# **UV Induced Evolution of Amino Acids under Possible Primitive Conditions of the Earth**

## LATA JOSHI

Department of Chemistry Kumaun University Campus, Almora-263 601, India

Amino acids *viz.* lysine, aspartic acid, serine, glycine, glutamic acid, threonine, α-alanine, β-alanine, α-amino-butyric acid, valine, isoleucine and leucine were formed as a result of irradiation of aqueous sterilized solution of allyl alcohol and aminonia with UV light of wavelength 2537Å. The presence of oxides *viz.*, ZrO<sub>2</sub>, TiO<sub>2</sub>, colloidal Fe<sub>2</sub>O<sub>3</sub>, MnO<sub>2</sub> and colloidal. MoO<sub>3</sub> exerted a marked effect on the nature and yield of the photo products.

## INTRODUCTION

Abiotic synthesis of possible precursors of life forming molecules has been studied by a number of workers. <sup>1-4</sup> Formation of amino acids from intermediates viz., saturated alcohols, aldehydes and acids<sup>5-9</sup> is also reported. However, synthesis of amino acids from unsaturated alcohols has not received much attention. The present work attempts to investigate UV induced synthesis of amino acids from reaction mixtures of allyl alcohol-ammonia and water in the presence and absence of various oxides.

#### MATERIALS AND METHODS

Low pressure mercury vapour lamp (250 w) was used to provide UV light of wavelength 2537 Å. Quartz glass vessels of 300 mL capacity were placed in specially designed air cooled chamber at room temperature (18  $\pm$  2°C). Prolonged time periods of irradiation upto 40 h were used. The mixtures in the present study were sterilized solutions of allyl alcohol in double distilled water (1/100 v/v) at pH  $\pm$  0.5. Oxide sensitizers (5 mg/100 mL water or 0.5 mL/100 mL water) were tested for their purity prior to addition in the reaction system. Portions of irradiated solutions were taken out aseptically after 10, 20, 30 and 40 h exposure, concentrated in vacuum evaporation and analysed chromatographically on Whatman No. 1 paper both by uni- and two-dimensional chromatography 10 using *n*-butanol-acetic acid-water (4 : 1 : 1 v/v) and *n*-butanol-acetic acidpyridine-water (15 : 3 : 10 : 12 v/v) as solvents. 11 Ninhydrin reagent 12 was used to locate amino acid spots on chromatogram. Further identification was made by colour tests with a variety of reagents viz., isatin, 13 Folin's reagent 14 and alloxan; 15 R<sub>f</sub> value measurements and also by spectral methods. 16 Products were eluted for chemical

402 Joshi Asian J. Chem.

tests by thin layer chromatography on silica gel G.<sup>17</sup> Photochem calorimeter MKIII was used for quantitative determination of amino acids. UV absorption spectrum of irradiated mixture was recorded in aqueous medium in Beckman DK-2 spectrophotometer.

#### RESULTS AND DISCUSSION

Tables 1 and 2 show the results of irradiation of allyl alcohol-water-ammonia mixture for different periods of time up to 40 h. Eight chromatographically detectable and ninhydrin positive products are found in 10 h exposure. Three more ninhydrin positive products are added in further 10 h irradiation. After 30 h exposure, fourteen products were detected. Further increase of irradiation time up to 40 h shows marked decrease in product yields. Of the fourteen products so formed, twelve were identified as lysine (I), aspartic acid (II), serine (IV), glycine (V), glutamic acid (VI), threonine (VII),  $\alpha$ -alanine (VIII),  $\beta$ -alanine (IX),  $\alpha$ -amino butyric acid (XI), valine (XII), isoleucine (XIII) and leucine (XIV). Relatively good yield for products IV, V, VIII, IX, XI, XII and XIV is also seen in Table 1. Products II, V, VIII and XII were also confirmed by UV absorption spectroscopy (Fig. 1).

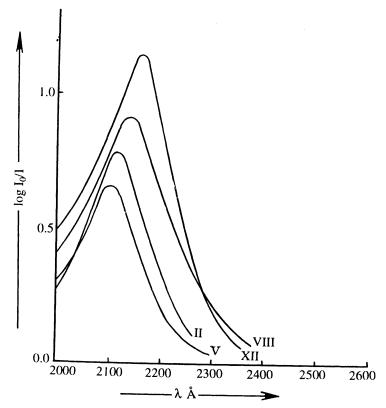


Fig. 1. UV absorption spectra of some photo products of irradiated aqueous solution of allyl alcohol and ammonia (30 h)

EFFECT OF UV RADIATION ON AQUEOUS STERILIZED SOLUTION OF ALLYL ALCOHOL AND AMMONIA (PH  $10\pm0.5$ )

Composition of Exposure	Exposure						Produc	Products of irradiation	adiation						
properties	(h)	_	=	Ξ	>	>	I/	VII	VIII	×	×	IX	IX	XIII	ΧIX
allyl alcohol +	10			F	+	+	+	F	‡	+					F
ammonia + water	20	1	۳	+	‡	‡	+	+	‡	- ‡		+		<b> -</b>	<b>→</b> +
	30	<b>+</b>	‡	+	‡	+ + +	+	<b>+</b>	+ + + +	† †	‡	‡	1	. 1	. 44
	40	+	+	+	‡	+	⊢	+	‡	‡	: +	: +	:	<u>-</u>	+ +
R <sub>f</sub> (%) in solvent										• •	-	-		-	-
systems (a) B.A.W.		8.8 8.8	12.2	13.2	14.0	15.2	18.0	20.0	22.2	24.0	25.2	31.4	44.0	52.2	56.4
(b) B.A.P.W.		38.0	40.0	42.4	43.0	4.0	50.0	52.0	54.0	56.1	58.0	62.4	4	8 2 9	74.0
Colour with										· ·	5		2	0	?
(a) Ninhydrin		>	BV	>	>	RV	>	>	BV	>	>	>	>	. >	>
(b) Folin's reagent		GBr	Ċ	Ŋ	GBr	GY	BG	Ö	Ö	Ö	. <sub>U</sub>	ت .	GrBr	· ლ	خ
(c) Isatin		BrP	ф	фB	Д	BrP	BB	Ы	ф	ф	ρ.	. д	RrP	RrP	RP .
(d) Alloxan		RV	ф	~	×	×	œ	OR	æ	O. S.	. ~	. e	£ 25	¥	ξ α
Stability in										;	:	5	Í	Í	٤
(a) ether		ins	ins	ins	ins	ins	ins	ins	ins	ins	ins	ins	ins	ins	Sui
(b) water	s	s	s	s	s	s		s	s	s	s	y.	v	v	
Amino acid identified		Lysine	Aspartic acid	1	Serine	Glycine	Glutamic acid	Threo- nine	α-alan- ine	β-alan- ine	α-amino acid	α-butyric acid	valine	iso- leucine	leucine

V, violet; B, blue; R, red: Br, brown: P, purple; G, green; Gr. gray; Y, yellow; O. orange; d, dull: ir.s. insoluble; s, soluble; —, not detected/ unidentified; T, trace amount; +, low yield: ++, moderate yield; +++, good yield; ++++, high yield.

B.A. W, *n*-butanol: acetic acid: water as 4: 1: 1 vol/vol at 16 ± 2°C.

B.A.P.W., *n*-butanol : acetic acid : pyridine: water as 15 : 3 : 10 : 14 vol./vol. at  $16 \pm 2^{\circ}$ C.

TABLE-2 DETERMINATION (g/1000 mL) OF PRODUCTS FORMED BY UV IRRADIATION (30 h)OF ALLYL ALCOHOL-AMMONIA-WATER SYSTEMS (PH =  $10.0 \pm 0.5$ ) IN THE PRESENCE AND ABSENCE OF OXIDES

Productions of irradiation

Reaction mixture						P	Productions of irradiation	of irradiat	ion					
MIN N	-	II	H	>1	>	N	VII	VIII	×	×	ΙX	IIX	шх	XIX
no oxide	0.002	0.006	0.003	0.056	0.048	0.005	0.028	0.126	0.140	0.035	0.062	0.061	0.048	0.072
$ZnO_2$	۲	۲	0.001	0.002	0.003	T	0.001	0.020	1	Н	1	1	Т	1
TiO <sub>2</sub>	0.003	0.033	0.015	0.064	0.031	0.046	0.025	0.072	0.080	0.050	0.042	L	0.038	Т
Colloidal Fe <sub>2</sub> O <sub>3</sub>	0.002	0.036	0.033	0.076	0.048	0.042	0.050	0.080	0.084	0.073	0.040	H	0.022	H
$MnO_2$	⊢	0.034	Т	0.042	0.032	0.018	0.028	0.065	0.038	0.074	0.026	⊢	0.025	H
Colloidal MoO <sub>3</sub>	0.001	0.001	0.003	0.076	0.064	0.125	0.129	0.095	0.135	0.226	0.025	0.061	0.079	0.080
—, not detected; T, trace amount; coll., colloidal	ace amou	nt; coll., ce	olloidal.											

A marked effect was noticed on the photochemical synthesis of amino acids when identical solutions of allyl alcohol, ammonia and water were irradiated separately in the presence of oxides. Photochemical formation of amino acids was retarded when ZrO2 was used as sensitizer. Products IX, XI, XII and XIV could not be detected and the other photo products showed faint presence only. Presence of TiO<sub>2</sub> in the reaction mixture gave all the fourteen products (Table 2). Of these serine (IV), α-alanine (VIII) and β-alanine (IX) were formed in appreciable amounts. Products glycine (V), glutamic acid (VI), α-amino butyric acid (XI) and isoleucine (XIII) were formed in moderate amounts whereas the others were formed in smaller amounts. In the presence of colloidal Fe<sub>2</sub>O<sub>3</sub>, same photo products were formed in moderate amounts. Products XII and XIV were formed in traces. Presence of MnO2 showed a moderate effect (Table 2). Highest yield of photo products was observed in the presence of colloidal MoO<sub>3</sub>.

Thus it may be inferred that allyl alcohol appeared as an intermediate during the photochemical evolution of amino acids under primitive earth conditions. The catalytic role of mineral oxides then present is also important.

## ACKNOWLEDGEMENT

Financial assistance from the UGC and CSIR, New Delhi is gratefully acknowledged. The author also thankfully acknowledges the facilities provided by the Kumaun University, Nainital.

# REFERENCES

- 1. J. Oro, Nature, 197, 756 (1963).
- 2. O.N. Perti and H.D. Pathak, *Proc. Natl. Acad. Sci.*, **30A**, 206 (1961).
- P.C. Joshi and H.D. Pathak, Science and Culture, 41, 260 (1975).
- 4. A.S.U. Choughuley, A.S. Subbaraman and Z.A. Kazi, *Blosystems*, 9, 73 (1877).
- 5. K. Bahadur, Nature, 173, 1141 (1954).
- 6. O.N. Perti and V. Paul, Agra Univ. J. Res. Sci., 13, 9 (1964).
- 7. L. Melkani, V.K. Melkani and H.D. Pathak, Acta Ciencia Indica, 9C, 35 (1983).
- 8. C.K. Pathak, J.N.Pant and H.D. Pathak, J. Brit. Interplanetary Soc. (London), 32, 103 (1978).
- 9. K.H. Arada snd T. Iwasaki, *Nature*, **250** (5465), 426 (1974).
- 10. T.L. Hardy, D.O. Holland and J.N.C. Nayler, Anal. Chem., 27, 971 (1955).
- 11. I.M. Hais and K. Macke, Paper Chromatography, Acad. Press, N.Y., p. 437 (1963).
- 12. I. Simth, Chromatographic and Electrophoretic Techniques, Vol. 1, Interscience Publishers, N.Y., p. 110 (1969).
- 13. J. Noworytko and M. Sarnecka-Keller, Acta Biochem. Polon., 2, 91 (1965).
- 14. K.U. Giri and A. Nagabhushanum, *Naturwiss*, **39**, 501 (1956).
- 15. A. Saifer and I. Oreskes, *Anal. Chem.*, **28**, 501 (1956).
- 16. J.P. Greenstein and M. Winitz, Chemistry of Amino Acids, Vol. 1-3, John Wiley and Sons, N.Y. (1961).
- 17. E. Stahl, Thin Layer Chromatography, Acad. Press, N.Y. (1965).