy (3420 cm<sup>-1</sup>), carbonyl (1752, 1654 cm<sup>-1</sup>) and aromatic groups (1602, 1538, 1455 cm<sup>-1</sup>) could also be observed in its IR spectrum. The <sup>13</sup>C and DEPT spectra of **1** revealed 16 carbon, composed of one methoxy group (δ<sub>c</sub> 55.9), one methine group (δ<sub>c</sub> 47.5), one oxidized methine group (δ<sub>c</sub> 65.2), one oxidized methylene group (δ<sub>c</sub> 65.1), four aromatic methines (δ<sub>c</sub> 137.6, 123.8, 106.2 and 108.5), six aromatic quatern (δ<sub>c</sub> 149.5, 162.8, 158.4, 110.9, 112.2 and 156.4) and two carbonyl carbons (δ<sub>c</sub> 183.2 and 176.4). The <sup>13</sup>C NMR signal 176.4 was assigned to a carbonyl group. The <sup>1</sup>H NMR spectrum revealed four aromatic protons [δ<sub>H</sub> (6.74 s), (6.95 s), (6.45 d, *J* = 9.9, 4.8 Hz) and (6.68 dd, *J* = 9.9 Hz)]. Two aromatic protons at δ<sub>H</sub> 6.45 (d, *J* = 9.9, 4.8 Hz) and 6.68 (dd, *J* = 9.9 Hz) were connected to those at δ<sub>H</sub> 4.68 (dd *J* = 3.8, 4.8 Hz) and 4.22 (d, *J* = 3.8 Hz).

This connectivity was confirmed by correlation of these four protons <sup>1</sup>H-<sup>1</sup>H COSY spectrum (Fig. 2). In addition, one methoxy and one hydroxymethyl protons appeared at δ<sub>H</sub> 3.81

(s) and 4.65 (s), respectively. The precise connectivities between proton and carbon signals were established by interpretation of HMBC data. In HMBC spectrum of **1**, the proton at δ<sub>H</sub> 4.22 (H-1) was correlated with C-2 (δ<sub>c</sub> 65.2), C-3 (δ<sub>c</sub> 137.6), C-4a (δ<sub>c</sub> 158.4), C-9 (δ<sub>c</sub> 183.2), C-9a (δ<sub>c</sub> 112.2). These correlations indicated the existence of dihydroxanthene skeleton<sup>11</sup>. The HMBC correlations (Fig. 2) of the methoxy proton signal (δ<sub>H</sub> 3.81) with C-8 (δ<sub>c</sub> 162.8) suggested the methoxy group should be located at C-8. The hydroxymethyl group located at C-6 was supported by the HMBC correlations of H-2' (δ<sub>H</sub> 4.65) with C-5 (δ<sub>c</sub> 106.2), C-6 (δ<sub>c</sub> 149.5) and C-7 (δ<sub>c</sub> 108.5). Furthermore, a hydroxy group should be located at C-2 to support the existence of an oxidized methylene. The configurations of 1R, 2R in **1** were deduced from the comparison of <sup>1</sup>H and <sup>13</sup>C NMR data, coupling constants and ROESY correlations with these of AGI-B4,<sup>11</sup> of which configuration was unambiguously established by an X-ray structure analysis. According to above informations, compound **1** was assigned as (1R, 2R)-2-hydroxy-6-(hydroxymethyl)-8-methoxy-9-oxo-2,9-dihydro-1H-xanthene-1-carboxylic acid.

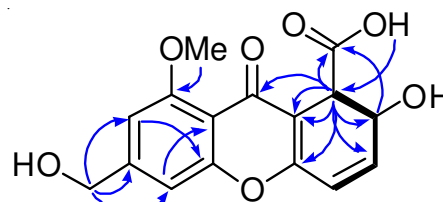


Fig. 2. Key HMBC (—) and <sup>1</sup>H-<sup>1</sup>H COSY (—) correlations of **1**

Xanthenes are known to exhibit cytotoxic effects<sup>6,9,12</sup>. Compound **1** were tested for its cytotoxicity against five tumor cells line (NB4, A549, SHSY5Y, PC3 and MCF7) using a previously reported procedure<sup>13</sup>. The results showed that **1** exhibited high cytotoxicity against NB4 and PC3 cell with IC<sub>50</sub> values of 4.5 and 3.8 μM.

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## REFERENCES

1. D. Udayanga, X. Liu, E.H.C. McKenzie, E. Chukeatirote, A.H.A. Bahkali and K.D. Hyde, *Fungal Divers.*, **50**, 189 (2011).
2. J.N. Choi, J. Kim, K. Ponnusamy, C. Lim, J.G. Kim, M.J. Muthaiya and C.H. Lee, *J. Antibiot.*, **66**, 231 (2013).
3. H. Hussain, M.K. Tchimene, I.K. Ahmed, M. Steinert, S. Draeger, B. Schulz and K. Krohn, *Nat. Prod. Commun.*, **6**, 1905 (2011).
4. H. Hussain, I. Ahmed, B. Schulz, S. Draeger and K. Krohn, *Fitoterapia*, **83**, 523 (2012).
5. H. Hussain, K. Krohn, I. Ahmed, S. Draeger, B. Schulz, S. Di Pietro and G. Pescitelli, *Eur. J. Org. Chem.*, 1783 (2012).
6. M. Isaka, A. Jaturapat, K. Rukseree, K. Danwisetkanjana, M. Tanticharoen and Y. Thebtaranonth, *J. Nat. Prod.*, **64**, 1015 (2001).
7. C. Hemtasin, S. Kanokmedhakul, K. Kanokmedhakul, C. Hahnvanawong, K. Soyong, S. Prabpai and P. Kongsaree, *J. Nat. Prod.*, **74**, 609 (2011).
8. E. Adelin, C. Servy, S. Cortial, H. Lévaïque, M.-T. Martin, P. Retailleau, G. Le Goff, B. Bussaban, S. Lumyong and J. Ouazzani, *Phytochemistry*, **72**, 2406 (2011).
9. K.S. Masters and S. Brase, *Chem. Rev.*, **112**, 3717 (2012).
10. S. Kaul, S. Gupta, M. Ahmed and M.K. Dhar, *Phytochem. Rev.*, **11**, 487 (2012).
11. H.S. Kim, I.Y. Park, Y.J. Park, J.H. Lee, Y.S. Hong and J.J. Lee, *J. Antibiot.*, **55**, 669 (2002).
12. Q. Hu, X. Gao, D. Niu, X. Li, Y. Qin, Z. Yang, G. Zhao, Z. Yang and Z. Chen, *Heterocycles*, **87**, 1127 (2013).
13. X.M. Gao, R.R. Wang, D.Y. Niu, C.Y. Meng, L.M. Yang, Y.T. Zheng, G.Y. Yang, Q.F. Hu, H.D. Sun and W.L. Xiao, *J. Nat. Prod.*, **76**, 1052 (2013).