

Chemical Constituents of *n*-Butanol Extract of Capparis spinosa L.

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The aim of this study was to ascertain chemical constituents of *n*-butanol extract of *Capparis spinosa* L. fruits. The *n*-butanol extract of *Capparis spinosa* L. was effective at controlling S180 tumour growth in a murine model. Silica gel column chromatography, Sephadex LH-20 and re-crystallization methods were used to isolate and purify the *n*-butanol extract. Chemical structures were elucidated by spectroscopic analyses and compared with literature. As results, nine known compounds were isolated and identified as: *bis*-(5-fomylfurfuryl) ether (1), 5-(hydroxymethyl)furfural (2), uracil (3), adenosine (4), 2-hydroxy-6-methoxy-1*H*-indol-3-ethanone (5), rutin (6), 2-methyl-6-(1',2',3',4'-tetrahydroxybutyl)pyrazine (7), *p*-hydroxy-benzoic acid (8) and stachydrine (9). There in, the compound 7 was identified in *Capparis spinosa* L. for the first time.

Keywords: Capparis spinosa L., n-Butanol extract, Antitumor activity, Chemical constituents, Spectroscopic analyses.

INTRODUCTION

Capparis spinosa L. is a perennate shrub belonging to the family Capparidaceae. It is widely grown on dry regions in west or central Asia and Mediterranean basin¹. In China, *Capparis spinosa* L. is mainly distributed in the Xinjiang Autonomy Region. Different parts of this plant, such as the flowers buds, fruits, seeds, shoots and bark of roots, have been used as food or folk medicines. *Capparis spinosa* L. has various pharmacological activities and is used in phytomedicine around the world for its antifungal ability², antiinflammatory³ and antihepatotoxic effects⁴, *etc*.

In our previous studies, the *n*-butanol extract of *Capparis* spinosa L. was confirmed for its antitumor effect against SGC-7901 and HepG-2 cells *in vitro* test. The present study was carried out to answer the question whether *n*-butanol extract of *Capparis spinosa* L. is also capable of inhibiting tumors *in vivo* or not. Evaluation was made in the sarcoma 180 mice. Then other doubt arises: which compound in *n*-butanol extract of *Capparis spinosa* L takes on the antitumor effect? So we carry a study on chemical constituents of *n*-butanol extract of *Capparis spinosa* L. fruits to find the answer.

EXPERIMENTAL

The dry mature fruits of *Capparis spinosa* L. were collected from Xinjinag Urumqi and identified by Prof. Q.H. Wang

from institute for drug control of Hei longjiang, Harbin, China. Petroleum ether (60-90 °C), chloroform, ethyl acetate, methanol, *n*-butanol, acetone and all chemicals and reagents used were analytically pure. NMR spectra in DMSO-*d*₆ were recorded on a Bruker-600MHz (600 MHz for ¹H; 100 MHz, ¹³C). Mass spectra were obtained in a MATLCQ electro-spray mass spectrometry. Analytically pure solvents were used in all experiments.

Extraction: The fruits of *Capparis spinosa* L. were extracted with ethanol (95 %). The extracts were concentrated under vacuum. Then the resulting residue was dissolved in water and extracted with petroleum ether (60-90 °C), ethyl acetate, *n*-butanol, respectively⁵.

Assay of antitumor activity *in vivo*: Sarcoma 180 ascites tumor cells $(1 \times 10^7 \text{ cell/mL})$ were implanted subcutaneously into the right anterior armpit of the mice. After 24 h inoculation, *n*-butanol extract of *Capparis spinosa* L.was dissolved in physiological saline and administered by gavage for 7 d in mice transplanted with Sarcoma 180 tumor. Cyclophosphamide was used as a positive control. The negative control was administrated by gavage with physiological saline. On day 8, the mice were sacrificed. Tumors were extirpated and weighed. The inhibition ratio (%) was calculated by the following formula: inhibition ratio (%) = [(A-B)/A] × 100, where A is the tumor weight average of the negative control and B is that of the treated group. Data are presented as mean ± SD. The differences between experimental groups were compared by ANOVA followed by least significant difference (LSD) test. Differences are considered significant when $p \le 0.05$.

Isolation: The dried fruits of *Capparis spinosa* L (10 kg) were extracted with 95 % EtOH. Evaporation of the solvent under reduced pressure gave the dry extract, which was partitioned successively with light petroleum, ethyl acetate and *n*-butanol. The *n*-butanol extract (108 g) was then subjected to further separation using repeated normal-phase silica gel column, Sephadex LH-20, ODS silica gel and HPLC to give nine compounds (Compound 1-9).

RESULTS AND DISCUSSION

Antitumor activity *in vivo* of *n*-butanol extract of *Capparis spinosa* L.: The effect of *n*-butanol extract of *Capparis spinosa* L. on the antitumor activity in sarcoma 180 bearing rats is shown in Table-1. The mean tumor weights at high, middle and low dose in the treated groups were compared to those in the untreated control. *n*-butanol extract of *Capparis spinosa* L. treated tumors were significantly smaller compared to control tumors on 200, 400, 800 mg kg⁻¹ d⁻¹. It was demonstrated that *n*-butanol extract of *Capparis spinosa* L. could significantly restrict further growth of Sarcoma 180.

TABLE-1				
INHIBITION EFFECTS OF n-BUTANOL EXTRACT OF				
Capparis spinosa L. ON S180 TUMOR IN MICE ($\overline{x} \pm SD$)				
Groups	Dose	N	Tumor	Inhibition
	(mg kg ⁻¹ d ⁻¹)		weight (g)	rate (%)
Control	-	12	1.121±0.158	-
n-Butanol extract	200	12	0.906±0.131**	19.16
of Capparis	400	12	$0.754 \pm 0.116^{**}$	32.76
spinosa L.	800	12	$0.604 \pm 0.107^{**}$	46.14
CTX	20	12	0.493±0.092**	55.17
** 0 1 1.1	. 1 0.001			

**Compared with control p < 0.001.

Identification of compounds of *n***-butanol extract of** *Capparis spinosa* L.: As a result, 9 compounds have been isolated and identified from *n*-butanol extract of *Capparis spinosa* L. and described as following (Fig. 1):

Compound 1: *bis*-(5-Fomylfurfuryl) ether⁶, colourless needles, melting point: 113.5-115.5 °C. ¹H NMR (600 MHz, DMSO-*d*₆) δ : 9.57 (2H, s), 6.73 (2H, d, *J* = 3.5, H-3, 3'), 7.50 (2H, d, *J* = 3.5, H-4,4'), 4.57 (2H, s, H-6,6'). ¹³C NMR (100 MHz, DMSO) δ : 178.5 (CHO), 157.9(C, C-2, C-2'), 153.4(C, C-5, C-5'), 122.3(CH, C-4, C-4'), 112.1(CH, C-3, C-3'), 65.2 (CH₂, C, C-6, C-6').

Compound 2: 5-(Hydroxymethyl)furfural⁷, colourless oil. ¹H NMR (600 MHz, DMSO- d_6) δ : 9.54 (1H, s), 7.48(1H, d, *J* = 3.5 Hz, H-3), 6.59 (1H, d, *J* = 3.5 Hz, H-4), 4.50 (2H, s). ¹³C NMR (100 MHz, DMSO) δ : 178.1 (CHO), 161.2(C, C-5), 152.9 (C, C-2), 123.9 (CH, C-3), 111.2 (CH, C-4).

Compound 3: Uracil⁷, white powder. ¹H NMR (600 MHz, DMSO-*d*₆) δ : 7.38 (1H, d, *J* = 7.4 Hz, H-6), 5.37(1H, d, *J* = 7.4 Hz, H-5), 3.52 (1H, brs, 1,3-NH₃). ¹³C NMR (100 MHz, DMSO) δ : 164.4(C-4), 151.6(C-2), 142.2(C-6), 100.3(C-5).

Compound 4: Adenosine⁸, white solid. ¹H NMR (600 MHz, DMSO-*d*₆) δ : 8.34 (H, s, H-8), 8.13(1H, s, H-2), 7.33(2H, s, NH₂), 5.87(1H, d, *J* = 6.2 Hz, H-1'), 4.61(1H, d, *J* = 3.1 Hz,



H-2'), 4.14 (1H, d, J = 3.1 Hz, H-3'), 3.96(1H, d, J = 3.1 Hz, H-4'), 3.67 (1H, dt, J = 11.9, 4.0 Hz, H-5'a), 3.58 (1H, dd, J =11.9, 4.0 Hz, H-5'b), 5.49-5.37 (2H, m, OH -2',5'), 5.16(1H, d, J = 4.1 Hz, OH-3'). ¹³C NMR (100 MHz, DMSO) δ : 156.3 (C-6), 152.7(C-2), 149.8 (C-4), 140.3 (C-8), 119.6 (C-5), 88.3 (C-1'), 73.8 (C-2'), 70.9 (C-3'), 86.2 (C-4'), 61.9 (C-5').

Compound 5: 2-Hydroxy-6-methoxy-1*H*-indol-3ethanone⁹, yellow crystal, ¹H NMR (600 MHz, DMSO- d_6) δ : 7.85 (1H, d, J = 8.5 Hz, H-4), 7.00 (1H, dd, J = 8.5, 2.6 Hz, H-5), 7.11 (d, J = 2.6 Hz, H-7), 3.69 (3H, s, H-11), 2.61(3H, s, H-12). **Compound 6:** Rutin¹⁰, pale yellow crystal. ¹H NMR (600 MHz, DMSO- d_6) δ : 12.59 (1H, s, OH-5), 10.84(1H, s, OH-7), 9.67(1H, s, OH-3'), 9.17(1H, s, OH-4'), 6.19(1H, s, H-6), 6.39 (1H, d, J = 2.5 Hz, H-8), 7.54 (1H, s, H-2'), 6.84 (1H, d, J = 8.2 Hz, H-5'), 7.53 (1H, s, H-6'), 5.34 (1H, d, J = 6.9 Hz, H-2"), 4.33 (1H, brs, H-2"), 0.99 (3H, d, J = 6.1 Hz, H-6"), 3.70-3.07 (10H, m). ¹³C NMR (100 MHz, DMSO) δ : 177.4(C-4), 164.1(C-7), 161.2(C-5), 156.4(C-2), 133.3(C-3), 100.8(C-6), 98.7(C-8), 148.4(C-3'), 144.8(C-4'), 121.6(C-1'), 121.2(C-6'), 116.3(C-5'), 115.2(C-2'), 93.6(C-1"), 75.9(C-5"), 74.1(C-2"), 71.9(C-3"), 67.00(C-6"), 70.0(C-4"), 104.0(C-1""), 76.5(C-4""), 70.6(C-2""), 70.4(C-3""), 68.3(C-5""), 17.7(C-6"").

Compound 7: 2-Methyl-6-(1',2',3',4'-tetrahydroxybutyl)pyrazine¹¹, colourless needles.¹H NMR (600 MHz, DMSO d_6) δ : 8.41(1H, s, H-5), 8.59(1H, s, H-3), 2.46(3H, s, H-2), 5.29(1H, d, *J*=6.4Hz, OH), 4.37(1H, d, *J* = 5.7 Hz, OH), 4.63 (1H, br, OH), 4.35(1H, br, OH), 3.5-3.61(3H, m, H-2'-4'). ¹³C NMR (100 MHz, DMSO) δ : 155.6 (C-2), 141.9 (C-3), 140.4 (C-5), 71.3 (C-1'), 73.8 (C-2'), 71.3 (C-3'), 63.6 (C-4'), 21.0 (CH₃).

Compound 8: *p*-Hydroxy-benzoic acid¹², colourless needles. ¹H NMR (600 MHz, CD₃OD) δ : 7.87(2H, d, *J* = 7.87 Hz, H-2, 6), 6.82 (2H, d, *J* = 7.87 Hz, H-3,5). ¹³C NMR (100 MHz, CD₃OD) δ : 122.8 (C-1), 130.0 (C-2), 116.0 (C-3), 163.3 (C-4), 116.0 (C-5), 133.0 (C-6), 170.3 (COOH).

Compound 9: Stachydrine, colourless oil. ¹H NMR (600 MHz, DMSO-*d*₆) δ: 3.77(1H, t, *J* = 9.6 Hz, H-2), 2.09, 2.17(1H, m, H-3), 1.92 (2H, m, H-4), 3.00(3H, s, H-1'), 3.22(3H, s, H-2'). ¹³C NMR (100 MHz, DMSO-*d*₆) δ: 76.0 (C-2), 24.5(C-3), 18.4(C-4), 65.7(C-5), 44.7(C-1'), 51.3(C-2'), 166.2(COOH).

Conclusion

The antitumor activity of *n*-butanol extract of *Capparis* spinosa L. was confirmed and nine compounds were isolated from this extract. They are *bis*-(5-fomylfurfuryl)ether (1),

5-(hydroxymethyl)furfural (**2**), uracil (**3**), adenosine (**4**), 2-hydroxy-6-methoxy-1*H*-indol-3-ethanone (**5**), rutin (**6**), 2-methyl-6-(1',2',3',4'-tetrahydroxybutyl)pyrazine (**7**), *p*-hydroxy-benzoic acid (**8**), stachydrine (**9**). More importantly, the compound (**7**) as a known alkaloid is extracted from *Capparis spinosa* L. for the first time.

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REFERENCES

- N. Tlili, W. Elfalleh, E. Saadaoui, A. Khaldi, S. Triki and N. Nasri, *Fitoterapia*, 82, 93 (2011).
- H. Wang, H. Wang, S. Shi, J. Duan and S. Wang, *Glycoconj. J.*, 29, 379 (2012).
- D. Trombetta, F. Occhiuto, D. Perri, C. Puglia, N.A. Santagati, A. De Pasquale, A. Saija and F. Bonina, *Phytother Res.*, **19**, 29 (2005).
- 4. C. Gadgoli and S.H. Mishra, J. Ethnopharmacol., 66, 187 (1999).
- 5. W.L. Li, J. Bai and Q.C. Dai, Asian J. Chem., 25, 8253 (2013).
- X. Li, J.H. Wang, D.Z. Meng and X. Li, J. Shenyang Pharm. Univ., 20, 173 (2003).
- J.C. Zhang, Y. Shen, G.Y. Zhu and M.S. Yang, J. Hebei Univ. (Nat. Sci. Edit.), 27, 262 (2007).
- 8. W. Jiang, W.H. Lin and S.D. Guo, J. Harbin Univ. Comm. (Nat. Sci. Edit.), 6, 684 (2005).
- 9. Y. Ban, Y. Sekine and T. Oishi, *Tetrahedron Lett.*, 19, 151 (1978).
- 10. Z.Z. Song and Z.J. Jia, J. Lanzhou Univ. (Nat. Sci. Ed.), 8, 99 (1992).
- 11. R. Wang, Y.S. Wen, L. Yang and W.J. Qin, *J. Chin. Mater. Med.*, **7**, 421 (1997).
- 12. J. Li, Y. Jiang and P.F. Tu, J. Chin. Mater. Med., 31, 45 (2006).