



HPLC Analysis of Fatty Acids and Triterpenoids in Fruit of Hawthorn (*Crataegus azarolus*) in Iraqi Kurdistan Region

BARAM AHAMD AMEEN*, SRWA N. MAJEED and DALIA A. ABDUL*

Department of Chemistry, School of Science, Faculty of Science & Science Education, University of Sulaimani, P.O. Box: 334, Sulaimani, Iraq

*Corresponding authors: E-mail: barmjaff@yahoo.com; dalyadya@yahoo.com

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Fatty acids and triterpenoids content of three types of hawthorn fruit (*Crataegus azarolus*) which grows in Iraqi Kurdistan region (season 2011) including Hawraman, Qaradax and house garden, were determined by RP-HPLC technique. The percentages of oils were found to be 12, 10 and 14 %, respectively. HPLC analysis of these oils show contain palmitic acid, palmitoleic (ω 7) acid and high levels of α -linoleic acid about [37.202, 31.402 and 33.816 %], linoleic acid [9.686, 21.556 and 19.135 %], respectively. Divert compounds of triterpenoids composition were also analyzed by HPLC technique in all of them mentioned four triterpenoids compound were detected oleanolic acid, ursolic acid, α -amyirin, β -amyirin and a high ratio (choline, acetylcholine) have also been detected. The ratio of acetylcholine is about 53.800, 24.6144 and 30.8198 % in each types, respectively and a high level of choline detected in Qaradax hawthorn fruits is about 39.2849 % and in house garden about 35.4523 %.

Key Words: HPLC analysis, Hawthorn, Fatty acids, Triterpenoids, ω -3 fatty acid, α -Amyirin, Acetylcholine.

INTRODUCTION

Hawthorn is a spiking bush or tree, the tree reaches 13 feet in height and grows along the edges of woods and forests. Hawthorn has smooth, gray dark and sharp thorns which grow along the branches. The medicinal parts are the flowers and the fruit (Fig. 1), the Latin name is *Crataegus azarolus* it belong to the genus (crataegus), the genus (crataegus) is a member of the rose family (Rosaceae). It is cultivated and grows wild in Kurdistan region-north Iraq, a local name of hawthorn fruit in kurdish is (goezh)¹.



Fig. 1. Dr. Baram while he harvest the hawthorn fruits and its leaves in Qaradax

Hawthorn contains amino acids and essential fatty acids. It also contains the minerals *e.g.*, calcium magnesium, chromium, potassium, selenium, iron and zinc vitamins B-1, B-2, B3 and C², oleanolic acid³ or oleanic acid (Fig. 2a) is a naturally occurring triterpenoid widely distributed in food and medicinal plants, related to betulinic acid. It is relatively non-toxic, antitumor and hepatoprotective as well as exhibiting antiviral properties⁴, oleanolic acid was found to exhibit strong anti HIV activity the related compound betulinic acid was use to create the first commercial maturation inhibitor drug.

An extremely potent synthetic triterpenoid analogue of oleanolic acid was found in 2005. It is a powerful inhibitor of cellular inflammatory processes⁵. Ursolic acid (Fig. 2b) is a pentacyclic triterpene acid, used in cosmetics that is also capable of inhibiting various types of cancer cells by inhibiting the STAT₃ activation pathway^{6,7} and human fibrosarcoma cells by reducing the expression of matrix metalloproteinase -9 by acting through the glucocorticoid receptor. It may also decrease proliferation of cancer cells induce apoptosis⁸. Ursolic acid can serve as a starting material for synthesis of more potent bioactive derivatives as antitumor agent⁹ other names for ursolic acid include urson, prunol and malol. α - and β -amyirin are pentacyclic triterpenes (Fig. 2c,d) found in plants and are known to exhibit pronounced anti-inflammatory effects¹⁰. Choline is 2-hydroxy-N,N,N-trimethylethanminium (Fig. 3a) the other name of it is bilineurine choline is a water soluble

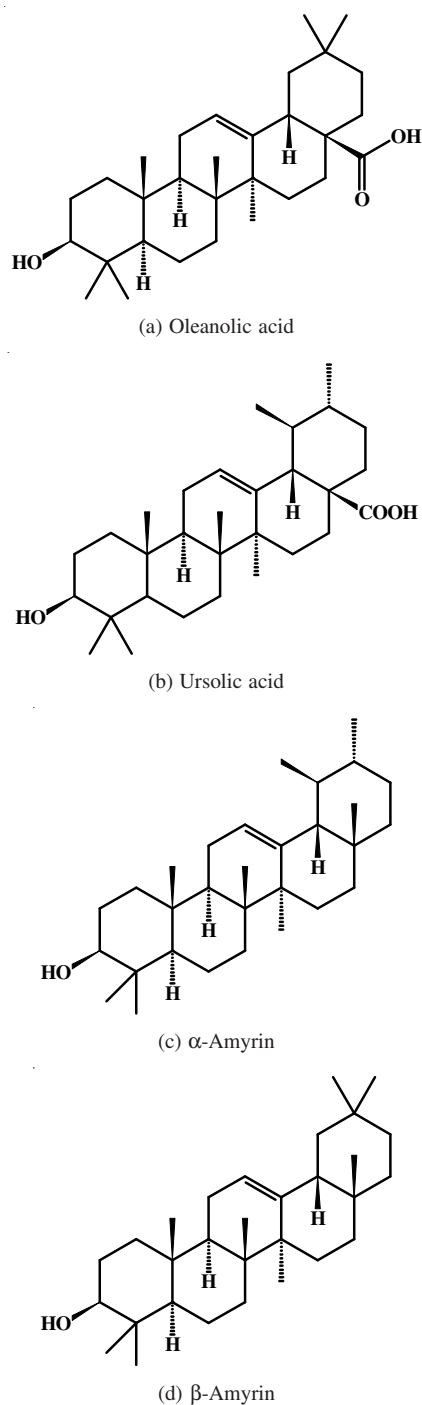


Fig. 2. Chemical structure of hawthorn triterpenoids

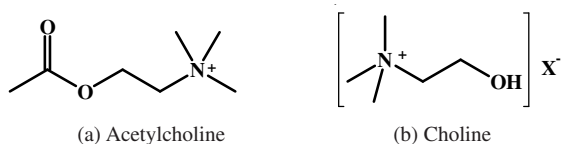


Fig. 3. Chemical structure of acetylcholine and choline

essential nutrient¹¹. It is usually grouped within the B-complex vitamins, generally refers to the various quaternary ammonium salts and it is a precursor molecule for the neurotransmitter, it must be consumed through the diet in order for the body to remain healthy¹². Acetylcholine(ash) is 2-acetoxy-N,N,N-trimethylethanaminium was first identified in 1914 by

Henry Hall *et al.*, for its action on heart tissue, is one of many neurotransmitters in the autonomic nervous system (ANS) and it slows the heart rate when functioning as an inhibitory neurotransmitters. However it also behaves at neuromuscular junction¹³.

EXPERIMENTAL

Three types of hawthorn [*Crataegus azarolus*] fruit, [Hawramane, Qaradax and house garden hawthorn] were harvested in its optimum state for two consecutive seasons in 2011 in Kurdistan region -north Iraq, Hawramane, Qaradax, garden Fig. 1. After a morphological and chemical characterization, the samples were prepared for determination of oils, fatty acids, triterpenoids and other chemical constituents.

A-Preparation of the sample for the determination of total oil contents: The three types of hawthorn fruits Hawramane, Qaradax, house garden were oven-dried at 50 °C for 18 h and ground through a wiley mill to pass a 30-40 mesh screen and stored tightly at 4 °C. 10 g of the samples were extracted with petroleum ether (40-60 °C) using Soxhlet apparatus according to AAcc methods¹⁴. The oils were recovered by petroleum ether evaporation in a rotary evaporator at a 60 °C (Heidolph Laboratory 4003 control). The oils were dried in desiccators for 1 h and finally the extracted oils were weighted. The percentage of the total oils content are (12, 10 and 14 %), respectively. The characteristic properties of the oil were determined by conventional methods¹⁵ and the results are presented in Table-1.

TABLE-1
SPECIFIC TESTS FOR OILS IN HAWTHORN FRUIT
[HAWRAMANE, QARADAX, HOUSE GARDEN]

Samples	Hawramane hawthorn	Qaradax hawthorn	House garden hawthorn
Oil	12 %	10 %	14 %
Colour	Light yellow	Light yellow	Light yellow
Saponification	172.10	169.00	170.10
Iodine value	152.8	152.0	150.0
Acid value	1.25	1.20	1.21
Peroxide value	1.20	1.19	1.21
Ash	0.2 g	0.19	0.21

B-Fatty acids composition: Fatty acids were separated on FLC (fast liquid chromatographic) column. Supelcosil LC, 3 μ m particle size (50 mm \times 4.6 mm i.d.) column, mobile phase:acetonitrile:acetone (59:41, v/v).

Detection: Refractive index detector LC6-RID flow rate 1.0 mL/min, injection: 20 μ L saturated and unsaturated fatty acid FAME fatty acid methyl ester. The sequences of the eluted fatty acids standard were as follow, each standard was 50 μ g/mL C16:1, C18:1, C18:2, C18:3 and C20:2¹⁶.

C-Extraction of samples

By hydro-distillation method: Hydro-distillation extraction method was used for extraction of essential oil, 1 g of the crushed fruit were immersed, the hot water help to release the aromatic molecules from the fruit, the molecules of these volatile oils evaporate in to steam, the steam, was condense in to liquid form. The oil was separated from water without losing

oil. The analysis of the oil and its constituents were analysis by HPLC, according to reference analysis procedure¹⁷.

D-Pentacyclic triterpenoids: The alcoholic extract of pentacyclic triterpenoids were separated on FLC (fast liquid chromatography) column, Um particle size (50 mm × 4.6 mm i.d.) chiral column, mobile phase were 0.1 % acetic acid in deionized water:acetonitrile (20:80 v/v) detection UV set at 264 nm, flow rate 1.3 mL/min. The sequence of the eluted standard was 25 µg/mL.

10 g of each samples were weighed, then dissolved in 10 mL HPLC methanol, the samples shaking and agitated in ultrasonic bath for 10 min, then concentrated by evaporating the solvent with a stream of liquid N₂ until reach 1 mL, then 20 µL were injected on HPLC column. The concentration for each compound were quantitatively determined by comparison the peak area of the standard with that of the samples.

Concentration of sample µg/mL = area of sample/area of standard × Con. of standard × dilution factor: The separation occurred on liquid chromatography Shimadzu 10AV-LC equipped with binary delivery pump model LC-10AV Shimadzu, the eluted peaks were monitored by UV-VIS 10A-SPD spectrophotometer.

E-Determination of elements: 10 g of samples were weighted in to a porcelain crucibles, placed in a muffle furnace temperature was increased to 600 °C and held at the temperature for 6 h, the samples were removed from the furnace

and weighted, the determine percentage of ash for samples were (0.2 g), (0.19 g) and (0.21 g) %, respectively. The residual ash was dissolved in 1:1 nitric acid filtered and the volume was completed to (100 mL) distilled water, the solution was injected to OES-ICP Perkin Elmer 2100 for determine.

RESULTS AND DISCUSSION

Hawthorn fruits [Hawraman, Qaradax and house garden] have oil contents 12, 10 and 14 %, respectively which have a yellow colour. The characteristics properties of the oils compared well to each other Table-1. The iodine value is often the most useful figure for indentifying oil or at least into a particular group which gives a reasonably quantitative measure for unsaturated of oil¹⁸. The saponification number represents the amount of saponifiable material which is inversely proportional to the mean of the molecular weights of the fatty acids in the glycerides present¹⁸. The peroxide value is an indicator of the products of primary oxidation measures rancidity or degree of oxidation but not stability of a fat¹⁹. A rancid test often begins to be noticeable when the peroxide value is between 10 and 20¹⁸.

The HPLC analysis of fatty acid composition in the three types of hawthorn fruits are presented in Tables 2-4 from the result the each types of hawthorn fruits contain high percentage of α-linoleic acid which is the essential fatty acid must

TABLE-2
FATTY ACID COMPOSITION IN HAWRAMANE HAWTHORN FRUIT

No	Subjects	Rt (min)	Area (standard)	df	Area (sample)	Conc. (µg/mL)	Conc. of sample	%
1	Palmitic C16 = 0	1.03	20765	3	7108	50	51.3460	7.000621297
2	Palmitoleic C16 = 1	1.87	23081	3	6741	50	43.8088	5.972976517
3	Stearic C18 = 0	2.7	27714	3	0	50	0.0000	0
4	Oleic C18 = 1	3.71	24155	3	20550	50	127.6133	17.399064
5	Linoleic C18 = 2	4.61	18272	3	8654	50	71.0431	9.686165951
6	α-Linoleic C18 = 3	6.02	12482	3	22706	50	272.8649	37.20296524
7	Arachidic C20 = 2	6.86	17984	3	19995	50	166.7732	22.738207
	Total	–	–	–	–	–	733.4494	–

TABLE-3
FATTY ACID COMPOSITION IN QARADAX HAWTHORN FRUIT

No	Subjects	Rt (min)	Area (standard)	df	Area (sample)	Conc. (µg/mL)	Conc. of sample	%
1	Palmitic C16 = 0	1.03	20765	3	8359	50	60.3829	7.51211981
2	Palmitoleic C16 = 1	1.87	23081	3	11605	50	75.4192	9.382760694
3	Stearic C18 = 0	2.7	27714	3	10697	50	57.8967	7.202825596
4	Oleic C18 = 1	3.71	24155	3	0	50	0.0000	0
5	Linoleic C18 = 2	4.61	18272	3	21107	50	173.2733	21.55661372
6	α-Linoleic C18 = 3	6.02	12482	3	21004	50	252.4115	31.40204614
7	Arachidic C20 = 2	6.86	17984	3	22111	50	184.4223	22.94363403
	Total	–	–	–	–	–	803.8058	–

TABLE-4
FATTY ACID COMPOSITION IN HOUSE GARDEN HAWTHORN FRUIT

No	Subjects	Rt (min)	Area (standard)	df	Area (sample)	Conc. (µg/mL)	Conc. of sample	%
1	Palmitic C16 = 0	1.03	20765	3	4490	50	32.4344	4.804159728
2	Palmitoleic C16 = 1	1.87	23081	3	37429	50	243.2455	36.02936732
3	Stearic C18 = 0	2.7	27714	3	7752	50	41.9571	6.214663013
4	Oleic C18 = 1	3.71	24155	3	0	50	0.0000	0
5	Linoleic C18 = 2	4.61	18272	3	15737	50	129.1895	19.13545931
6	α-linoleic C18 = 3	6.02	12482	3	18998	50	228.3048	33.81635063
7	Arachidic C20 = 2	6.86	17984	3	0	50	0.0000	0
	Total	–	–	–	–	–	675.1313	–

TABLE-5
TRITERPENOID COMPOSITION IN HAWRAMANE HAWTHORN AND OTHER CHEMICAL CONSTITUENT*

No	Subjects	Rt (min)	Area (standard)	df	Area (sample)	Conc. (µg/mL)	Conc. of sample	%
1	Chlorogenic acid	0.89	21630	3	0	25	0.0000	0
2	Oleanolic acid	2.03	20404	3	17070	25	62.7450	2.772614876
3	Ursolic acid	2.87	22267	3	13878	25	46.7441	2.065553827
4	α-Amyrin	3.87	13168	3	27898	25	158.8966	7.021414227
5	β-Amyrin	4.94	16621	3	85319	25	384.9904	17.01217924
6	*Choline	5.87	20287	3	106067	25	392.1243	17.32741588
7	*Acetylcholine	6.77	31108	3	504998	25	1217.5276	53.80082195
	Total	–	–	–	–	–	2263.0280	–

TABLE-6
TRITERPENOID COMPOSITION IN QARADAX HAWTHORN AND OTHER CHEMICAL CONSTITUENT*

No	Subjects	Rt (min)	Area (standard)	df	Area (sample)	Conc. (µg/mL)	Conc. of sample	%
1	Chlorogenic acid	0.89	21630	3	0	25	0.0000	0
2	Oleanolic acid	2.03	20404	3	28840	25	106.0086	3.582365987
3	Ursolic acid	2.87	22267	3	22177	25	74.6969	2.524242642
4	α-Amyrin	3.87	13168	3	69842	25	397.7939	13.44271097
5	β-Amyrin	4.94	16621	3	108542	25	489.7810	16.55124555
6	*Choline	5.87	20287	3	314452	25	1162.5129	39.28498067
7	*Acetylcholine	6.77	31108	3	302115	25	728.3858	24.61445418
	Total	–	–	–	–	–	2959.1791	–

TABLE-7
TRITERPENOID COMPOSITION IN HOUSE GARDEN HAWTHORN AND OTHER CHEMICAL CONSTITUENT*

No	Subjects	Rt (min)	Area (standard)	df	Area (sample)	Conc. (µg/mL)	Conc. of sample	%
1	Chlorogenic acid	0.89	21630	3	0	25	0.0000	0
2	Oleanolic acid	2.03	20404	3	17579	25	64.6160	4.611192631
3	Ursolic acid	2.87	22267	3	17733	25	59.7285	4.262407024
4	α-Amyrin	3.87	13168	3	21322	25	121.4421	8.666475927
5	β-Amyrin	4.94	16621	3	50270	25	226.8365	16.18773736
6	*Choline	5.87	20287	3	134378	25	496.7886	35.45232792
7	*Acetylcholine	6.77	31108	3	179130	25	431.8744	30.81985914
	Total	–	–	–	–	–	1401.2862	–

be taking from diet because the human body not able to synthesized it. Hawraman hawthorn does not contain stearic acid, while Qaradax hawthorn does not contain oleic acid and house garden hawthorn does not contain oleic acid and arachidic acid but contain high percentage of palmitoleic acid *ca.* 36.029 %.

HPLC analysis of triterpenoids compounds in hawthorn samples are presented in Tables 5-7. All the four different triterpenoids compound are indicated in all three samples of hawthorn oleanolic acid, ursolic acid, α-amyirin and β-amyirin (Fig. 2), all samples contain high percentage of β-amyirin while Qaradax hawthorn contain high percentage of α-amyirin (13.442 %) by comparison with Hawraman and house garden hawthorn (7.021 and 8.664 %). Another natural occurring compounds like choline and acetylcholine are detected in these hawthorn fruits as shown in Fig. 3, Qaradax and house garden hawthorn contain high percentage of choline 39.284 and 35.452 %, while Hawraman hawthorn fruit contain high percentage of acetylcholine *ca.* 53.800 %.

Elemental analysis have been done by Inductive couple plasma [ICP] techniques (OES-ICP Perkin Elmer 2100) for each types of hawthorn and the result cited in Table-8 which a high amount of Cu and Mo elements have been detected in each of these three types of hawthorn fruits [in ppb].

TABLE-8
ELEMENTAL ANALYSIS OF HAWTHORN FRUITS
[HAWRAMANE, QARADAX, HOUSE GARDEN, ppb]

Samples	Hawramane hawthorn	Qaradax hawthorn	House garden hawthorn
Co	2.722	2.511	2.00
Cr	16.65	15.60	17.15
Cu	35.31	34.31	34.00
Mo	41.63	40.00	41.50
Ni	94.491	9.50	9.00
Se	6.491	6.50	6.00

Conclusion

The three types of hawthorn fruits [Hawraman, Qaradax and house garden] in Iraqi Kurdistan region are rich source of polyunsaturated fats including α-linoleic acid (according to HPLC analysis) and triterpenoids compound including β-amyirin which is knowing as exhibit pronounced anti-inflammatory effect, also all three types of hawthorn fruit contain another phytochemical compounds like choline and acetylcholine which they have special physiological effect in our body especially action of neurotransmitter and the amount of these elements like Co, Cr, Cu, Mo, Ni and Se are exist in these fruits, for that we must consume it through our diet in order to the remain healthy.

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REFERENCES

- H.I. Chakravary and Ali-Al-Rawi, Medicinal Plant of Iraq, Ministry of Agriculture Baghdad, edn. 2, 1 (1964).
- Prescription for Nutritional Healing Phyllis Balch, University of Maryland Medical Center-Hawthorn (2003).
- T. Cui, J.-Z. Li, H. Kayahara, L. Ma, L.-X. Wu and K. Nakamura, *J. Agric. Food Chem.*, **54**, 4574 (2006).
- J. Liu, *J. Ethnopharmacol.*, **49**, 57 (1995).
- A.T. Dinkova-Kostova, K.T. Liby, K.K. Stephenson, W.D. Holtzclaw, X. Gao, N. Suh, C. Williams, R. Risingsong, T. Honda, G.W. Gribble, M.B. Sporn and P. Talalay, *Proc. Nat. Acad. Sci. USA*, **102**, 4584 (2005).
- S. Shishodia, S. Majumdar, S. Banerjee and B.B. Aggarwal, *Cancer Res.*, **63**, 4375 (2003).
- A.K. Pathak, M. Bhutani, A.S. Nair, K.S. Ahn, A. Chakraborty, H. Kadara, S. Guha, G. Sethi and B.B. Aggarwal, *Mol. Cancer Res.*, **5**, 943 (2007).
- X. Wang, F. Zhang, L. Yang, Y. Mei and H. Long, *J. Biomed. Biotechnol.*, **2011**, 419343 (2011).
- M. Mac, S.Q. Cai, J.R. Cui and R.Q. Wang, *Eur. J. Med. Chem.*, **40**, 582 (2005).
- C.E. Vitor, C.P. Figueiredo, D.B. Hara, A.F. Bento, T.L. Mazzuco and J.B. Calixto, *Br. J. Pharmacol.*, **157**, 1034 (2009).
- Dietary Reference Intakes for Thiamine, Riboflavin, Niacin, Vitamin B6, Folate, Vitamin B12, Panthothenic Acid, Biotin and Chloine, Institute of Medicine (1998).
- <http://ipi.oregonstate.edu/infocenter/othernvts/choline> (at Linus paulin Inst)AJCN 92 n5 1113-1119(Nov2010)
- N.A. Compbell and J.B. Reece, 48 Biology, SanFrancisco, CA, Pearson Education, Inc., edn. 6, p. 1037 (2002).
- AAcc, Approved Methods of the AAcc, American Association of Coreal Chemists INC, stpau, Minn (1987).
- J.Y. Goe, *J. Am. Oil Chem. Soc.*, **72**, 120 (1995).
- ISO/IEC 17025: 1999. The Present List of Methods Reflects the Amendments Adopted by the 34th Session of the Codex Alimentarius Commission in (2011).
- www.che.utah.edu/.../Instrumental%20Analysis%20CHE5503/.../HPLC%20Procedure%20Final
- D. Pearson, The Chemical Analysis of Food, Chemical Publishing Company INC. New York, edn. 6 (1971).
- D.T. Plummer, An Introduction to Practical Biochemistry, Academic Press, London, edn. 3 (1987).