Palladium(II) Complex derived from Harmaline; an Alkaloid Isolated from *Peganum harmala* Seeds: Synthesis, Characterization and Cytotoxic Activity

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The 1:1 molar ratio reaction of trans-[Pd(DMSO)₂Cl₂] (I) with harmaline, the naturally occuring alkaloid, isolated from Peganum harmala seeds, in non-aqueous solvent, was found to give, mainly, the monomer trans-[Pd(harmaline)(DMSO)Cl₂] (II) as a major component and the dimer [Pd(harmaline)Cl₂]₂ (III) as a minor one. The monomer (II) can be formed, as a single product, either by the direct reaction between I and harmaline in DMSO or by dissolving II in DMSO. In both cases the pure monomer (II) can be precipitated from its DMSO solution by the addition of 0.05 M HCl. The complexes have been characterized by their C, H, N analysis, ¹H, ¹³C, IR and mass spectroscopic methods. The monomer (II) has been tested for its cytotoxic activity against several tumour cell lines, i.e., P₃₈₈, L₁₂₁₀, K₅₆₂, Raji, HeLa, RD and Hep-2. This activity was compared with those of reference standards, cisplatin, carboplatin and 5-fluorouracil (5-FU).

INTRODUCTION

In our recent work on the palladium(II) complexes, we have given a brief survey about the importance of platinum complexes as antitumour agents and reported the first antileukaemic *trans*-palladium(II) complex with 7-methoxy-1-methyl-9H-pyrido[3,4-b]indole (harmaline); a naturally occurring β -carboline alkaloid, isolated from *Peganum harmala* seeds. As a continuation of our comprehensive interest in the biological evaluation of metal cmplexes with *e.g.*, β -carboline alkaloids, $^{2-5}$ we are presenting here another new *trans*-palladium(II) complex derived from 4,9-dihydro-7-methoxy-1-methyl-3H-pyrido[3,4-b]indole (harmaline) as another naturally occurring β -carboline alkaloid, isolated from *Peganum harmala* seeds. The present study also includes the examination of this complex against several tumour cell lines, both the fluid suspension cell lines (P₃₈₈, K₅₆₂, L₁₂₁₀ and Raji) and the solid ones (HeLa, RD and Hep-2).

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EXPERIMENTAL

¹H and ¹³C NMR spectra were recorded at the University of Jordan, Amman, Jordan, on a Burker-DPX 300 MHz spectrometer using DMSO-d₆ as a solvent with TMS as an internal standard. IR spectra were recorded on Nicolet FT-IR (Impact 400) spectrometer, using KBr discs in the range 4000–400 cm⁻¹ or CsI discs in the range 4000–200 cm⁻¹ on SP 2000 Pye Unicam spectrophotometer. Elemental analyses were performed at the Atlantic Microlab, Inc., Norcross, Georgia-30091 (USA). Mass spectrum of complex (II) was performed at Sussex University, Brighton, UK, using FAB technique with cold inlet and 3-nitrobenzoyl alcohol (3-NBA) as a solvent.

Starting Materials

The compounds K_2PdCl_4 , $PdCl_2$ and harmaline were commerical products (Fluka) and used without further purification. The complex *trans*- $[Pd(DMSO)_2Cl_2]$ was prepared as described in our previous article. ¹

Preparation of trans-[Pd (DMSO)(harmaline) Cl₂]

The complex trans-[Pd(DMSO)₂Cl₂] (0.5 g, 1.5 mmol) was suspended in acetone (40 mL) and placed in a 150 mL two-necked round-bottomed flask fitted with water condenser. A suspension of harmaline (0.32 g, 1.5 mmol) in acetone (40 mL) was gradually added to the above supension. The reaction mixture was immediately turned clear orange; it was left under reflux for ca. 3 h, then filtered through celite. The clear orange solution was taken to dryness to give an orange residue. This was redissolved in a small amount of DMSO (to destroy the dimeric product if there is any) and the product was precipitated by 0.05 M hydrochloric acid solution. The solid was collected by filtration, washed with water and air dried first, then under vacuum at 60°C for several hours. The yield was 0.55 g (78%).

This complex can also be prepared by dissolving the complex *trans*-[Pd(DMSO)₂Cl₂] in DMSO with gentle heating and after cooling the solution to room temperature it was treated with 0.05 M HCl until complete precipitation. The solid thus formed was treated as above. In both cases, the solid can be recrystallized from CHCl₃/n-hexane to give fine orange crystals.

Biological Methods

- (1) Complex: The complex (II) was dissolved in 10% DMSO. Serial dilutions of 0.1, 1.0 and 10.0 μ g/mL were used and millipore (0.2 μ m) filtered under laminar flow conditions. Reference standards (cisplatin, carboplatin and 5-FU) were purchased from Bristol Myers (USA).
- (2) Cell lines: Several cell lines were used in the present study; these include both the fluid suspension cell lines (e.g., P₃₈₈, K₅₆₂, L₁₂₁₀ and Raji) and solid cell lines (e.g., HeLa, RD and Hep-2). The fluid suspension cell lines were maintained in RPMI-1640 and supplemented with 5% fetal calf serum, L-glutamine and antibiotics, whereas the solid cell lines were maintained in Dulyeccos minimum

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essential mdium (DEME) and supplemented with 5% fetal calf serum, L-glutamine and antibiotics.

(3) Cytotoxicity test: MTT(3-[4,5-dimethylthiazol-2-yl]-2,5-diphenyltetrazolium bromide) colorimetric assay was performed in a 96-well plate^{6,7}. The above cell lines $(2 \times 10^4 \text{ cells/mL})$ were seeded in each well with 100 μ L of the growth medium and antibiotics, using the method described in our previous work.³

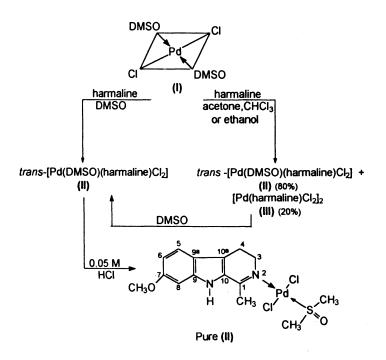
RESULTS AND DISCUSSION

The preparation of the starting material trans-[Pd(DMSO)₂Cl₂] and its reaction with harmaline are summarized in Scheme 1. The complex trans-[Pd harmaline) (DMSO) Cl₂] (II) was isolated from its DMSO solution and purified as described in the experimental part. The physical properties of II are listed in Table-1.

SCHEME 1 THE PREPARATIVE ROUTE FOR PURE (II)

$$K_2PdCl_4 + DMSO \longrightarrow trans-[Pd(DMSO)_2Cl_2] + K[Pd(DMSO)_3Cl] + KCl$$

$$PdCl_2 + DMSO \longrightarrow trans-[Pd(DMSO)_2Cl_2]$$
(I)



Colour

m.p. (°C)

¹H NMR‡

13C NMR‡

Results Method of analysis Orange > 146 with decomposition IR (cm⁻¹)* v(N-H) = 3311m; v(C-H) = 2996, 2919m; v(C-C) = 1633s; v(C=N) = 1556s; v(S=O) = 1121s; v(Pd-Cl) = 320mFound (%): C = 38.6, H = 4.5, N = 6.1; calculated (%) for Elemental analysis $C_{15}H_{20}N_2O_2Cl_2SPd$: C = 38.5, H = 4.3 N = 6.0Mass spectrum† m/z = 470, 460, 435, 391, 370, 215, 136, 69 $\delta(CH_3) = 3.2s (3H); \delta(CH_3O) = 3.8s (3H) \delta(NH) = 11.6b (1H);$ $\delta(HC-3) = 3.0d$ (1H), (J = 31) and 3.9m (1H); $\delta(HC-4) = 2.9b$ (2H); $\delta(HC-5) = 6.8dd$ (1H) (J = 1.8); $\delta(HC-6) = 7.5dd$ (1H) (J = 2.8);

TABLE-1 PHYSICAL PROPERTIES AND ANALYSES OF COMPLEX (II)

 $\delta(HC-8) = 6.9b(1H)$.

 $\delta(C-1) = 164.4$; $\delta(C-3) = 52.9$, $\delta(C-4) = 19.8$, $\delta(C-5) = 118.3$, $\delta(C-6) = 112.5$, $\delta(C-7) = 159.3$,

 $\delta(C-10a) = 122.1$, $\delta(CH_3) = 24.6$, $\delta(CH_3O) = 55.8$

 $\delta(C-8) = 94.7$, $\delta(C-9) = 139.7$, $\delta(C-9a) = 119.0$, $\delta(C-10) = 128.5$,

We have previously reported that the reaction of trans-[Pd(DMSO)₂Cl₂] with β-carboline alkaloids isolated from Peganum harmala seeds leads to the formation of various palladium(II)-alkaloid complexes, both the monomer and the dimer.² Later studies of ours on such complexes, but with platinum metal, using the FAB-mass spectroscopic technique, showed that two products could be detected the monomer (with one coordinated DMSO molecule) and the dimer (without DMSO).8 Recently, we have found that palladium metal gave almost certainly similar products when trans-[Pd(DMSO)2Cl2] was treated with harmine¹, the monomer trans-[Pd(harmine)(DMSO)Cl₂] (30% proportion) and the dimer [Pd(harmine)Cl₂]₂ (70% proportion). In the present study, the reaction of trans-Pd(DMSO)₂Cl₂ with harmaline was found to give, again, the monomer and the dimer (Scheme-1) but with ca. 80% and 20% proportions, respectively, i.e., opposite trend to that of harmine one. This may be due to the fact that harmaline possesses some saturation on C-3 and C-4, which in turn allows some structural modification and this prefers the formation of the monomer, in contrary to harmine which does not possess such a structural flexibility, i.e., planar molecule.

However, the characterization data are clearly assigned to the formal trans-[Pd(harmaline)(DMSO)Cl₂] (II). The DMSO molecule has been coordinated via sulphur and this is very clear from the strong IR band appearing at 1121 cm⁻¹. The rest of other spectral data obtained from ¹H, ¹³C and mass techniques are in a good agreement with those expected for complex II and compare very well with those obtained for the harmine analogue complex.¹

The cytotoxic activity of complex II (Scheme-1) against the different cell lines

^{*}Selected IR bands; s. strong; m. medium bands.

[†]FAB technique used with cold inlet, using 3-NBA as a solvent; molecular ion m/z = 469.4.

[‡]Downfield from internal TMS at room temperature, using DMSO-d₆ as a solvent; s, singlet; dd, doublet of doublet; m, multiplet; b, broad signals

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compared with those of the reference standards are shown in Table-2. It appears from these results that complex II exhibited different cytotoxic activities against all the cell lines. However, this complex showed certain selectivity especially towards the fluid suspension cell lines (P_{388} , K_{562} , L_{1210} and Raji), where the IC₅₀ values were ranging from 0.25–0.45 µg/mL. This activity is somewhat approaching to the IC₅₀ values of cisplatin against the same cell lines and superior to those of carboplatin where the IC₅₀ values were greater than 10 µg mL against the same cell lines. Furthermore, complex II showed no cytotoxic activities against the solid cell lines (HeLa, RD and Hep-2) at concentrations \leq 10 µg/mL.

TABLE-2
CYTOTOXIC ACTIVITIES OF COMPLEX II WITH STANDARD REFERENCES
AGAINST DIFFERENT TUMOUR CELL LINES

Compound -	IC ₅₀ (μg/mL)						
	P ₃₈₈	L ₁₂₁₀	K ₅₆₂	Raji	HeLa	RD	Нер-2
(II)	0.35	0.45	0.40	0.25	>10	> 10	>10
Cisplatin	0.15	6.00	0.25	NT	5.5	> 10	1.8
Carboplatin	> 10	> 10	> 10	> 10	> 10	> 10	>10
5-FU	0.15	NT	0.15	NT	NT	NT	NT

NT: Not tested.

Preliminary, the selectivity of complex II against the leukaemic cell lines (P_{388} , K_{562} and L_{1210}) and the Buerkitts lymphoma cell lines (Raji) is very interesting and promising. Nevertheless it is premature to conclude about such activity unless otherwise confirmed by *in vivo* tests in animal models. Therefore, further studies are necessary to confirm the above activity.

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