

Chemical Analysis of Burdock Root Constituents

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An ethanolic solution of extracted burdock root was concentrated, suspended by water and extracted by petroleum ether, chloroform, ethyl acetate and *n*-butanol. Then, the solution was purified by silica gel, Sephadex LH-20 and C_{18} column chromatography to obtain 17 compounds. Nine structures of compounds, identified according to physical and chemical properties using IR, UV, ESIMS, ¹H NMR, ¹³C NMR and other methods, include 3 petroleum ethers (β -sitosterol, oleanolic acid, ursolic acid), 4 chloroforms (β -sitosterol, daucosterol, syringaresinol, ethyl- β -D-pyran fruit glycosides), 1 ethyl acetate (1,5-O-two caffeoylquinic acid) and *n*-butanol parts (succinic acid and 5-hydroxy maltol). Oleanolic acid, ursolic acid, clove lignans, ethyl- β -D-pyran fruit glycoside, succinic acid and 5-hydroxy maltol were first isolated from burdock.

Key Words: Burdock root, Petroleum ether, Chloroform, Ethyl acetate, n-Butanol.

INTRODUCTION

Burdock, an herb belonging to the Asteraceae family, is a traditional Chinese medicine with high food value and medicinal benefits. A number of scholars have focused on the pharmacological activity and chemical composition of the edible burdock root.

The pharmacological activities of burdock root include antibacterial^{1,2}, antifungal³, antimutagenic⁴, antitumor⁵, antioxidant liver⁶, hypoglycemic⁷ and promotion of plant growth as well as resistance induction⁸. Fresh burdock root is chemically composed of *ca*. 70 % water, 2.8 % protein, 25 % carbohydrate and 0.6 % ash. Burdock root contains mostly inulin, amino acids, sulphur-acetylene class, multi-polyacetylenes, polyphenols and volatile oil, among others.

Although there are several reports on the pharmacological activity of burdock root, few studies have addressed the chemical composition, separation, purification and structural identification of the chemical composition of burdock root.

Currently, only a few details on the chemical composition and structure has been identified in burdock root, including essential oils, polyphenols, polyacetylenes, sulphur polyacetylenes and inulin. Majority of the chemical composition of this plant have not been purified. In the present study, identification of the chemical composition, purification and structure of the burdock root was performed.

EXPERIMENTAL

The burdock root medicinal pieces were from Qingdao Hao Tian Food Co. Ltd. (China).

Extraction and isolation: Burdock root herbs (20 kg) were obtained and extracted twice with two volumes of 95 % ethanol at 50 °C for 6 h. Ethanol was then recovered and the concentrated solution was combined and extracted. The extract was mixed with distilled water and then extracted by petroleum ether, chloroform, ethyl acetate and *n*-butanol to produce four parts with different polarities. These parts were concentrated to obtain 98 g petroleum ether, 100 g chloroform, 80 g ethyl acetate and 300 g *n*-butanol.

Petroleum ether was eluted by petroleum ether-acetone $(100:0 \rightarrow 0:100 \text{ gradient})$ on a silica gel column. At the 90:10 eluent, compound **1** (20 mg) was obtained, whereas compounds **2** (54 mg) and **3** (46 mg) were obtained at 85:15 eluent.

Chloroform was eluted by petroleum ether-acetone (100:0 \rightarrow 0:100 gradient) on a silica gel column. At the 85:15 eluent, compounds **1** (63 mg) and **4** (44 mg) were obtained. The eluate (60:40) was collected, concentrated and further separated with Sephadex-LH 20 reversed-phase column. Compound **5** (48 mg) was obtained at 40 %. The eluate (50:50) was collected, concentrated and further separated with C₁₈ reversed-phase column. Compound **6** (55 mg) was obtained at 15 %.

The ethyl acetate was eluted with chloroform-methanol (100:0 \rightarrow 0:100 gradient) on silica gel column. The eluate (50:50) was collected, concentrated and further separated using Sephadex-LH 20 reversed-phase column. Compound **7** (74 mg) was obtained at 15 %.

n-Butyl alcohol was eluted with chloroform-methanol (100:0 \rightarrow 0:100 gradient) on silica gel column. Compound **8** (52 mg) was obtained at 15 %. At 92:2 eluent ratio, compound **8** (52 mg) was obtained, whereas compound **9** (62 mg) was obtained at the 94:6 eluent ratio.

RESULTS AND DISCUSSION

The structures of nine compounds isolated from burdock root were shown in Fig. 1. Compound **1** was a white needle crystal (petroleum ether), with m.p. from 135-137 °C. Based on its purple colour at 10 % H_2SO_4 ethanol solution and Liebermann-Burchard positive test, the compound may either be triterpenoid or steroidal type. The sample and standard products were separated by thin layer chromatography (TLC), where a single spot was shown in three different systems. The R_f values of the compound were the similar to those of standard product when coloured in iodine vapour and 10 % H_2SO_4 ethanol solution and the melting point of the mixture did not decrease. Therefore, the compound was identified as β -sitosterol.

Compound **2** was a white needle crystal (acetic ether). In the chloroform solution, the Liebermann-Burchard reaction was positive, at the same time, based on the ¹H and ¹³C NMR spectral characteristics, the compounds may be the neat trick fruit type five-ringed triterpenes. ESI-MS m/z values were 479.2 ([M + Na]⁺, excimer ion peak), 457.2 ([M + H]⁺, excimer ion peak) and 439.2 ([M-H₂O + H]⁺). The relative molecular weight was 456 and combined with ¹H and ¹³C NMR spectra, the formula may be $C_{30}H_{48}O_3$. In the ¹H NMR (C_5D_5N , 600 MHz) map, seven single-peak methyl proton signals were shown, as follows: δ : 0.88 (3H, s), 0.94 (3H, s), 0.99 (3H, s),



1.02 (3H, s), 1.02 (3H, s), 1.21 (3H, s), 1.27 (3H, s). A proton signal linked with carbon oxygen 3.44 (1H, dd, J = 9.6 Hz, 6.0 Hz) and one alkene hydrogen proton signal. 5.49 (1H, s); ¹³C NMR (C₅D₅N, 150 MHz): 38.8 (C-1), 28.2 (C-2), 77.9 (C-3), 39.8 (C-4), 55.7 (C-5), 17.3 (C-6), 33.2 (C-7), 39.4 (C-8), 48.0 (C-9), 37.2 (C-10), 23.6 (C-11), 122.5 (C-12), 144.7 (C-13), 42.1 (C-14), 28.2 (C-15), 23.7 (C-16), 46.6 (C-17), 41.9 (C-18), 46.4 (C-19), 30.9 (C-20), 34.1 (C-21), 33.1 (C-22), 28.6 (C-23), 16.5 (C-24), 15.4 (C-25), 17.3 (C-26), 26.1 (C-27), 180.1 (C-28), 33.5 (C-29), 23.6 (H-30). The ¹³C NMR data were the similar to those of a previous report⁹. Both R_f values were exactly the same when the compound and oleanolic acid control were separated with TLC. Thus, the compound was identified as oleanolic acid.

Compound **3** was a white needle crystal (acetic ether). In the chloroform solution, the Liebermann-Burchard reaction was positive. ESI-MS m/z values were 479.2 ([M + Na]⁺, excimer ion peak), 457.2 ([M + H]+, excimer ion peak) and 439.2 ($[M - H_2O + H]^+$). At the same time, based on the ¹H and ¹³C NMR spectral characteristics, the compounds may be ursane five-ringed triterpenes. The relative molecular weight was 456 and the formula was $C_{30}H_{48}O_3$. In the ¹H NMR (C_5D_5N , 600 MHz) map, seven methyl proton signals could be seen, consisting of five single and two twin peaks, as follows: δ : 0.88 (3H, s), 0.95 (3H, d, J = 6.6 Hz), 0.99 (3H, d, J = 4.8 Hz),1.00 (3H, s), 1.02 (3H, s), 1.22 (3H, s), 1.24 (3H, s). One proton signal linked with carbon oxygen 3.46 (1H, dd, J =10.3 Hz, 6.6 Hz) and one alkene hydrogen proton signal 5.49 (1H, s). ¹³C NMR (C₅D₅N, 150 MHz): 39.0 (C-1), 28.0 (C-2), 78.0 (C-3), 39.3 (C-4), 55.7 (C-5), 18.7 (C-6), 33.5 (C-7), 39.6 (C-8), 48.0 (C-90), 37.3 (C-10), 17.4 (C-11), 125.5 (C-12), 139.2 (C-13), 42.4 (C-14), 28.7 (C-15), 24.8 (C-16), 48.0 (C-17), 53.4 (C-18), 39.8 (C-19), 39.4 (C-20), 31.0 (C-21), 37.3 (C-22), 28.7 (C-23), 15.6 (C-24), 16.5 (C-25), 17.4 (C-26), 23.5 (C-27), 179.8 (C-28), 23.8 (C-29), 21.3 (H-30). The ¹³C NMR data were similar to those of a previous report¹⁰. The $R_{\rm f}$ values were exactly the same when the compound and the ursolic acid control were separated with TLC. Therefore, the compound was identified as ursolic acid.

Compound **4** was a white needle crystal (80 % EtOH), m.p. > 300 °C. The Liebermann-Burchard reaction was positive and the colour was purple when dyed with 10 % H₂SO₄ ethanol solution. IR (KBr, v_{max} , cm⁻¹): produced 3432, 2936, 1636, 1540, 1461, 1383 and 1025 absorption peaks, which were similar with the IR values. The R_f values of the compound were the same as the daucosterol standard when they were separated with TLC and the melting point of the mixture did not decrease. Therefore, the compound was identified as daucosterol.

Compound **5** was colourless crystalline (methanol), where m.p. is from 177-178 °C, $[\alpha]_D^{20} = -3.36^\circ$ (c = 0.24, chloroform). ESI-MS showed m/z values of 441.1 ([M + Na]⁺), 417.0 ([M - H]⁺) and 859.4 ([2M + Na]⁺) excimer ion peak. Combined with ¹H and ¹³C NMR data, the relative molecular mass of the compound was determined as 418, molecular formula was C₂₂H₂₆O₈ and its unsaturation was 10. In the ¹H NMR (CDCl₃, 600 MHz) map, δ : 3.92 (12H, s, 4 × OCH₃) was the hydrogen signal on four 7-hydroxy, δ : 6.61 (4H, s) was the hydrogen signal on benzene rings and δ : 3.12 (2H, m, H-8 H-8'), δ : 3.92 (2H, m, H-9e, H-9'e), δ : 4.31 (2H, m, H-9a, H-9'a), δ : 4.75 (2H, d, J = 3.0 Hz, H-7, H-7'), δ : 5.58 (2H, br s, 4-OH, 4'-OH) proton signals. Combined with δ 86.1 (C-7, C-7'), δ 71.8 (C-9, C-9') and δ 54.3 (C28, C28'), three carbon signals in the ¹³C NMR spectrum suggest that the compound may be epoxy lignin. The fragrant area of ¹³C NMR spectrum appeared at δ 102.6 (C-2, C-2', C-6, C-6'), 132.1 (C-1, C-1'), 134.2 (C-4, C-4') and 147.1 (C-3, C-3', C-5, C-5') carbon signals, suggesting the symmetry of the molecular structure. The above data were similar to those of a previous report¹¹, identifying the compound as syringaresinol.

Compound 6 was colourless, crystalline and granular (aquiferous ethanol). Molisch reaction and fennel aldehyde acid reaction were positive. ESI-MS m/z were 439.9 ([M + M + Na]⁺, excimer ion peak), 230.8 ([M + Na]⁺, excimer ion peak), 209.0 ($[M + H]^+$ excimer ion peak), 179.9 $[M-C_2H_5]^+$. In the ¹H NMR (D_2O , 600 MHz) map, the mutual coupling of proton signal in two groups could be seen, δ 1.05 (3H, t, J =7.2 Hz, H-2') and δ 3.66 (2H, q, J = 7.2 Hz, H-1'). The compound contained CH₃-CH₂-O-segment, where 14.5 (C-2 ') and 56.6 (C-1') could be seen in the ${}^{13}C$ NMR (D₂O, 150 MHz) map data and further confirmed that the compound included the CH₃-CH₂-O segment. Five even oxygen carbon signals (69.5 [C-3], 69.1 [C-4], 68.1 [C-5], 63.7 [C-6] and 61.2 [C-1]) and a ketal carbon signal 100.5 (C-2) were also observed in the carbon map. These data were similar to those of a previous report¹², identifying the compound as ethyl- β -D-pyranoid fructoside.

Compound 7 was yellow powder with positive FeCl₃ reaction. The ¹H NMR data were (600 MHz, DMSO- d_6) δ : 7.48 (1H, d, J = 15.6 Hz, H-7"), 7.37 (1H, d, J = 15.6 Hz, H-7'), 7.10 (1H, s, H-2"), 7.05 (1H, s, H-2'), 6.96 (1H, d, J = 7.8 Hz, H-6"), 6.93 (1H, d, J = 7.8 Hz, H-6'), 6.75 (1H, d, J = 7.8 Hz, H-5"), 6.73 (1H, d, J = 7.8 Hz, H-5'), 6.25 (1H, d, J = 15.6 Hz, H-8"), 6.18 (1H, d, J = 15.6 Hz, H-8'), 5.26 (1H, m), 4.04 (1H, m) and 3.53 (1H, m). The ¹³C NMR data were (125 MHz, m)DMSO-d6) & 81.8 (C-1), 34.7 (C-2), 68.6 (C-3), 72.1 (C-4), 70.3 (C-5), 37.4 (C-6), 174.0 (C-7), 125.6 (C-1), 125.2 (C-1'), 115.8 (C-2'), 115.9 (C-2"), 143.6 (C-3'), 144.6 (C-3"), 148.1 (C-4'), 148.8 (C-4"), 114.9 (C-5', C-5"), 120.1 (C-6'), 121.2 (C-6"), 145.8 (C-7', C-7"), 114.2 (C-8"), 165.0 (C-9') and 166.2 (C-9"). The physical and chemical properties, as well as spectral data, were similar to those of a previous report¹³, identifying the compound as 1-O-, 5-O-dicaffeoylquinic acid.

Compound **8** consisted of colourless particles (methanol) with negative ESI-MS m/z values, namely, 116.8 ([M-H]⁻ excimer ion peak) and 101.0 [M + H - H₂O]⁺. The relative molecular mass was 118. In the ¹H NMR (DMSO-*d*₆, 600 MHz) map, two groups of single-peak proton signals could be seen, with a ratio of 1:2, δ : 2.42 (4H, s) and 12.15 (2H, s). A carboxyl and aliphatic carbon was shown in the ¹³C NMR (DMSO-*d*₆, 150 MHz) mapping data and both R_f values were exactly the same when the compound and succinic acid standard product were separated with TLC. Therefore, the compound was identified as succinic acid.

Compound **9** was a light yellow crystal with m.p. from 218-222 °C and showed deep purple in TLC with FeCl₃ solution at 105 °C. ESI-MS m/z values were 142. 9 $[M + H]^+$ and 164.8 $[M + Na]^+$. The relative molecular mass was 142.

¹H NMR data were (600 MHz, DMSO- d_6) δ : 8.96 (1H, br s, -OH), 8.79 (1H, br s, -OH), 7.95 (1H, s, H-6), 2.23 (3H, s, CH₃) and ¹³C NMR data were (125 MHz, DMSO- d_6) δ : 168.9 (C-4), 149.6 (C-2), 144.7 (C-5), 141.8 (C-3), 139.4 (C-6) and 14.8 (C-7). The physical and chemical properties, as well as spectral data, were similar to those of a previous report¹⁴, identifying the compound as 5-oxymaltol.

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REFERENCES

- F.B. Holetz, G.L. Pessini, N.R. Sanches, D.A. Cortez, C.V. Nakamura and B.P. Filho, *Mem. Inst. Oswaldo Cruz.*, 97, 1027 (2002).
- M. Takasugi, S. Kawashima and N. Katsui, *Phytochemistry*, 26, 2957 (1987).

- 3. L.C. Chiang, W. Chiang, M.Y. Chang, L.T. Ng and C.C. Lin, *Antiviral Res.*, **55**, 53 (2002).
- K. Miyamoto, M. Nomura, M. Sasakura, E. Matsui, R. Koshiura, T. Murayama, T. Furukawa, T. Hatano, T. Yoshida and T. Okuda, *Cancer Res.*, 84, 99 (1993).
- 5. C. Tamayo, M.A. Richardson, S. Diamond and I. Skoda, *Phytother*. *Res.*, **14**, 1 (2000).
- 6. S.C. Lin, C.H. Lin, C.C. Lin, Y.H. Lin, C.F. Chen, I.C. Chen and L.Y. Wang, *J. Biomed. Sci.*, **9**, 401 (2002).
- 7. M. Mitsuo, Y. Nobuo and T. Katsuya, J. Oleo. Sci., 54, 589 (2005).
- 8. F.D. Wang, G.H. Feng and K.S. Chen, *Postharvest Biol. Technol.*, **52**, 110 (2009).
- 9. H. Kojinna and H. Ogura, Phytochemistry, 25, 729 (1986).
- 10. D.T. Coxon and J.W. Wells, *Phytochemistry*, **19**, 1247 (1980).
- M.M. Badawi, S.S. Handa, A.D. Kinghorn, G.A. Cordell and N.R. Farnsworth, J. Pharm. Sci., 72, 1285 (1983).
- H.X. Kuang, R. Kasai, K. Ohtani, Z.S. Liu, C.S. Yuan and O. Tanaka, *Chem. Pharm. Bull.*, **37**, 2232 (1989).
- 13. Z.J. Jia, Y. Zhao and R.X. Tan, J. Nat. Prod., 56, 494 (1993).
- Y. Shinoda, M. Murata, S. Homma and H. Komura, *Biosci. Biotechnol. Biochem.*, 68, 529 (2004).