

Biological Activities of Herbaceous Peony Flower Extracts

YING-SHAN JIN¹, YONG-HAO XUAN², YIN-ZHE JIN³, MAN-LI CHEN¹ and JUN TAO^{4,*}

¹College of Bioscience and Biotechnology, Yangzhou University, Yangzhou 225009, Jiangsu Province, P.R. China
 ²Yangzhou University Library, Yangzhou University, Yangzhou 225009, Jiangsu Province, P.R. China
 ³College of Food Science and Technology, Shanghai Ocean University, Shanghai 201306, P.R. China
 ⁴College of Horticulture and Plant Protection, Yangzhou University, Yangzhou 225009, Jiangsu Province, P.R. China

*Corresponding author: E-mail: taojun@yzu.edu.cn

(Received: 29 March 2012;

Accepted: 14 January 2013)

AJC-12705

Herbaceous peony (*Paeonia lactiflora* Pall.) root is an important Traditional Chinese Medicine. To determine if the flower has any beneficial properties, we investigated antioxidant, antiinflammatory and anti α -glucosidase activities of petroleum ether, ethyl ether, ethyl acetate, *n*-butanol and water extracts of herbaceous peony flower. Antioxidant activity was evaluated by measuring total phenolic content, total antioxidant activity and DPPH radical scavenging activities. The ethyl ether and ethyl acetate extracts exhibited higher antioxidant ability than the other extracts. Moreover, the ethyl ether and ethyl acetate extracts had greater antiinflammatory activity on murine macrophage cells (RAW264.7). On the other hand, the petroleum ether extract and ethyl acetate extract showed strong inhibitory effect on α -glucosidase. These results suggest that flower from Herbaceous peony may have health-enhancing effects.

Key Words: Paeonia lactiflora Pall. Flower, Antioxidant activity, Anti α-glucosidase, Antiinflammatory.

INTRODUCTION

Herbaceous peony (*Paeonia lactiflora* Pall.) root is an important Traditional Chinese Medicine (TCM). A decoction of the root has been used in the treatment of rheumatoid arthritis, systemic lupus erythematosus, hepatitis, dysmenorrhea, muscle cramping and spasms and fever with a long history. Herbaceous peony is rich in different kinds of bioactive components, such as monoterpenes¹⁻³, triterpenes^{4.5}, flavonoids^{5.6}, stilbenes⁷, volatile oil⁵ and polyphenols⁸⁻¹⁰, among which paeoniflorin a kind of monoterpene is the most studied medicinal compound^{2,11-13}. During the cultivation of medicinal herbaceous peony, above ground biomass is often discarded as litter, resulting in enormous waste of resources.

In recent years, much attention has been focused on the biological properties of natural herbs and foods^{14,15}. Many important bioactivities (such as antioxidant, antiinflammatory, antidiabetes, anticancer) have been investigated in plant. Biological activities are correlated to the presence of certain compounds that may assist in predicting some traditional uses of medicinal plants¹⁶. Thus, we can assume that some plants can be less toxic to humans and can be used as medicine for the treatment of diseases. Previous studies of herbaceous peony were mainly focused on the root^{2,10,12,17}, with monoterpenes as the main compounds. There were few reports about the above ground parts of herbaceous peony.

In this study, we evaluated the possible biological activities such as antioxidant, anti α -glucosidase, antiinflammatory activities of flowers from herbaceous peony.

EXPERIMENTAL

Plant material and preparation of extracts: Herbaceous peony flower were collected from Yangzhou and identified by Professor Jun Tao (College of Horticulture and Plant Protection, Yangzhou University, China). Herbaceous peony flower was dried in the shade at room temperature and a 200 g sample of the powder was subjected to a methanol extraction process for 3 d. The extract was filtered through filter paper (100 mm; Whatman, Maidstone, UK) and evaporated using a vacuum rotary evaporator (CCA-1110; Eyela, Tokyo, Japan) to produce a crude extract of 108.7 g. The crude extract was suspended in 500 mL deionized water and partitioned sequentially with 500 mL of petroleum ether, 500 mL of ethyl ether, 500 mL of ethyl acetate and 500 mL of n-butanol. The solvent of petroleum ether fraction, ethyl ether fraction, ethyl acetate fraction and *n*-butanol fraction was then evaporated using a vacuum rotary evaporator. The final yields of petroleum ether fraction (PEF), ethyl ether fraction (EEF), ethyl acetate fraction (EAF), n-butanol fraction (BF) and water fraction (WF) were 6.92, 22.28, 21.12, 12.18 and 35.9 %, respectively (Table-1).

TABLE-1				
YIELD, TOTAL PHENOLIC AND DPPH FREE				
RADICAL SCAVENGING ACTIVITY OF EXTRACTS				
FROM Paeonia lactiflora PALL. FLOWERS				
Extracts	Yield (%)	Total phenolic	DPPH radical	
		content (tannic	scavenging activity ²	
		$mg/g)^1$	(IC ₅₀ : µg/mL)	
PEF	6.92	81.26	42.44	
EEF	22.28	42.59	7.31	
EAF	21.12	33.70	4.70	
BF	12.18	2.97	44.89	
WF	35.9	1.19	273.12	
Vitamin C	_	_	1.09	
(nositive control)				

^(positive control) ¹Total phenolic content is expressed as tannic equivalent mg/g. ²Amount required for 50 % reduction of DPPH (0.1 mM) after 0.5 h.

Determination of total phenolic contents: Each extract (1 mL, 1 mg/mL) was mixed with 9 mL, reagent solution (0.1 N Folin-Ciocalteu reagent and 7.5 % sodium carbonate). The reaction mixture was incubated at room temperature for 0.5 h against a blank. The total phenolic content was determined with respect to the standard curve of tannic acid and was expressed as tannic acid equivalent¹⁸.

Total antioxidant activity: The effects of herbaceous peony flower extracts on total antioxidant were investigated according to the method of Prieto *et al.*¹⁹, with several modifications. Briefly, 0.2 mL of each extract (at concentration of 12.5 μ g/mL) was mixed with 0.6 mL of the reagent solution (0.6 M sulphuric acid, 28 mM sodium phosphate and 4 mM ammonium molybdate). The reaction mixture was incubated at 95 °C for 90 min. The absorbance of the cooled mixture was measured at 695 nm against a blank. The total antioxidant activity was expressed as the absorbance of the sample. The higher absorbance value indicates higher antioxidant activity. Paeoniflorin and BHT were used as standards.

DPPH radical scavenging activity: The free radical scavenging activities of the extracts, based on the scavenging activity of the stable 1,1-diphenyl-2-picrylhydrazyl (DPPH) free radical, were determined by Kilani *et al.*²⁰, with some modifications. Aliquots (0.5 mL) of varying concentration of the extract were mixed with freshly prepared DPPH in methanol (final concentration 250 μ M) and the absorbance at 517 nm was determined after incubation for 0.5 h in the dark at room temperature. Distilled water was used as control and ascorbic acid as reference compound. Each dilution was assayed in triplicate, the three readings were averaged and the percentage inhibition of activity was calculated as: (1-A_{sample}/A_{control}) × 100 %.

Protection from protein damage: The effect of different fractions on protein oxidation was carried out according to the method of Hu *et al.*²¹ with some modifications. One hundred microliters of 4 mg/mL bovine serum albumin (BSA) was mixed with 100 μ L of H₂O or 1 mg/mL different extracts. The mixture was incubated at 37 °C for 3 h with 300 μ L reagent solution (50 μ M FeCl₃, 1 mM H₂O₂ and 100 μ M ascorbic acid). Bovine serum albumin without Fenton's reagent was used as the positive control. After incubation, the mixture was mixed with loading buffer and further incubated at 95 °C for 10 min. After treatment, protein samples were subjected to SDS-polyacrylamide gel electrophoresis (SDS-PAGE) to quantify

protein damage. The protein sample was loaded in a 12 % polyacrylamide gel and electrophoresed. After running for 2 h, gels were stained with 0.15 % Coomassie brilliant blue R-250 for 12 h, washed for 24 h. Densitometric analysis was done with image analysis software (Quantity one; Bio-Rad, Hercules, CA).

Rat intestinal α -glucosidase inhibitory activity: α -Glucosidase inhibitory activity was assayed by Kim *et al.*²² with some modifications. α -Glucosidase (0.075 unit) was mixed with various concentrations (0.01-200 µg/mL) of herbaceous peony flower extracts in phosphate buffer. After 15 min preincubation at 37 °C, 100 µL of 3 mM *p*-nitrophenyl glucopyranoside (*p*NPG) was added to the mixtures as substrate. The reactions were incubated at 37 °C for 10 min and stopped by adding 750 µL of 0.1 M Na₂CO₃. α -Glucosidase activity was determined by measuring the *p*-nitrophenol released from the *p*-nitrophenyl glucopyranoside at 405 nm.

Cell lines and cell culture: Mutine macrophage RAW264.7 cell line were purchased from the Korean Cell Bank (Seoul, Korea) and grown in Roswell Park Memorial Institute medium 1640 (RPMI 1640), supplemented with 10 % fetal bovine serum, 100 U/mL penicillin and 100 μ g/mL streptomycin. Cells were cultured in a humidified atmosphere and incubated at 37 °C in 5 % CO₂.

Nitric oxide assay: After pre-incubation of RAW264.7 cells $(2 \times 10^6 \text{ cells/mL})$ with LPS $(1 \ \mu\text{g/mL})$ for 24 h, the quantity of nitrite in the culture medium was measured as an indicator of NO production. Aliquots of 100 μ L cell culture medium were mixed with 50 μ L of 1 % sulfanilamide (in 5 % phosphoric acid) and 50 μ L of 0.1 % naphthyl-ethylenediamine dihydrochloride. Subsequently, the mixture was incubated at room temperature for 10 min and the absorbance was measured at 550 nm.

MTT assay for measurement of cell proliferation: The cytotoxicity of herbaceous peony flower extracts was determined by a conventional MTT assay, as previously reported²³.

Statistical analyses: All tests were carried out independently in triplicate (n = 3). Data are expressed as the mean \pm standard derivation (SD). The results were processed using Excel 2003 (Microsoft, Redmond, WA, USA).

RESULTS AND DISCUSSION

Total phenolic contents: Antioxidant compounds can reduce the risk of many diseases, such as diabetes, inflammation, cancer, several degenerative diseases and $aging^{24}$. Carotenoids, ascorbic acid, flavonoids and phenolic acids might be some of the components able to contribute to their antioxidant activity²⁵. The total phenolic contents of herbaceous peony flower extracts are shown in Table-1. The order of total phenolic contents was: PEF EEF >> EAF > BF > WF. EEF and EAF exhibited higher phnolic contents (42.59 and 33.70 mg/g) than BF and WF (2.97 and 1.19 mg/g). These results imply that herbaceous peony flower extracts EEF and EAF contain considerable amounts of phenolics.

Total antioxidant activity: High absorbance value of a sample indicates high antioxidant activity (Fig. 1). All sample exhibited varying degrees of antioxidant activity. The order of total antioxidant activity was: Paeoniflorin > EAF > EEF >



(12.5 µg/mL). Paeoniflorin and BHA were used as standards

BHA > BF > PEF > WF. At 12.5 μ g/mL, Paeoniflorin, EAF and EEF showed higher total antioxidant activity than BHA.

DPPH radical scavenging activity: It was reported that DPPH is the method of choice for evaluating the free radical scavenging activity of natural compounds²⁶. The DPPH radical scavenging activities of different extracts are shown in Table-1. The order of DPPH radical scavenging activity was: EAF > EEF > PEF > BF > WF. The DPPH scavenging activities of the extracts, expressed as an IC₅₀ value, ranged from 4.70-273.12 µg/mL. The EAF exhibited the strongest antioxidant activity (IC₅₀ value of 4.70 µg/mL), followed by the EEF (IC₅₀ value of 7.31 µg/mL) and the PEF and BF (IC₅₀ value of 42.44 and 44.89 µg/mL) which showed the weakest activity. These results suggested that the EAF and EEF contained the strongest free radical scavenging compounds.

Protection against protein damage: The oxidative protein damage induced by free radicals has been shown to play a significant role in aging and in several pathological events²⁷. Hydroxyl radical is recognized as a protein-damaging agent with physiological significance. In the present study, protein damage was induced by the $Fe^{3+}/H_2O_2/ascorbic$ acid system. The density of the bovine serum albumin band decreased to than that of the control after incubation with the Fenton's reagent (Fig. 2). The herbaceous peony flower extracts restored the bovine serum albumin band intensity, EAF and EEF showed greater protein protection than WF, BF and PEF.



Fig. 2. (A) SDS-PAGE profile of the BSA protein treated with Fe³⁺/H₂O₂/ ascorbic acid system in the presence of the fractions of herbaceous peony flower (1 mg/mL). (B) Histogram showing the protective effect densitometric measurements

Antiinflammatoty activity: Inflammation is a complex biological response to pathogens and damaged cells. However, chronic and uncontrolled inflammation may serve as an important and common pattern in various diseases²⁸. Therapy of inflammatory diseases is usually directed at the inflammatory processes. Many antiinflammatory drugs have been prepared and marketed²⁹; however, these complex drugs are known to provoke gastrointestinal irritation. Thus, more gentle antiinflammatory natural herbs are being investigated. In present study, the antiinflammatory activity of herbaceous peony flower extracts was measured on lipopolysaccharide (LPS)induced RAW264.7 cells. LPS can activate immune cells to upregulate inflammatory states. The level of nitric oxide (NO) produced is an important indicator of the inflammatory process. The overproduction of NO can create cytotoxicity and tissue damage in an organism³⁰. Compared to the group with LPS treatment, the herbaceous peony flower EAF and EEF reduced NO product (Fig. 3A). LPS can induce apoptosis in the cells, with a viability of 61.99 % (Fig. 3B). Herbaceous peony flower extracts protected the cells against LPS-induced apoptosis. The antiinflammatory effect of herbaceous peony flower EAF and EEF is considerable, the results suggest that herbaceous peony flower EAF and EEF is suitable to be used as an antiinflammatory agent. The molecular mechanism of the antiinflammatory effect elicited by herbaceous peony flower will be investigated in further study.



Fig. 3. Effect of herbaceous peony flower extracts on production of NO in LPS-activated RAW264.7 cells. (A) RAW264.7 cells (1×10^6 cells/mL) were incubated with herbaceous peony flower extracts in the presence of LPS (1 µg/mL) for 24 h. Culture supernatants were assayed for NO determination by Griess assay. (B) RAW264.7 cells (1×10^6 cells/mL) were incubated with herbaceous peony flower extracts for 24 h. Cell viability was determined by MTT assay. *: p < 0.05 and **:p < 0.01 compared to c©ontrol

 α -Glucosidase inhibition: Inhibitors of intestinal α -glucosidase are used in the treatment of non-insulin-dependent diabetes mellitus (NIDDM) and represent a huge proportion of the antidiabetic drug market³¹. It has been reported that the inhibition of α -glucosidase reduces the bioavailability of glucose, the determination of 40 drugs that inhibit carbohydrate hydrolyzing enzymes have been proved to decrease postprandial hyperglycemia and improve impaired glucose metabolism without promoting insulin secretion in NIDDM patients. To determine if herbaceous peony flower extracts possess antidiabetic properties, we studied the effect of these extracts in α -glucosidase inhibition assays. All the extracts demonstrated α -glucosidase inhibitory activity (Table-2). Based on the calculated IC₅₀ values, PEF ($21.98 \pm 0.55 \,\mu g/mL$) and EAF (33.84 \pm 0.78 µg/mL) showed the highest activity, followed by EEF (141.46 \pm 1.23 µg/mL), BF (213.12 \pm 2.11 μ g/mL) and WF (244.77 ± 2.98 μ g/mL). The extracts of herbaceous peony flower had α -glucosidase inhibitory activity, showing that they contain effective α -glucosidase inhibitors that may have potential antidiabetic activity.

TABLE-2 EFFECTS OF DIFFERENT EXTRACTS OF HERBACEOUS PEONY ON RAT INTESTINAL & GLUCOSIDASE INHUBITION ASSAY			
Sample	α -Glucosidase inhibitory activity (IC ₅₀ ^a µg/mL)		
PEF	21.98 ± 0.55 ^b		
EEF	141.46 ± 1.23		
EAF	33.84 ± 0.78		
BF	213.12 ± 2.11		
WF	244.77 ± 2.98		
Acarbose	0.003 ± 0.01		
^a Amount required for 50 % inhibition. ^b Values represent the mean ±			
SD(n = 3).			

Conclusion

In this study, the extracts from herbaceous peony flower showed different biological activities. The herbaceous peony flower extracts showed higher phenolic, free radical scavenging, antiinflammatory and anti α -glucosidase activities. These results suggest that herbaceous peony flower be investigated for further research of its antioxidant and antidiabetes activity. The mechanisms of antiinflammatory effect elicited by ethyl ether fraction and ethyl acetate fraction will be explored in further studies.

ACKNOWLEDGEMENTS

This work was financially supported by Agricultural Science & Technology Independent Innovation Fund of Jiangsu Province CX(12)2019, P.R. China.

REFERENCES

- A. Braca, P.V. Kiem, P.H. Yen, N.X. Nhiem, T.H. Quang, N.X. Cuong and C.V. Minh, *Fitoterapia*, **79**, 117 (2008).
- N. Kim, K.R. Park, I.S. Park and Y.H. Park, *Food Chem.*, **96**, 496 (2006).
 H.B. Wang, W.F. Gu, W.J. Chu, S. Zhang, X.C. Tang and G.W. Qin, *J.*
- Nat. Prod., 72, 1321 (2009).
- 4. A. Ikuta, K. Kamiya, T. Satake and Y. Saiki, *Phytochemistry*, **38**, 1203 (1995).
- K. Kamiya, K. Yoshioka, Y. Saiki, A. Ikuta and T. Satake, *Phytochemistry*, 44, 141 (1997).
- N. Jia, Q.Y. Shu, L.S. Wang, H. Du, Y.J. Xu and Z.A. Liu, *Sci. Hortic.*, 117, 167 (2008).
- H.J. Kim, E.J. Chang, S.J. Bae, S.M. Shim, H.D. Park, C.H. Rhee, J.H. Park and S.W. Choi, *Arch. Pharm. Res.*, 25, 293 (2002).
- 8. N. Kumar and M.G. Motto, Phytochemistry, 25, 250 (1985).
- 9. D. Guo, G. Ye and H. Guo, *Fitoterapia*, **77**, 613 (2006).
- S.C. Lee, Y.S. Kwon, K.H. Son, H.P. Kim and M.Y. Heo, *Arch. Pharm. Res.*, 28, 775 (2005).
- 11. F. Chen, H.T. Lu and Y. Jiang, J. Chromatogr. A, 1040, 205 (2004).
- 12. B. Lee, Y.W. Shin, E.A. Bae, S.J. Han, J.S. Kim, S.S. Kang and D.H. Kim, *Arch. Pharm. Res.*, **31**, 445 (2008).
- 13. L. Xiao, Y.Z. Wang, J. Liu, X.T. Luo, Y. Ye and X.Z. Zhu, *Life Sci.*, **784**, 413 (2005).
- Y. Yang, T. Yu, H.J. Jang, S.E. Byeon, S.Y. Song, B.H. Lee, M.H. Rhee, T.W. Kim, J. Lee, S. Hong and J.Y. Cho, *J. Ethnopharmacol.*, **139**, 616 (2012).
- P. McCue, Y.I. Kwon and M.K. Shetty, *Asia Pacific J. Clin. Nutri.*, 14, 145 (2005).
- 16. T.L. Shale, W.A. Stirk and J. Staden, *J. Ethnopharmacol.*, **67**, 347 (1999).
- R.R. Baumgartner, D. Steinmann, E.H. Heiss, A.G. Atanasov, M. Ganzera, H. Stuppner and V.M. Dirsch, *Nat. Prod.*, **73**, 1578 (2010).
- 18. M.J. Jung, S.I. Heo and M.H. Wang, Food Chem., 108, 482 (2008).
- 19. P. Prieto, M. Pineda and M. Aguilar, Anal. Biochem., 269, 337 (1999).
- S. Kilani, R.B. Ammar, I. Bouhlel, A. Abdelwahed, N. Hayder, A. Mahmoud, K. Ghedira and L. Chekir-Ghedira, *Environ. Toxicol. Pharmacol.*, 20, 478 (2005).
- 21. W. Hu, T. Shen and M.H. Wang, J. Food Sci. Nutr., 14, 277 (2009).
- 22. Y.M. Kim, M.H. Wang and H.I. Rhee, Carbohydr. Res., 339, 715 (2004).
- J.Y. Cho, K.U. Baik, J.H. Jung and M.H. Park, *Eur. J. Pharmacol.*, 398, 399 (2000).
- 24. D.B. Choi, S.S. Park, J.L. Ding and W.S. Cha, *Biotechonol. Bioprocess Eng.*, **12**, 516 (2007).
- 25. D. Steinberg, J. Internal. Med., 233, 227 (1993).
- K. Shimada, K. Fujikawa, K. Yahara and T. Nakamura, J. Agric. Food Chem., 40, 945 (1997).
- 27. E.R. Stadtman and R.L. Levin, Ann. NY Acad. Sci., 899, 191 (2000).
- L. Ferrero-Miliani, O.H. Nielsen, P.S. Andersen and S.E. Girardin, *Clin. Exp. Immunol.*, **147**, 227 (2007).
- 29. P.O. Osadebe and F.B.C. Okoye, J. Ethnopharmacol., 89, 19 (2003).
- C. Nicholas, S. Batra, M.A. Vargo, O.H. Voss, M.A. Gavrilin, M.D. Wewers, D.C. Guttridge, E. Grotewold and A.I. Doseff, *J. Immunol.*, **179**, 7121 (2007).
- 31. S.E. Inzucchi, J. Am. Med. Assoc., 287, 360 (2002).