

Amino Acid Composition of Some New Varieties of Oil Seeds

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The amino acid composition of some new varieties of oil seeds (*Arachis hypogaea* JGN-3, *Brassica campestris* VARUNA, *Carthamus tinctorius* JSF-1, *Glycine max* JS-90-41 and *Helianthus annuus* KBSH-1) were analyzed by high performance liquid chromatography.

Key Words: Amino acid composition, High performance liquid chromatography, *Arachis hypogaea* JGN-3, *Brassica campestris* VARUNA, *Carthamus tinctorius* JSF-1, *Glycine max* JS-90-41 and *Helianthus annuus* KBSH-1.

INTRODUCTION

Food consists of three main classes of natural products: carbohydrates, proteins and fat, apart from the essential ingredients such as vitamins and minerals, while proteins supply the building materials of the body for its growth and for proper and balanced functioning of the various important organs. Protein deficiency in babies and young children causes not only physical impairment, but also permanent and irreversible brain damage¹. As a result, protein-calorie malnutrition has become one of the most outstanding dietary problems in India and other developing countries². From a survey of the conventional and new proteins used in various food preparations, it will be evident that to supplement protein intake, maximum use will have to be made of oilseed proteins. Oilseeds are an excellent source of protein and energy for improving cereals³.

Proteins present in various foods differ in their nutritive value on account of the difference in the amino acid composition. An analysis for amino acid by modern methods has given an excellent nutritive evaluation of the absorbed fraction of the food protein and thus, for this reason, one must be concerned with qualitative and quantitative adequacy of amino acids⁴.

EXPERIMENTAL

The oil seeds (*Arachis hypogaea* JGN-3, *Brassica campestris* VARUNA, *Carthamus tinctorius* JSF-1, *Glycine max* JS-90-41 and *Helianthus annuus* KBSH-1) were procured from Department of Plant Breeding and Genetics, Jawaharlal Nehru Krishi Vishwavidyalaya, Jabalpur.

The samples were hydrolyzed in 6 N hydrochloric acid for 18 h at 110°C. The hydrolysate was analysed by JASCO high performance liquid chromatograph UV-975 amino acid analysis system.

RESULTS AND DISCUSSION

From the perusal of the data (Table-1), it appears that in the seed proteins of *Arachis hypogaea* JGN-3, *Brassica campestris* VARUNA, *Glycine max* JS-90-41 and *Helianthus annuus* KBSH-1 the percentage of aspartic acid was maximum (84.27, 82.91, 14.83 and 89.27 g/100 g proteins respectively), in the seed proteins of *Carthamus tinctorius* JSF-1 the percentage of glutamic acid was maximum (14.83 g/100 g proteins), in the seed proteins of *Brassica campestris* VARUNA valine (0.02 g/100 g proteins), in the seed proteins of *Carthamus tinctorius* JSF-1 phenyl alanine (0.15 g/100 g proteins), in the seed proteins of *Glycine max* JS-90-41 threonine (0.95 g/100 g proteins) and in the seed proteins of *Helianthus annuus* KBSH-1 tyrosine (0.04 g/100 g proteins) were present in lowest percentage.

TABLE-1
AMINO ACID COMPOSITION OF SOME NEW VARIETIES OF OIL SEEDS

Amino acid	Amino acid analysis (g/100 g proteins)				
	<i>Arachis hypogaea</i> JGN-3	<i>Brassica campestris</i> VARUNA	<i>Carthamus tinctorius</i> JSF-1	<i>Glycine max</i> JS-90-41	<i>Helianthus annuus</i> KBSH-1
Aspartic acid	84.27	82.91	3.22	14.83	89.27
Glutamic acid	3.40	12.86	25.35	11.99	0.33
Serine	6.56	0.60	7.67	10.06	4.55
Glycine	0.20	0.11	14.41	2.68	0.59
Histidine	0.10	0.66	11.26	2.83	3.11
Alanine	1.03	0.32	17.14	3.33	0.23
Arginine	3.01	0.22	10.19	2.12	0.22
Threonine	0.09	0.11	1.01	0.95	0.09
Proline	0.10	—	0.28	1.00	0.25
Lycine	0.02	0.03	3.43	5.76	0.15
Tyrosine	0.06	0.08	0.49	8.98	0.04
Valine	0.21	0.02	2.43	10.41	0.05
Methionine	0.06	0.06	0.29	4.56	0.08
Cystine	0.09	—	—	9.29	0.17
Isoleucine	—	0.04	—	—	—
leucine	0.26	0.19	—	—	—
Phenylalanine	0.30	0.80	0.15	11.11	0.16

Isoleucine has not been reported in the seed proteins of *Arachis hypogaea* JGN-3. The seed proteins of *Arachis hypogaea* JGN-3 were found to contain highest amount of aspartic acid. The values obtained in the present studies are

considerably higher than those reported earlier by various investigators⁵⁻⁷. However, other amino acids in the decreasing order were serine, glutamic acid, arginine, alanine, phenyl alanine, leucine, valine, glycine, histidine, proline, threonine, cystine, tyrosine, methionine and lycine.

Brassica campestris VARUNA is lacking in proline and cystine. The seed protein of *Brassica campestris* VARUNA was found to contain highest amount of aspartic acid (82.91 g/100 g proteins) followed by glutamic acid (12.86 g/100 g proteins). Other amino acids in the decreasing order were phenylalanine, histidine, serine, alanine, arginine, leucine, threonine, glycine, tyrosine, methionine, isoleucine, lycine and valine. A study of amino acid composition of the seed proteins of *Brassica campestris* VARUNA as reported in the present study and by various workers⁸⁻¹⁰ reveals that considerable varietal variations exist.

Carthamus tinctorius JSF-1 is lacking in isoleucine and leucine. Phenylalanine (0.15 g/100 g proteins) is somewhat sacrificed in the seed proteins of *Carthamus tinctorius* JSF-1. However, other amino acids in the increasing order were tyrosine, threonine, valine, aspartic acid, lycine, serine, arginine, histidine, glycine, alanine and glutamic acid. The amino acid composition of seed protein of *Carthamus tinctorius* JSF-1 was found to be in good accordance with reported values¹¹⁻¹³.

Glycine max JS-90-41 is lacking in isoleucine and leucine. Threonine (0.95 g/100 g proteins) is somewhat sacrificed in the seed proteins of *Glycine max* JS-90-41. However other amino acids in the increasing order were proline, arginine, glycine, histidine, alanine, methionine, lycine, tyrosine, cystine, serine, valine, glutamic acid and aspartic acid. However, not much variation exists with the reported values^{4, 14-17}.

Helianthus annuus KBSH-1 is lacking in isoleucine and leucine. Tyrosine, valine, methionine, threonine, lysine, phenylalanine and cystine are somewhat sacrificed in the seed proteins of *Helianthus annuus* KBSH-1. However, other amino acids in the increasing order were arginine, alanine, proline, glutamic acid, histidine, serine and aspartic acid. Considerable variation has been observed in the amino acids composition of seed proteins of *Helianthus annuus* KBSH-1¹⁸⁻²⁰.

There is much variation in the amino acid composition of seed proteins in the oilseeds of *Arachis hypogaea* JGN-3, *Brassica campestris* VARUNA and *Helianthus annuus* KBSH-1 under investigation. Variations in the various constituents of oilseeds have been attributed to the variety^{21, 22}, conditions, fertilizer treatments and climatic conditions²³⁻²⁵.

The present results seem to confirm the good quality of the protein in *Carthamus tinctorius* JSF-1 and *Glycine max* JS-90-41 as compared to other oilseeds.

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REFERENCES

1. C.R. Mitra, *Khadi Gramodyog*, **17**, 712 (1971).
2. Y.P. Gupta, *Intern., J. Trop. Agri.*, **5**, 247 (1987).
3. R.J. Bressani, *Am. Oil. Chem. Soc.*, **52**, 2541 (1975).
4. H.P. Saxena and S.K. Shrivastava, *Acta Cienc. Indica*, **23C**, 99 (1997).
5. M.M.S. Basha and S.K. Pancholy, *J. Agric. Food Chem.*, **29**, 331 (1981).
6. H.F. Jiang, N. Duan, H.F. Jiang and N.X. Duan, *Crop Genetic Resources*, **4**, 29 (1994).
7. C. Andres, *Food Processing*, **38**, 5 (1977).
8. F.X. Aherne and J.J. Kennelly, Oilseed meals for livestock feeding, in: D.J.A. Cole and W. Haresingh (Eds.), *Recent Development in Big Nutrition*, Butterworths, London, UK, p. 278 (1985).
9. Y.G. Liu, S.K. Jensen and B.O. Eggum, *J. Sci. Food Agric.*, **67**, 135 (1995).
10. B. Digna, R. Rodrige, J. Nakouzi, C.O. Chichester, E. Yanez and F. Monckeberg, *J. Sci. Food Agric.*, **21**, 139 (1970).
11. F.W. Sosulski and G. Sarwar, *J. Inst. Can. Technol. Aliment*, **1**, 6 (1973).
12. C.H. Van Etten, J.J. Rackis, T.W. Miller and A.K. Smith, *J. Agri. Food Chem.*, **11**, 137 (1963).
13. *The Wealth of India*, **3(Ca-Ci)**, 311 (1963).
14. S. Kuppaswami, M. Srinivasan and V. Subramanyan, Protein in Foods, Indian Council of Medical Research, Special Reports Series No. 33, pp. 50–51 (1958).
15. C. Gopalan, B.V. Ramashastri and S.C. Balasubramanian, Nutritive Value in Indian Foods, National Institute of Nutrition, Hyderabad (India), pp. 140–141 (1980).
16. M.N. Islam and R.A. Lea, *J. Food Sci.*, **46**, 658, 663 (1981).
17. S. Pushpalata and S.K. Shrivastava, *Asian J. Chem.*, **14**, 1071 (2002).
18. M.A. Sabir, F.W. Sosulski and S.L. Mackenize, *J. Agric. Food Chem.*, **21**, 988 (1973).
19. K.J. Smith, *Feed Stuffs*, **8**, 20 (1968).
20. J. Baerdet and J. Mosse, *J. Am. Oil Chem. Soc.*, **54**, 82A (1977).
21. W.D. Bressani, E. Marcucci, C.E. Robles and N.S. Schimsaw, *Food Res.*, **19**, 263 (1954).
22. S.J. Afroj, R.K. Bajpai, S.P. Netke and B.S. Gupta, *Indian J. Anim. Sci.*, **483**, 161 (1978).
23. R.J. Evans, J.L. St. John, P.M. Craven and S.P. Huddock, *Cereal Chem.*, **24**, 150 (1947).
24. G.C. Esh and J.M. Som, *Indian J. Physiol. All Sci.*, **7**, 158 (1956).
25. W.V. Royes, Nutr. Improve Food Legumes Breed Proc. Symp., pp. 193–196 (1975).

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