

Juncenolide B, a New Briarane Diterpene from the Gorgonian *Junceella juncea* of the Indian Ocean

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Chemical examination of the Indian Ocean gorgonian, *Junceella juncea* Pallas (Gorgonaceae) furnished a new non-chlorinated briarane diterpene, Juncenolide-B (1). The structure of 1 was determined as (1S, 2S, 7S, 8R, 9S, 10S, 11R, 12R, 13R)-11,20-epoxy-13-(3-methylbutanoyl)-2,9,13-triacetoxylbriaran-4-en-18,7-olide on the basis of ^1H and ^{13}C NMR, DEPT, COSY and FABMS experiments.

Key Words: Briarane, *Junceella juncea*, Juncenolide-B, Gorgonian, Indian Ocean.

INTRODUCTION

In continuation of our search for bioactive secondary metabolites from gorgonians¹, we have investigated *Junceella juncea* Pallas (Gorgonaceae) collected from the Tuticorin coast (8°45' E, 78°12' N) of the Indian Ocean. Previous studies on gorgonian corals of the genus *Junceella* have resulted in the isolation of structurally novel and highly oxygenated briarane-type diterpenes, which contain a γ -lactone in a bicyclo-[8,4,0] system²⁻⁹. Among these metabolites some briaranes showed potential biological activities such as antitumor, anti-inflammatory, antiviral and antibacterial activities¹⁰⁻¹³. We report herein the isolation and characterization of a new non-chlorinated diterpene (1S, 2S, 7S, 8R, 9S, 10S, 11R, 12R, 13R)-11,20-epoxy-13-(3-methylbutanoyl)-2,9,13-triacetoxylbriaran-4-en-18,7-olide named Juncenolide-B(1).

EXPERIMENTAL

Melting points were measured on a Boitus melting pont apparatus and are uncorrected. Optical rotations were taken on an Autopol polarimeter. Purification of the compound was done on Shimadzu preparative RPHPLC system with LC-8A pump, SPD-10A detector with Selectosil C8, 5 μ , 110 A (10 \times 250 mm) column. IR spectra were recorded on a Perkin-Elmer BXFTIR spectrophotometer. ^1H , ^{13}C NMR, DEPT and COSY spectra were recorded using an AMX 400 MHz NMR instrument. TLC was carried out on silica gel-G (ACME) thin layers.

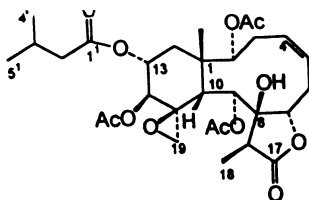
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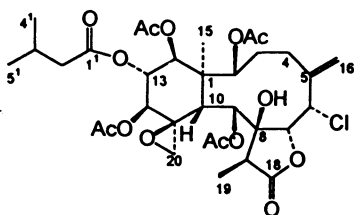
Animal Material: The gorgonian *Junceela juncea* (Pallas) was collected from Tuticorin coast (8°45' E, 78°12' N) of the Indian Ocean in March 1999. It was identified by Dr. P.A. Thomas, CMFRI, Trivandrum. A voucher specimen (AU2-164) was deposited in the Dept. Of Pharmaceutical Sciences, Andhra University, Visakhapatnam, Andhra Pradesh, India.

Extraction and Isolation: The collected organism was washed thoroughly with fresh water at the site and shade dried. The outer red layer of the gorgonian (dry wt. 1 kg) was carefully separated, powdered and successively extracted using n-hexane (5 L), dichloromethane (5 L) and methanol (5 L).

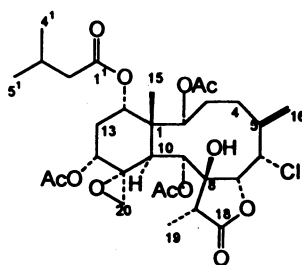
A white solid (1.2 g) separated out on concentration of the dichloromethane extract. The solid on column chromatography over silica gel (100–200 #) yielded an impure sample of the compound 1 along with Gemmacolide-B (1 g) and Juncin-H (165 mg) already reported⁸. The compound 1 was further purified by preparative RPHPLC (water : acetonitrile, 40 : 60) yielded pure sample (8 mg).



Compound 1



(+)- Gemmacolide B



Juncin H

RESULTS AND DISCUSSION

Juncenolide-B (1): It was obtained as colourless crystals, m.p. 213°C, $[\alpha]_D^{25} - 14.0^\circ$ (c 1.0, CHCl₃); its m.f. of C₃₀H₄₂O₁₁ is derived from a molecular ion at m/z 579 [M + H]⁺ in the FABMS. Its IR bands indicated the presence of a tertiary hydroxyl (3435 cm⁻¹), a five membered lactone (1765 cm⁻¹) and an ester carbonyl (1731 cm⁻¹).

The ¹H NMR data of compound 1 (Table-1) showed three singlets (δ 2.23, 3H; 2.02, 3H; 2.01, 3H) of methyls on O-acetyl groups, a typical methyl doublet (δ 1.26, J = 7.1 Hz), a methyl singlet (δ 1.24), a pair of doublets (δ 2.38, J = 3.4 Hz, 2H-19), a methine singlet (δ 3.65, H-10) and five oxygenated methine protons (δ 5.91, d, J = 8.4 Hz, H-2; 5.79, brs, H-9; 4.92, m, H-13; 4.60 brs, H-12; 4.44,

brs, H-7), two olefinic protons (δ 5.25, m, H 4, H 5) and a tertiary hydroxyl (δ 3.45, brs, 8-OH). The ^{13}C NMR and DEPT spectra of compound 1 showed signals for six methyl carbons, five methylene carbons and eight quaternary carbons including five ester carbons (δ 169.358, 169.769, 170.266, 171.358, 174.395).

TABLE-1
 ^1H AND ^{13}C NMR SPECTRAL DATA OF JUNCENOLIDE-B (1) IN CDCl_3 , TMS AS REFERENCE AT 400 MHz; CHEMICAL SHIFT (δ), J IN Hz

Position	^{13}C	^1H	COSY
1	47.271 s	—	—
2	73.081 d*	5.91 (d, 8.4)	H-3
3	28.296 t	2.74 (m, 2H)	H-2, H-4
4	120.624 d*	5.25 (m)	H-3, H-5
5	121.148 d*	5.25 (m)	H-4, H-6
6	28.443 t	2.45 (m)	H-5, H-7
7	71.836 d	4.44 (brs)	H-6
8	81.435 s		
9	81.219 d	5.79 (brs)	H-10
10	51.445 s	3.66 (brs)	H-9
11	56.849 s		
12	73.655 d*	4.60 (brs)	H-13
13	72.746 d*	4.92 (m)	H-12, H-14
14	29.474 t	2.15 (d, 6)	H-13
15	14.219 q	1.24 (brs)	
16	35.400 d	2.94 m	H-18
17	174.395 s		
18	5.982	1.26 (d, 6.8)	H-16
19	50.769	2.38 (d, 3.4) 2.89 (d, 3.4)	
2-OAc	169.358 s*, 20.587 q*	2.23 (s)	
9-OAc	169.769 s*, 20.832 q*	2.02 (s)	
12-OAc	170.266 s*, 21.214 q*	2.01 (s)	
1 ¹	171.358 s*		
2 ¹	43.356 t	2.10 (s)	H-3'
3 ¹	24.990 d	2.08 (s)	H-2', H-4', H-5'
4 ¹	22.706 q	0.92 (d, 6.4)	H-3'
5 ¹	22.403 q	0.92 (d, 6.4)	H-3'
8-OH		3.45 s	

Multiplicities are assigned on the basis of DEPT spectrum.

*Close δ_c values may be interchangeable.

A study of mass spectrum and ^{13}C NMR showed that there is no chlorine bearing carbon atom in compound 1. It is observed that in most of the C6 chlorinated briarane diterpenes the chlorine bearing carbon atom resonates at about δ 53.6 and the characteristic methine proton appears at δ 4.6^{7,8}. Absence of these resonances both in ^{13}C and ^1H NMR indicates the absence of chlorine atom at C6 in compound 1. Further the presence of an additional methylene at δ 28.44 besides the methylene at C-3 and C-13/C-14 in the rings and the appearance of multiplet at δ 2.45 (2 H) in ^1H NMR confirms the absence of chlorine atom in compound 1. Thus compound 1 shows close structural similarity with a briarane Gemmacolide-B, but differs from it by lack of chlorine atom at C-6, exocyclic methylene at C-5 and absence of a fourth O-acetyl group. The position of the double bond is fixed as follows.

In general it is noted that the exocyclic methylene carbons in briarane diterpenes appear at δ 121.1, δ 146.6 and the methylene protons resonate at δ 5.50, δ 5.80 as doublets^{7,8}. In compound 1 olefinic carbons appeared at δ 120.62, δ 121.14 and their protons resonated at δ 5.25 (2H) as multiplet. This shows the absence of exocyclic methylene in compound 1. Further the appearance of ^{13}C resonances at δ 120.62 and δ 121.14 indicates the presence of same environment for the two carbons of the double bond. This is possible only if the double bond is Δ^4 . This observation is confirmed by the connectivities of H-3/H-4/H-5/H-6/H-7 using ^1H - ^1H COSY spectrum.

The FABMS fragmentation showed the presence of three acetoxy and one isovalerate group in compound 1. The positions and stereochemistry of three O-acetyl groups could be ascertained by detailed study and comparison of ^1H , ^{13}C and ^1H - ^1H COSY data with those of Juncin-H and Gemmacolide-B. The position of the isovalerate could be on C-13/C-14; if present on C-14, the methylene protons of C-13 appear as multiplet at δ 2.20 as observed in Juncin-H.⁸ The compound 1 has showed a doublet at δ 2.15 attributed to 14-H₂ coupled to 13-H and 13-H as a multiplet at δ 4.9. The ^1H - ^1H COSY connectivities of H-12/H-13/H-14 also add to this view. Based on the above data the structure of compound 1 has been determined as (1S, 2S, 7S, 8R, 9S, 10S, 11R, 12R, 13R)-11, 20-epoxy-13-(3-methylbutanoyl)-2,9,13-triacetoxylbriaran-4-en-18,7-olide.

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REFERENCES

1. K.V. Ramana, D.V. Rao, C.B.S. Rao, E. Fahy and D.J. Faulkner, *J. Nat. Prod.*, **51**, 954 (1988).
2. Y. Lin and K. Long, *Zhongshan Daxue Xuebao, Ziran Kexueban*, **2**, 46 (1983).
3. K. Long, Y. Lon and W. Huang, *Zhongshan Daxue Xuebao, Ziran Kexueban*, **2**, 15 (1987); *Chem. Abstr.*, **107**, 215049m (1987).
4. J. Shin, M. Park and W. Fenical, *Tetrahedron*, **45**, 1633 (1989).
5. B.F. Bowden, J.C. Coll and G.M. Konig, *Aust. J. Chem.*, **43**, 151 (1990).
6. S. Isaacs, S. Carmely and Y. Kashman, *J. Nat. Prod.*, **53**, 596 (1990).
7. H.Y. He and D.J. Faulkner, *Tetrahedron*, **47**, 3271 (1991).
8. A.S.R. Anjaneyulu and N.S.K. Rao, *J. Chem. Soc., Perkin Trans.*, **1**, 959 (1997).
9. M. Garcia, J. Rodriguez and C. Jimenez, *J. Nat. Prod.*, **62**, 257 (1999).
10. J. Rodriguez, R.M. Nieto and C. Jimenez, *J. Nat. Prod.*, **61**, 313 (1998).
11. Y.C. Shen, C.V.S. Prakash, Y.T. Chang, M.C. Hung, S.J. Chen, H.J. Chen and M.C. Hsu, *Chin. Pharm. J.*, **52**, 341 (2000).
12. Y.C. Shen, C.V.S. Prakash and Y.T. Chang, *Steroids*, **66**, 721 (2001).
13. J.H. Kwak, F.J. Schmitz and G.C. Williams, *J. Nat. Prod.*, **64**, 754 (2001).

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