

Synthesis, Characterization and Biological Screening of *N*-Substituted (5-Chloro-2-methoxyphenyl)benzene Sulfonamide

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In the present study, a series of *N*-substituted (5-chloro-2-methoxyphenyl)benzene sulfonamide have been synthesized. The reaction of benzene sulfonyl chloride (1) with 2-amino-4-chloroanisole (2) yielded *N*-(5-chloro-2-methoxyphenyl)benzene sulfonamide (3). Finally the target compounds (**5a-k**) were obtained by stirring *N*-(5-chloro-2-methoxyphenyl)benzene sulfonamide with different electrophiles (**4a-k**) in the presence of *N*,*N*-dimethyl formamide and sodium hydride. The structures of the synthesized compounds were established by spectroscopic techniques like ¹H NMR and EI-MS. These compounds were assayed for their antioxidant activities by using 2,2-diphenyl-1-picrylhydrazil (DPPH) scavenging and other biological activities *via* screening them against acetylcholinesterase, butyrylcholinesterase and lipoxygenase enzymes, however, these showed prominent activity against acetylcholinesterase enzyme. It is clearly evident from the results that the compounds *N*-methyl-(5-chloro-2-methoxyphenyl)benzene sulfonamide (**5a**), *N*-allyl-*N*-(5-chloro-2-methoxyphenyl)benzene sulfonamide (**5b**) and *N*-2"-phenylethyl-*N*-(5-chloro-2-methoxyphenyl)benzene sulfonamide (**5j**) were found to be promising inhibitors against acetylcholinesterase enzyme having IC₅₀ value of 34.61 ± 0.62, 40.21 ± 0.25 and 45.11 ± 0.22 µmol/L, respectively, relative to Eserine, a reference standard with IC₅₀ value of 0.04 ± 0.001 µmol/L.

Key Words: 2-Amino-4-chloroanisole, Benzenesulfonyl chloride, Sulfonamides, Acetylcholinesterase enzyme.

INTRODUCTION

The sulfonamides belong to class of compounds that comprise at least five different classes of pharmacologically active agents¹. The basic sulfonamide group -SO₂NH- occurs in various biological dynamic compounds including antimicrobial drugs, antithyroid agents, antitumor, antibiotics and inhibitors of carbonic anhydrase². Sulfonamides are widely used to treat microbial infections by inhibiting the growth of gram negative and gram positive bacteria, some protozoa and fungi³. Clinically sulfonamides are used to treat several urinary tract infections and gastrointestinal infections⁴. Sulfonamides that are aromatic or hetroaromatic are responsible for the inhibition of the growth of tumor cells. They act as antitumor agents by inhibiting the carbonic anhydrase. Sulfonamides are structurally similar to p-aminobenzoic acid (PABA) which is a cofactor that is needed by the bacteria for the synthesis of folic acid. Sulfonamides antibiotics inhibit the conversion of *p*-aminobenzoic acid into folic acid and thus ultimately inhibit the synthesis of purine and DNA. Sulfonamide antibiotics are used as veterinary medicines to treat infections in livestock herds^{5,6}.

Acetyl cholinesterase (AChE, EC 3.1.1.7) and butyryl cholinesterase (BChE, EC 3.1.1.8) comprise a family of enzymes which include serine hydrolases. The different specificities for substrates and inhibitors for these enzymes are due to the differences in amino acid residues of the active sites of AChE and BChE. The enzyme system is responsible for the termination of acetylcholine at cholinergic synapses. These are key components of cholinergic brain synapses and neuromuscular junctions. The major function of AChE and BChE is to catalyze the hydrolysis of the neurotransmitter acetylcholine and termination of the nerve impulse in cholinergic synapses⁷. It has been found that BChE is present in significantly higher quantities in Alzheimer's plaques than in the normal age related non dementia of brains. H₁ and H₂ receptor antagonists possess AChE inhibitory activities. Cholinesterase inhibitors increase the amount of acetylcholine available for neuronal and neuromuscular transmission through their ability to reversibly or irreversibly. Hence, the search for new cholinesterase inhibitors is considered an important and ongoing strategy to introduce new drug candidates for the treatment of Alzheimer's disease and other related diseases⁸⁻¹⁰.

Literature survey revealed that slight modification in the structure of sulfonamide can lead to quantitative as well as qualitative changes in the biological activity. It prompted us to synthesize the various *N*-substituted derivatives of sulfonamides with an objective to search new contenders of drug having significant enhanced activity and can be helpful in controlling many degenerative diseases. For this, the parent sulfonamide N-(5-chloro-2-methoxyphenyl)benzene sulfonamide (**3**), was first prepared by reacting benzene sulfonyl chloride with 2-amino-4-chloroanisole at room temperature in basic medium. Simple stirring gave the desired compound in excellent yield. Then it was further processed to obtain different new *N*-alkyl substituted sulfonamides.

EXPERIMENTAL

¹H NMR spectra were recorded in CD₃OD on a Burker Aspect AM-500 MHz using TMS as an internal standard. Chemical shifts are given in ppm. IR spectra were recorded in KBr on a Shimadzu IR-460 and Burker vector 22 spectrometer (wave number in cm⁻¹). Mass spectra (EIMS) were measured on Finnigan MAT-112 instrument. High resolution EIMS were recorded in Jeol JMS HX-110 spectrometer. The melting points were recorded on a Griffin & George melting point apparatus. TLC was conducted on 0.25 mm silica gel plates. Visualization was made with ultraviolet light.

Procedure for the synthesis of N-(5-chloro-2-methoxyphenyl)benzene sulfonamide (3): The nucleophilic substitution reaction of amine with aryl sulfonyl chloride was carried out in aqueous media as described in the literature¹¹: a mixture 2-amino-4-chloroanisole (1; 10.0 mmol; 1.47 g) and benzene sulfonyl chloride (2; 10.0 mmol; 1.45 mL) and was suspended in 25 mL water. The pH of the suspension was maintained at 9.0 by adding aqueous solution of Na₂CO₃ at room temperature. The mixture was stirred and monitored by using analytical technique TLC for the completion of reaction. Then concentrated HCl was added gradually to adjust the pH to 2.0. The precipitates were collected by filtration, washed with distilled H₂O and dried to afford the title compound **3**. Methyl alcohol was used to dissolve the product and then it was re-crystallized by slow evaporation of the solvent, to generate colourless needle like crystals of N-(5-chloro-2-methoxyphenyl)benzene sulfonamide.

General method for the synthesis of compounds (5ak): The calculated amount of compound 3 (0.1 mmol) was taken in a round bottomed flask (50 mL) and *N*,*N*-dimethyl formamide (10 mL) was added to dissolve it followed by the addition of sodium hydride/lithium hydride (0.1 mmol) to the reaction mixture. The mixture was stirred for 0.5 h at ambient temperature and then slowly added the electrophile to the mixture and was further stirred for 2 h. The progress of reaction was monitored *via* TLC. The product was precipitated by adding cold distilled water. It was filtered, washed with water and crystallized from aqueous methanol.

Acetylcholinesterase assay: The AChE inhibition activity was performed according to the reported method¹² with slight modifications. Total volume of the reaction mixture was 100 μL. It contained 60 μL Na₂HPO₄ buffer with concentration of 50 mM and pH 7.7. Ten µL test compound (0.5 mM well⁻¹) was added, followed by the addition of 10 μ L (0.005 unit well⁻¹) enzyme. The contents were mixed and pre-read at 405 nm. Then contents were pre-incubated for 10 min at 37 °C. The reaction was initiated by the addition of 10 µL of 0.5 mM well⁻¹ substrate (acetylthiocholine iodide), followed by the addition of 10 µL DTNB (0.5 mM well-1). After 15 min of incubation at 37 °C absorbance was measured at 405 nm using 96-well plate reader Synergy HT, Biotek, USA. All experiments were carried out with their respective controls in triplicate. Eserine (0.5 mM well⁻¹) was used as a positive control. The percent inhibition was calculated by the help of following equation

Inhibition (%) =
$$\frac{\text{Control} - \text{Test}}{\text{Control}} \times 100$$

IC₅₀ values were calculated using EZ-Fit Enzyme kinetics software (Perrella Scientific Inc. Amherst, USA).

Butyrylcholinesterase assay: The BChE inhibition activity was performed according to the reported method¹² with slight modifications. Total volume of the reaction mixture was 100 µL containing 60 µL, Na₂HPO₄ buffer, 50 mM and pH 7.7. Ten µL test compound 0.5 mM well⁻¹ was added followed by the addition of 10 µL (0.5 unit well⁻¹) BChE (Sigma Inc.). The contents were mixed and pre-read at 405 nm and then pre-incubated for 10 min at 37 °C. The reaction was initiated by the addition of 10 µL of 0.5 mM well⁻¹ substrate (butyrylthiocholine chloride). Followed by the addition of 10 µL DTNB, 0.5 mM well⁻¹. After 15 min of incubation at 37 °C, absorbance was measured at 405 nm using 96-well plate reader Synergy HT, Biotek, USA. All experiments were carried out with their respective controls in triplicate. Eserine (0.5 mM well⁻¹) was used as positive control. The percent inhibition was calculated by the help of following equation.

Inhibition (%) =
$$\frac{\text{Control} - \text{Test}}{\text{Control}} \times 100$$

IC₅₀ values were calculated using EZ-Fit Enzyme kinetics software (Perrella Scientific Inc. Amherst, USA).

Lipoxygenase assay: Lipoxygenase (LOX) activity was assayed according to the reported method¹³⁻¹⁵ with slight modifications. A total volume of 200 µL lipoxygenase assay mixture contained 150 µL sodium phosphate buffer (100 mM, pH 8.0), 10 µL test compound and 15 µL purified lipoxygenase enzyme (600 units well⁻¹, Sigma Inc.). The contents were mixed and pre-read at 234 nm and preincubated for 10 min at 25 °C. The reaction was initiated by addition of 25 µL substrate solution. The change in absorbance was observed after 6 min at 234 nm using 96-well plate reader Synergy HT, Biotek, USA. All reactions were performed in triplicates. The positive and negative controls were included in the assay. Baicalin (0.5 mM well⁻¹) was used as a positive control. The percentage inhibition (%) was calculated by formula given below.

Inhibition (%) =
$$\frac{\text{Control} - \text{Test}}{\text{Control}} \times 100$$

where, Control = Total enzyme activity without inhibitor. Test = Activity in the presence of test compound. IC_{50} values were calculated using EZ-Fit Enzyme Kinetics software (Perrella Scientific Inc. Amherst, USA).

DPPH assay: The stable 1,1-diphenyl-2-picrylhydrazyl radical (DPPH) was used for the determination of antioxidant activity. Different concentrations of compounds in respective solvents were added at an equal volume (10 μ L) to 90 μ L of 100 μ M methanolic DPPH in a total volume of 100 μ L in 96-well plates. The contents were mixed and incubated at 37 °C for 0.5 h. The absorbance was measured at 517 nm using Synergy HT BioTek® USA microplate reader. Quercetin and L-ascorbic acid were used as standard antioxidants. The experiments were carried out in triplicates. IC₅₀ values were calculated using EZ-Fit5 Perrella Scientific Inc. Amherst USA software. The decrease in absorbance indicates increased radical scavenging activity which was determined by the following formula¹⁶.

Antiradical activity (Inhibition %) = $\frac{\text{Control} - \text{Test}}{\text{Control}} \times 100$

Statistical analysis: All the measurements were done in triplicate and statistical analysis was performed by Microsoft Excel 2003. Results are presented as mean \pm sem.

Spectral characterization of the synthesized compounds

N-(**5-Chloro-2-methoxyphenyl)benzene sulfonamide** (**3**): Colourless needle like crystals, Yield: 91 %, m.p. 140-142 °C. IR (KBr, v_{max} , cm⁻¹): 3120 (*N*-H str.), 3020 (C-H aromatic str.), 1447, 1338 (SO₂), 1225 (-OCH₃); ¹H NMR (500 MHz, CD₃OD): δ 7.71 (dd, *J* = 7.5, 1.5 Hz, 2H, H-2' and H-6'), 7.55 (br.t, *J* = 7.5 Hz, 1H, H-4'), 7.45 (br.t, *J* = 7.5 Hz, 2H, H-3' and H-5'), 7.41 (d, *J* = 2.5 Hz, 1H, H-6), 7.05 (dd, *J* = 8.5, 2.5 Hz, 1H, H-4), 6.79 (d, *J* = 8.5 Hz, 1H, H-3), 3.51 (s, 3H, -OCH₃); EIMS: m/z 297 [M⁺], 288 [M-CH₃]⁺, 266 [M-OCH₃]⁺, 233 [M-SO₂]⁺, 156 [M-PhSO₂]⁺, 141 [M-C₆H₃(Cl)(OCH₃NH)]⁺.

N-Methyl-(5-chloro-2-methoxyphenyl)benzene sulfon amide (5a): Yellow coloured crystals, yield: 83 %. m.p. 110-112 °C. IR (KBr, v_{max} , cm⁻¹): 3018 (aromatic C-H), 1445, 1340 (SO₂), 1220 (-OCH₃)), 1150 (aliphatic C-N str.); ¹H NMR (500 MHz, CD₃OD): δ 7.65 (dd, *J* = 7.5, 1.5 Hz, 2H, H-2' and H-6'), 7.62 (br.t, *J* = 7.5 Hz, 1H, H-4'), 7.53 (br.t, *J* = 7.5 Hz, 2H, H-3' and H-5'), 7.29 (dd, *J* = 8.5, 2.5 Hz, 1H, H-4), 7.23 (d, *J* = 2.5 Hz, 1H, H-6), 6.89 (d, *J* = 8.5 Hz, 1H, H-3), 3.34 (s, 3H, -OCH₃), 3.18 (s, 3H, -CH₃); EIMS: m/z 311 [M⁺], 296 [M-CH₃]⁺, 280 [M-OCH₃]⁺, 247 [M-SO₂]⁺, 170 [M-PhSO₂]⁺, 141 [PhSO₂]⁺.

N-Ethyl-*N*-(5-chloro-2-methoxyphenyl)benzene sulfon amide (5b): White amorphous powder, Yield: 79 %, m.p. 79-81 °C. IR (KBr, v_{max} , cm⁻¹): 3022 (aromatic C-H), 1444, 1337 (SO₂), 1227 (-OCH₃), 1151 (aliphatic C-N str.); ¹H NMR (500 MHz, CD₃OD): δ 7.64 (dd, J = 7.5, 1.5 Hz, 2H, H-2' and H-6'), 7.61 (br.t, J = 7.5 Hz, 1H, H-4'), 7.52 (br.t, J = 7.5 Hz, 2H, H-3' and H-5'), 7.32 (dd, J = 8.5, 2.5 Hz, 1H, H-4),7.19 (d, J = 2.5 Hz, 1H, H-6), 6.80 (d, J = 8.5 Hz, 1H, H-3), 3.63 (br.s, 2H, CH₂-1"), 3.34 (s, 3H,-OCH₃), 1.03 (t, J = 7.5 Hz, 3H, CH₃-2"); EIMS: m/z 325 [M⁺], 310 [M-CH₃]⁺, 294 [M-OCH₃]⁺, 261 [M-SO₂]⁺, 184 [M-PhSO₂]⁺, 141 [PhSO₂]⁺. *N*-Isopropyl-*N*-(5-chloro-2-methoxyphenyl)benzene sulfonamide (5c): Brown needle like crystals, Yield: 59 %, m.p. 90-92 °C. IR (KBr, v_{max} , cm⁻¹): 3019 (aromatic C-H), 1446, 1342 (SO₂), 1229 (-OCH₃), 1149 (aliphatic C-N str.); ¹H NMR (500 MHz, CD₃OD): δ 7.64 (dd, *J* = 7.5, 1.5 Hz, 2H, H-2' and H-6'), 7.61 (br.t, *J* = 7.5 Hz, 1H, H-4'), 7.52 (br.t, *J* = 7.5 Hz, 2H, H-3' and H-5'), 7.32 (dd, *J* = 8.5, 2.5 Hz, 1H, H-4), 7.19 (d, *J* = 2.5 Hz, 1H, H-6), 6.80 (d, *J* = 8.5 Hz, 1H, H-3), 4.50 (m, 1H, H-1"), 3.34 (s, 3H, -OCH₃), 1.01 (d, *J* = 6.6 Hz, 6H, CH₃-2" and CH₃-3"); EIMS: m/z 339 [M⁺], 324 [M-CH₃]⁺, 308 [M-OCH₃]⁺, 275 [M-SO₂]⁺, 198 [M-PhSO₂]⁺, 198 [PhSO₂]⁺.

N-(2-Bromoethyl)-*N*-(5-chloro-2-methoxyphenyl)benzene sulfonamide (5d): Dark brown solid, Yield: 64 %, IR (KBr, v_{max} , cm⁻¹): 3022 (aromatic C-H), 1449, 1337 (SO₂), 1232 (-OCH₃), 1145 (aliphatic C-N str.); ¹H NMR (500 MHz, CD₃OD): δ 7.96 (br.s, H-2' and H-6'), 7.62 (m, 1H, H-4'), 7.51 (br.t, *J* = 7.5 Hz, 2H, H-3' and H-5'), 7.37 (d, *J* = 2.5 Hz, 1H, H-6), 7.32 (dd, *J* = 8.5, 2.5 Hz, 1H, H-4), 6.88 (d, *J* = 8.5 Hz, 1H, H-3), 3.90 (br.s, 2H, CH₂-1"), 3.43 (t, *J* = 6.5, 2H, CH₂-2"), 3.27(s, 3H, -OCH₃); EIMS: m/z 404 [M⁺], 389 [M-CH₃]⁺, 373 [M-OCH₃]⁺, 340 [M-SO₂]⁺, 263 [M-PhSO₂]⁺, 141 [PhSO₂]⁺.

N-Allyl-*N*-(5-chloro-2-methoxyphenyl)benzene sulfon amide (5e): Dark brown gammy solid, Yield: 82 %, IR (KBr, v_{max} , cm⁻¹): 3019 (aromatic C-H), 1443, 1336 (SO₂), 1227 (-OCH₃), 1151 (aliphatic C-N str.); ¹H NMR (500 MHz, CD₃OD): δ 7.64 (dd, *J* = 7.5, 1.5 Hz, 2H, H-2' and H-6'), 7.62 (br.t, *J* = 7.5 Hz, 1H, H-4'), 7.51 (br.t, *J* = 7.5 Hz, 2H, H-3' and H-5'), 7.27 (dd, *J* = 8.5, 2.5 Hz, 1H, H-4), 7.17 (d, *J* = 2.5 Hz, 1H, H-6), 6.85 (d, *J* = 8.5 Hz, 1H, H-3), 5.74 (m, 1H, H-2"), 5.06 (dd, *J* = 17.5, 1.5 Hz, 1H, H_b-3"), 5.00 (dd, *J* = 10, 1.0 Hz, 1H, H_a-3"), 4.20 (br.s, 2H, H-1"), 3.33 (s, 3H, -OCH₃); EIMS: m/z 337 [M⁺], 322 [M-CH₃]⁺, 306 [M-OCH₃]⁺, 273 [M-SO₂]⁺, 183 [M-PhSO₂]⁺, 141 [M-C₆H₃(C1)(OCH₃) NCH₂CH=CH₂]⁺.

N-**Butyl**-*N*-(**5**-chloro-2-methoxyphenyl)benzene sulfon amide (**5f**): Colourless sticky solid, Yield: 72%, IR (KBr, v_{max} , cm⁻¹): 3015 (aromatic C-H), 1445, 1337 (SO₂), 1225 (-OCH₃), 1150 (aliphatic C-N str.); ¹H NMR (500 MHz, CD₃OD): δ 7.62 (dd, *J* = 7.5, 1.5 Hz, 2H, H-2' and H-6'), 7.60 (br.t, *J* = 7.5 Hz, 1H, H-4'), 7.51 (br.t, *J* = 7.5 Hz, 2H, H-3' and H-5'), 7.30 (dd, *J* = 8.5, 2.5 Hz, 1H, H-4), 7.18 (d, *J* = 2.5 Hz, 1H, H-6), 6.88 (d, *J* = 8.5 Hz, 1H, H-3), 3.57 (br.s, 2H, H-1"), 3.33 (s, 3H,-OCH₃), 1.38 (m, 2H, H-3"), 1.27 (m, 2H, H-2"), 0.85 (t, *J* = 7.0 Hz, 3H, H-4"); EIMS: m/z 353 [M⁺], 338 [M-CH₃]⁺, 322 [M-OCH₃]⁺, 289 [M-SO₂]⁺, 212 [M-PhSO₂]⁺, 141 [PhSO₂]⁺.

N-**Pentyl**-*N*-(**5**-**chloro-2**-**methoxyphenyl**)**benzene sulfonamide** (**5g**): Off white solid, Yield: 67 %, IR (KBr, v_{max} , cm⁻¹): 3016 (aromatic C-H), 1442, 1339 (SO₂), 1224 (-OCH₃), 1152 (aliphatic C-N str.); ¹H NMR (500 MHz, CD₃OD): δ 7.62 (dd, *J* = 7.5, 1.5 Hz, 2H, H-2' and H-6'), 7.60 (br.t, *J* = 7.5 Hz, 1H, H-4'), 7.51 (br.t, *J* = 7.5 Hz, 2H, H-3' and H-5'), 7.32 (dd, *J* = 8.5, 2.5 Hz, 1H, H-4), 7.18 (d, *J* = 2.5 Hz, 1H, H-6), 6.90 (d, *J* = 8.5 Hz, 1H, H-3), 3.58 (br.s, 2H, H-1"), 3.32 (s, 3H, -OCH₃), 1.36 (m, 2H, H-2"), 1.33-1.30 (m, 4H, H-3" and H-4"), 0.86 (t, *J* = 7.5 Hz, 3H, H-5"); EIMS: m/z 367 [M⁺], 352 [M-CH₃]⁺, 336 [M-OCH₃]⁺, 303 [M-SO₂]⁺, 226 [M-PhSO₂]⁺, 141 [PhSO₂]⁺.

N-Benzyl-*N*-(5-chloro-2-methoxyphenyl)benzene sulfonamide (5h): Colourless sticky solid, Yield: 75 %, IR (KBr, v_{max} , cm⁻¹): 3020 (aromatic C-H), 1447, 1338 (SO₂), 1225 (-OCH₃), 1150 (aliphatic C-N str.); ¹H NMR (500 MHz, CD₃OD): δ 7.60 (dd, *J* = 7.5, 1.5 Hz, 2H, H-2' and H-6'), 7.59 (br.t, *J* = 7.5 Hz, 1H, H-4'), 7.48 (br.t, *J* = 7.5 Hz, 2H, H-3' and H-5'), 7.30 (dd, *J* = 8.5, 2.5 Hz, 1H, H-4), 7.18 (d, *J* = 2.5 Hz, 1H, H-6), 7.12-6.95 (m, 5H, H-2" to H-6"), 6.88 (d, *J* = 8.5 Hz, 1H, H-3), 4.67 (s, 2H, -CH₂-of benzyl), 3.31 (s, 3H, -OCH₃); EIMS: m/z 387 [M⁺], 372 [M-CH₃]⁺, 356 [M-OCH₃]⁺, 323 [M-SO₂]⁺, 246 [M-PhSO₂]⁺, 141 [PhSO₂]⁺.

N-4''-Bromobenzyl-*N*-(5-chloro-2-methoxyphenyl)benzene sulfonamide (5i): Brown coloured sticky solid, Yield: 81 %, IR (KBr, v_{max} , cm⁻¹): 3013 (C-H aromatic str.), 1442, 1334 (SO₂), 1221 (-OCH₃), 1145 (C-N str.); ¹H NMR (500 MHz, CD₃OD): δ 7.69 (d, *J* = 8.5 Hz, 2H, H-3" and H-5"), 7.63 (dd, *J* = 7.5, 1.5 Hz, 2H, H-2' and H-6'), 7.61 (br.t, *J* = 7.5 Hz, 1H, H-4'), 7.53 (br.t, *J* = 7.5 Hz, 2H, H-3' and H-5'), 7.35 (dd, *J* = 8.5, 2.5 Hz, 1H, H-4), 7.28 (d, *J* = 8.5 Hz, 2H, H-2" and H-6"), 7.21 (d, *J* = 2.5 Hz, 1H, H-6), 6.93 (d, *J* = 8.5 Hz, 1H, H-3), 4.75 (s, 2H, -CH₂-of benzyl), 3.35 (s, 3H,-OCH₃); EIMS: m/z 466 [M⁺], 451 [M-CH₃]⁺, 435 [M-OCH₃]⁺, 402 [M-SO₂]⁺, 325 [M-PhSO₂]⁺, 141 [PhSO₂]⁺.

N-2''-Phenylethyl-*N*-(5-chloro-2-methoxyphenyl)benzene sulfonamide (5j): Colourless sticky solid, Yield: 58 %, 67-69 °C. IR (KBr, v_{max} , cm⁻¹): 3019 (aromatic C-H), 1448, 1337 (SO₂), 1223 (-OCH₃), 1154 (aliphatic C-N str.); ¹H NMR (500 MHz, CD₃OD): δ 7.61 (dd, *J* = 7.5, 1.5 Hz, 2H, H-2' and H-6'), 7.58 (br.t, *J* = 7.5 Hz, 1H, H-4'), 7.49 (br.t, *J* = 7.5 Hz, 2H, H-3' and H-5'), 7.30 (dd, *J* = 8.5, 2.5 Hz, 1H, H-4), 7.19 (d, *J* = 2.5 Hz, 1H, H-6), 7.12-6.92 (m, 5H, H-2" to H-6"), 6.87 (d, *J* = 8.5 Hz, 1H, H-3), 3.70 (t, *J* = 7.8 Hz, 2H, H-8"), 3.34 (s, 3H, -OCH₃), 2.75 (t, *J* = 7.8 Hz, 2H, H-7"); EIMS: m/ z 480 [M⁺], 465 [M-CH₃]⁺, 449 [M-OCH₃]⁺, 416 [M-SO₂]⁺, 339 [M-PhSO₂]⁺, 141 [PhSO₂]⁺.

N-3''-Phenylpropyl-*N*-(5-chloro-2-methoxyphenyl)benzene sulfonamide (5k): Colourless gammy solid, Yield: 78 %, IR (KBr, v_{max} , cm⁻¹): 3015 (C-H aromatic str.), 1445, 1335 (SO₂), 1224 (-OCH₃), 1155 (aliphatic C-N str.); ¹H NMR (500 MHz, CD₃OD): δ 7.64 (dd, *J* = 7.5, 1.5 Hz, 2H, H-2' and H-6'), 7.61 (br.t, *J* = 7.5 Hz, 1H, H-4'), 7.51 (br.t, *J* = 7.5 Hz, 2H, H-3' and H-5'), 7.30 (dd, *J* = 8.5, 2.5 Hz, 1H, H-4), 7.19 (d, *J* = 2.5 Hz, 1H, H-6), 7.16-6.98 (m, 5H, H-2" to H-6"), 6.91 (d, *J* = 8.5 Hz, 1H, H-3), 3.59 (t, *J* = 7.8 Hz, 2H, H-9"), 3.36 (s, 3H,-OCH₃), 2.30 (t, *J* = 7.8 Hz, 2H, H-7"), 1.74 (m, 2H, H-8"); EIMS: m/z 494 [M⁺], 479 [M-CH₃]⁺, 463 [M-OCH₃]⁺, 430 [M-SO₂]⁺, 353 [M-PhSO₂]⁺, 141 [PhSO₂]⁺.

RESULTS AND DISCUSSION

Our objective was to synthesize some new *N*-substituted derivatives of sulfonamides and to screen out their enzymatic activities. Keeping that objective in mind, we synthesized *N*-substituted-(5-chloro-2-methoxyphenyl)benzene sulfonamide in excellent yield and having good biological activities. The synthesis was carried out through the reaction of benzene sulfonyl chloride (1) with 2-amino-4-chloroanisole (2) yielded

N-(5-chloro-2-methoxyphenyl)benzene sulfonamide (3) and finally a series of N-substituted (5-chloro-2-methoxyphenyl) benzenesulfonamide (5a-k) derivatives were obtained by the reaction of parent compound (3) with different electrophiles (4a-k) in the presence of DMF and sodium hydride (NaH) which act as a strong base represented in Scheme-I. Complete conversion was achieved within 2-4 h by simple stirring at room temperature. The product was isolated by adding cold water in the reaction mixture. It was filtered, washed with distilled water and dried to afford the derivatives (5a-k). The structures of the synthesized compounds were established by ¹H NMR (Table-1), IR and mass spectral data as described in experimental section. Compound 3 was synthesized as colourless needle like crystals. The molecular formula C13H12NO3SCl was established by HRMS showing molecular ion peak at m/z 297.7585 (calcd. for C₁₃H₁₂NO₃SCl, 297.7582). The IR spectrum revealed the presence of methoxy group (1225 cm⁻¹), a sulfonyl group (1338, 1447 cm⁻¹) and -NHgroup (3100 cm⁻¹) in the molecule. The EIMS gave a distinct peak at m/z 233 after the removal of -SO₂ group and further two fragment ion peaks were observed at m/z 282 and 266 which showed the presence of methyl and methoxy groups, respectively in the molecule. In the aromatic region of the ¹H NMR spectrum of 3 signals which resonated at d 7.41 (d, J =2.5 Hz), 6.79 (d, J = 8.5 Hz), 7.05 (dd, J = 8.5, 2.5 Hz) and 7.41 (d, J = 2.5 Hz) were assigned to the protons of the tri-substituted ring and the signals appeared at δ 7.55 (t, J =7.5 Hz), 7.45 (t, J = 7.5 Hz) and 7.71 (dd, J = 7.5, 1.5 Hz) were assigned to the protons of the mono-substituted ring. In the aliphatic region of ¹H NMR spectrum, a singlet was appeared at δ 3.51 corresponding to the methoxy group present in the molecule. On the basis of these evidences the structure of 3 was assigned as N-(5-chloro-2-methoxyphenyl) benzenesulfonamide. The structure of compound 3 was also confirmed by single-crystal XRD^{11,17}. Similarly on the basis of spectral evidences from IR, EI-MS and ¹H NMR, the structures of other derivatives were elucidated as described in experimental section.

Enzyme inhibition activity: The screening of these synthesized compounds against acetylcholinesterase (AChE), butyrylcholinesterase (BChE) and lipoxygenase (LOX) enzymes revealed that these molecules exhibited good inhibitory potential against acetylcholinesterase as it was evident from their IC₅₀ values. It is obvious from Table-1 that compounds 5a, 5e and 5j were found to be promising inhibitors against acetylcholinesterase enzyme having IC₅₀ value of 34.61 \pm 0.62, 40.21 \pm 0.25 and 45.11 \pm 0.22 µmol/L, respectively, relative to Eserine, a reference standard with IC₅₀ value of 0.04 $\pm 0.001 \,\mu$ mol/L, probably due to the N-substitution of methyl, allyl and 2-phenylethyl groups, respectively in these molecules. The screening against butyrylcholinesterase enzyme exposed that the compounds 5a and 5g exhibited promising inhibitory potential having IC₅₀ 44.57 \pm 0.04 and 54.57 \pm 0.05 μ mol/L as compared to standard. However, only few compounds (Table-1) showed inhibition against lipoxygenase enzyme but all other showed very weak activity. DPPH is a stable free radical at room temperature. DPPH radical is scavenged by antioxidants through the donation of a proton and form reduced DPPH.





Scheme-I: Outline for synthesis of N-substituted (5-chloro-2-methoxyphenyl)benzene sulfonamide

BIOACTIVITY STUDIES OF N-SUBSTITUTED (5-CHLORO-2-METHOXYPHENYL)BENZENE SULFONAMIDE									
	DPPH		LOX		AChE		BChE		
C. No.	Inhibition (%) at 0.5 mM	IC ₅₀ (µmol)	Inhibition (%) at 0.5 mM	IC ₅₀ (µmol)	Inhibition (%) at 0.5 mM	IC ₅₀ (µmol)	Inhibition (%) at 0.5 mM	IC ₅₀ (µmol)	
3	89.23±0.63	98.11±0.44	50.79±0.31	398.91±0.34	63.82±0.31	139.11±0.41	58.07±0.31	295.6±0.22	
5a	3.63±0.13	Nil	44.46±0.41	Nil	87.50±0.14	34.61±0.62	25.60±0.19	44.57±0.04	
5b	0.57±0.54	Nil	76.86±0.34	166.11±0.24	84.65±0.22	75.81±0.04	45.78±0.45	<400	
5c	0.51±0.22	Nil	64.56±0.34	<400	74.01±0.71	132.51±0.66	64.46±0.24	180.41±0.11	
5d	0.44 ± 0.18	Nil	68.94±0.61	179.11±0.62	69.85±0.35	165.61±0.32	52.08±0.51	<400	
5e	0.76±0.16	Nil	69.67±0.13	Nil	69.08±0.52	40.21±0.25	67.95±0.44	<400	
5f	89.23±0.63	Nil	95.13±0.87	<400	71.93±0.01	66.71±0.92	71.72±0.59	295.6±0.22	
5g	3.63±0.11	Nil	60.54±0.25	Nil	54.57±0.05	213.25±0.07	39.16±0.05	54.57±0.05	
5h	0.57±0.54	Nil	56.45±0.22	384.91±0.28	57.57±0.34	192.91±0.14	37.63±0.22	57.57±0.34	
5i	0.61±0.22	Nil	70.16±0.19	260.81±0.34	59.10±0.36	360.81±0.25	60.09±0.11	59.10±0.36	
5j	0.56±0.12	Nil	47.50±0.09	Nil	78.73±0.59	45.11±0.22	92.84±0.09	<400	
5k	0.58±0.15	Nil	77.71±0.59	144.43±0.18	66.12±0.73	67.91±0.41	63.82±0.59	66.12±0.73	
Control	Quercetin 93.21±0.97	16.96±0.14	Baicalein 93.79±1.27	22.4±1.3	Eserine 91.29±1.17	0.04±0.001	Eserine 82.82±1.09	0.85±0.001	

Note: IC_{50} values (concentration at which there is 50 % enzyme inhibition) of compounds were calculated using EZ: Fit enzyme kinetics software (Perella Scientific Inc. Amherst, USA). LOX = Lipoxygenase. AChE = Acetyl cholinesterase. BChE = Butyryl cholinesterase. DPPH = 1,1-diphenyl-2-picrylhydrazyl radical.

The colour changes from violet to yellow after reduction of DPPH and it can be quantified by decrease of absorbance at wavelength 517 nm. Radical scavenging activity increased with increasing percentage of the free radical inhibition. The colour change from violet to yellow and fall in absorbance of the stable radical DPPH was measured for three different concentrations of samples and the results are shown in Table-1. These results showed that the parent compound N-(5-chloro-2methoxyphenyl)benzene sulfonamide (3) showed scavenging activity against 1,1-diphenyl-2-picrylhydrazyl (DPPH) radical but all others remained inactive. The IC₅₀ value for the parent compound 98.11 \pm 0.44 μ mol/L, relative to quercetin, a reference standard with IC₅₀ value of $16.96 \pm 0.14 \mu mol/L$, was calculated from the curves plotted. IC₅₀ is the concentration of fraction causing 50 % inhibition of absorbance and lower its value means greater antioxidant activity of the fraction.

Conclusion

The proposed structure of the synthesized compound is well supported by spectroscopic data. From the enzyme inhibition data (Table-1), it might be concluded that the three compounds **5a**, **5e** and **5j** have promising activity against acetylcholinesterase enzyme as compared to relative standard. Some synthesized compounds exposed moderate activity against butyrylcholinesterase and lipoxygenase enzymes but only parent compound **3** showed scavenging activity against DPPH but all were remained inactive. Hence on the basis of aforesaid results, these synthesized derivatives provide an overall indispensable basis to introduce new drug candidates for the treatment of Alzheimer's disease and other associated diseases.

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