



Chromones from the Flowers of *Rosa rugosa* and Their Cytotoxicity

LIYING YANG¹, JIANHUA YAO², LIDAN SHU¹, YANQIONG SHEN^{1,2}, XUEMEI GAO^{1*} and QIU-FEN HU¹

¹Key Laboratory of Chemistry in Ethnic Medicinal Resources, State Ethnic Affairs Commission & Ministry of Education, Yunnan University of Nationalities, Kunming 650031, P.R. China

²Key Laboratory of Tobacco Chemistry of Yunnan Province, Yunnan Academy of Tobacco Science, Kunming 650106, P.R. China

*Corresponding author: Fax: +86 871 5910017; Tel: +86 871 5910013; E-mail: gao_xuemei@hotmail.com

(Received: 25 October 2012;

Accepted: 21 August 2013)

AJC-13949

A new chromone, 2,2-dimethyl-4-oxo-8-(2-oxopropyl)-chroman-6-yl acetate (**1**), together with five known chromones (**2-6**), were isolated from the flowers of *Rosa rugosa*. The structures of **1-6** were elucidated by spectroscopic methods including extensive 1D and 2D NMR techniques. Compounds **1** were evaluated for their cytotoxicity against five human tumor cell lines and it showed high cytotoxicity against A549 and MCF7 cell with IC₅₀ values of 3.6 and 5.0 μM, respectively.

Key Words: Chromones, *Rosa rugosa*, Isolation, Structure elucidation, Cytotoxicity.

INTRODUCTION

The species of *Rosa rugosa* are one of the most important ornamental flowers widely distributed in temperate regions of eastern Asia including Japan, Korea and China¹. This species had widely been cultivated in several areas of Yunnan Province because of its high commercial values². Meanwhile, the petals and buds of *R. rugosa* have also been used as food, incense materials and in traditional Chinese medicine for treating stomachache, diarrhoea and women's diseases³. The recent study also report the previous phytochemical researches on *R. rugosa* has revealed that tannins⁴, sesquiterpenes⁵, terpenoids^{6,7}, as well as flavonoids^{8,9} are major components isolated from this plant. The biological activities of antitumour had also been reported for recent study on this plant^{7,9,10}. Motivated by a search for new bioactive metabolites from this plant, our group had reinvestigated the chemical constituents of the flowers of *R. rugosa*, which led to the isolation and characterization of a new chromone (**1**), along with five known chromones (**2-6**). The structures of the isolated compounds were established by means of spectroscopic methods including extensive 1D and 2D NMR techniques. This paper deals with the isolation, structural characterization and biological activities of these compounds.

EXPERIMENTAL

General experimental procedures: Optical rotations were measured with a Horiba SEPA-300 polarimeter. UV spectrometry was obtained using a Shimadzu UV-2401A spectro-

photometer. 1D and 2D NMR spectrometry were recorded on a DRX-500 NMR spectrometer with TMS as internal standard. Unless otherwise specified, chemical shifts (δ) are expressed in ppm with reference to the solvent signals. HRESIMS was performed on a VG Autospec-3000 spectrometer. Semi-preparative HPLC was performed on a Shimadzu LC-8A preparative liquid chromatograph with Zorbax PrepHT GF (21.2 mm × 25 cm) or Venusil MP C₁₈ (20 mm × 25 cm) columns. Column chromatography was performed using silica gel (200-300 mesh, Qing-dao Marine Chemical, Inc., Qingdao, People's Republic of China), Lichroprep RP-18 gel (40-63 μm, Merck, Darmstadt, Germany), Sephadex LH-20 (Sigma-Aldrich, Inc, USA) and MCI gel (75-150 μm, Mitsubishi Chemical Corporation, Tokyo, Japan). The fractions were monitored by TLC and spots were visualized by heating silica gel plates sprayed with 5 % H₂SO₄ in EtOH.

The flowers of *R. rugosa* were collected in Dali Prefecture, Yunnan Province, People's Republic of China, in September 2010. The identification of the plant material was verified by Prof. Chen Y.J. (Yunnan Nationalities University). A voucher specimen (YNNI 10-9-56) has been deposited in our laboratory.

Extraction and isolation: The air-dried and powdered flower buds of *R. rugosa* (5.2 kg) were extracted four times with 95 % aqueous methanol (4 × 5 L) at room temperature and filtered. The filtrate was evaporated under reduced pressure and the crude extract (465 g) was applied to silica gel (150-200 mesh) column chromatography, eluting with a CHCl₃-MeOH gradient system (20:1, 9:1, 8:2, 7:3, 6:4, 5:5), to give

six fractions A-F. The further separation of fraction B (9:1, 32.8 g) by silica gel column chromatography, eluted with petroleum ether-acetone (9:1-1:2), yielded mixtures B1-B6. Fraction B2 (8:2, 6.47 g) was subjected to silica gel column chromatography using petroleum ether-ethyl acetate and semi-preparative HPLC (52-55 % MeOH-H₂O or 40-45 % CH₃CN-H₂O, flow rate 12 mL/min) to give **1** (11.3 mg), **2** (25.4 mg) and **5** (20.1 mg). Fraction B3 (7:3, 6.22 g) was subjected to silica gel column chromatography using petroleum ether-ethyl acetate and semi-preparative HPLC (45-50 % MeOH-H₂O or 36-40 % CH₃CN-H₂O, flow rate 12 mL/min) to give **4** (18.4 mg) and **6** (13.6 mg). Further separation of fraction D (8:2, 31.8 g) by silica gel column chromatography, eluted with petroleum ether-acetone (8:2-1:2), yielded mixtures D1-D5. Fraction D3 (6:4, 8.22 g) was subjected to silica gel column chromatography using petroleum ether-ethyl acetate and semi-preparative HPLC (35-38 % MeOH-H₂O or 28-35 % CH₃CN-H₂O, flow rate 12 mL/min) to give **3** (21.4 mg).

2,2-Dimethyl-4-oxo-8-(2-oxopropyl)-chroman-6-yl acetate (1): Obtained as pale yellow oil; UV (MeOH) max (log ϵ) 210 (4.26), 262 (3.92), 355 (2.87) nm; IR (KBr, ν_{\max} , cm⁻¹): 3435, 2914, 2876, 1716, 1680, 1668, 1612, 1550, 1439, 1356, 1132, 941, 853; ¹H and ¹³C NMR data (CDCl₃, 500 and 125 MHz), see Table-1; negative ESIMS m/z 313 [M + Na]⁺; negative HRESIMS m/z 313.1044 [M + Na]⁺ (calcd. (%) for C₁₆H₁₈NaO₅, 313.1052).

	δ_c (m)	δ_H (m, J, Hz)
2	79.7 s	–
3	50.7 t	2.67 s
4	192.2 s	–
5	112.0 d	7.42, d, J = 2.4
6	145.7 s	–
7	123.9 d	7.07, d, J = 2.4
8	130.8 s	–
9	154.2 s	–
10	120.1 s	–
11	49.6 t	4.04 s
12	206.3 s	–
13	30.2 q	2.22 s
14,15	25.2 q	1.49 s
-OAc	21.1 q, 169.6 s	2.03 s

RESULTS AND DISCUSSION

The flowers of *R. rugosa* were extracted with 95 % methanol. The extract was subjected repeatedly to column chromatography on silica gel, Sephadex LH-20, RP-18 and semi-preparative RP-HPLC separation to afford seven new chromone, 2,2-dimethyl-4-oxo-8-(2-oxopropyl)-chroman-6-yl acetate (**1**), together with five known chromones (**2-6**). The structures of the compounds **1-6** were as shown in Fig. 1 and the ¹H and ¹³C NMR data of the compound **1** were listed in Table-1. The known compounds, compared with literature data, were identified as, peuceenin-7-methyl ether (**2**)¹¹, mulberroside C (**3**)¹², greveichromenol (**4**)¹³, 6-(3-hydroxy-4-methoxystyryl)-4-methoxy-2H-pyran-2-one (**5**)¹⁴, pestaloficial J (**6**)¹⁵.

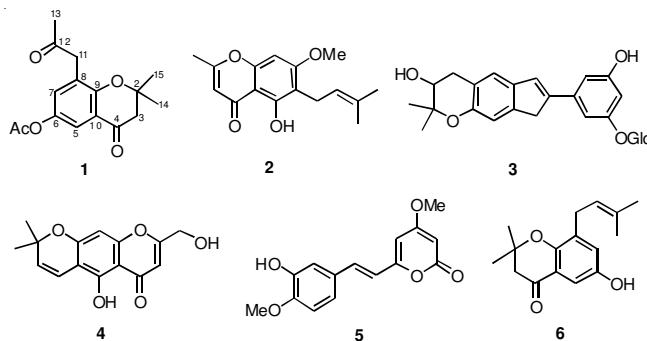


Fig. 1. Structure of chromones from the flowers of *Rosa rugosa*

Compound **1** was obtained as pale yellow oil. It gives a parent ion by HR-ESIMS at m/z 313.1044 [M + Na]⁺ (calcd. (%) for 313.1052) corresponding to a molecular formula C₁₆H₁₈O₅, requiring eight degrees of unsaturation. The ¹H and ¹³C NMR spectra of compound **1** along with analysis of the DEPT spectra (Table-1) displayed 16 carbon signals and 18 proton signals, respectively, corresponding to a chromanone nucleus¹⁴ (δ_c 79.7 s, 50.7 t, 192.2 s, 112.0 d, 145.7 s, 123.9 d, 130.8 s, 154.2 s, 120.1 s, 25.2 q (2C)), an acetylonyl group (-CH₂C(O)CH₃) (δ_c 49.6 t, 206.3 s, 30.2 q; δ_H 4.04 s, 2.21 s) and an acetoxy group (δ_c 21.2 q, 169.6 s; δ_H 2.03 s). Strong absorption bands accounting for hydroxy (3435 cm⁻¹), carbonyl group (1716, 1680, 1668 cm⁻¹) and aromatic groups (1612, 1550, 1439 cm⁻¹) could also be observed in its IR spectrum. The UV spectrum of compound **1** showed absorption maxima at 262 and 210 nm, which confirmed the existence of the aromatic functions. The HMBC correlations (Fig. 2) of H-11 (δ_H 4.04) with C-7 (δ_c 123.9), C-8 (δ_c 130.8) and C-9 (δ_c 154.2), of H-7 (δ_H 7.07) with C-11 (δ_c 49.6) indicated that the acetylonyl group should be located at C-8 on the chromone ring (Fig. 2). The position of acetylonyl group was determined, the acetoxy group should be located at C-6 to support the ¹H NMR signals of aromatic proton (δ_H 7.42, d, J = 2.4 and 7.07, d, J = 2.4). Thus, the structure of **1** was established as 2,2-dimethyl-4-oxo-8-(2-oxopropyl)-chroman-6-yl acetate.

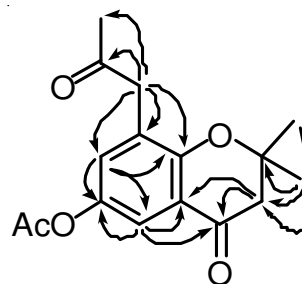


Fig. 2. Key HMBC correlations (\curvearrowright) of **1**

Since certain chromone derivatives exhibit potential cytotoxicity^{16,17}, the compound **1** was tested for their cytotoxicity against five human tumor cell lines (NB4, A549, SHSY5Y, PC3 and MCF7) using the MTT method as reported previously¹⁸. The results showed that compound **1** showed high cytotoxicity against A549 and MCF7 cell with IC₅₀ values of 3.6 and 5.0 μ M, respectively.

ACKNOWLEDGEMENTS

This project was supported financially by the Excellent Scientific and Technological Team of Yunnan High School (2010CI08), the Yunnan University of Nationalities Green Chemistry and Functional Materials Research for Provincial Innovation Team (2011HC008) and Open Research Fund Program of Key Laboratory of Ethnic Medicine Resource Chemistry (Yunnan University of Nationalities) (2010XY08).

REFERENCES

1. S.M. Gault and P.M. Syngé, *The Dictionary of Roses in Colour*, Ebury Press, London (1971).
2. J.L. Lu, *China Flowers Horticult.*, **11**, 26 (2012).
3. L. Putian and Y. Jiang, *Flora of China*, Chinese Science Press, Beijing (1977).
4. S. Ochir, B. Park, M. Nishizawa, T. Kanazawa, M. Funaki and T.J. Yamagishi, *Nat. Med.*, **64**, 383 (2010).
5. Y. Hashidoko, S. Tahara and J. Mizutani, *Phytochemistry*, **32**, 387 (1993).
6. Y. Hashidoko, *Phytochemistry*, **43**, 535 (1996).
7. H.J. An, I.T. Kim, H.J. Park, H.M. Kim, J.H. Choid and K.T. Lee, *Int. Immunopharm.*, **11**, 504 (2011).
8. Z.P. Xiao, H.K. Wu, T. Wu, H. Shi, B. Hang and H.A. Aisa, *Chem. Nat. Comp.*, **42**, 736 (2006).
9. X.M. Gao, L.Y. Yang, L.D. Shu, Y.Q. Shen, Y.J. Zhang and Q.F. Hu, *Heterocycles*, **85**, 1925 (2012).
10. M.H. Ma, B. Cui, H.F. Yu and C.Z. Yuan, *J. Shandong Inst. Light. Ind.*, **22**, 38 (2008).
11. S. Thadaniti, W. Archakunakorn, P. Tuntiwachwuttikul and J.B. Bremner, *J. Sci. Soc. Thailand*, **20**, 183 (1994).
12. T. Fukai, Y. Hano, K. Hirakura, T. Nomura and J. Uzawa, *Chem. Pharm. Bull.*, **33**, 4288 (1985).
13. F.M. Dean and M.L. Robinson, *Phytochemistry*, **10**, 3221 (1971).
14. N.T. Dat, X.J. Jin, Y.S. Hong and J.J. Lee, *J. Nat. Prod.*, **73**, 1167 (2010).
15. L. Liu, S.C. Liu, S.B. Niu, L.D. Guo, X.L. Chen and Y.S. Che, *J. Nat. Prod.*, **72**, 1482 (2009).
16. D.R. Mou, W. Zhao, Tao Zhang, L. Wan, G.Y. Yang, Y.K. Chen, Q.F. Hu and M.M. Miao, *Heterocycles*, **85**, 1251 (2012).
17. A. Lazarenkow, J. Nawrot-Modranka, E. Brzezinska, U. Krajewska and M. Rozalski, *Med. Chem. Res.*, **21**, 1861 (2012).
18. T. Mosmann, *J. Immunol. Methods*, **65**, 55 (1983).