Effects of Flyash and Plant Hormones Treated Soil on the Increased Protein and Amino Acid Contents in the Seeds of Ground Nut (*Arachis hypogaea*)

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Soils of newly formed Chhattisgarh state are acidic (red, yellow) and soil acidity is responsible for Al³⁺ ion toxicity which leads to reduced micro organism activity, as well as Mo and Zn deficiency. In plants indole acetic acid and giberellic acid were used as growth hormones. Plant growth parameters were compared for plain and flyash and hormone treated soil.

Key Words: Flyash, *Arachis hypogaea*, Giberellic acid, Protein hydrolyzate, Indole acetic acid.

INTRODUCTION

Ground nut is an important oil seed crop of India. It covers an area of 7.4 million hectares with a production of 8.4 million tonnes. The yield and poor, quality of seed kernels are the major problems in ground nut production. Ground nut seeds contain about 50% edible oil and 50% of high quality of proteins¹. Indiscriminate use of NPK fertilizers removes trace elements fastly from the soils². Flyash has following properties. (i) It is basic in nature to ameliorate soil acidity, (ii) It is a reserve source of trace elements to replenish its deficiency in the soil and (iii) Its physicochemical properties, texture helps to mix homogeneously with soils³.

Biotechnologically, the present work on ground nut is unique in one aspect of its nutrition. The seeds develop *via* nutrients it gathers directly from the soil rather than those transported from roots to shoots and back to the seeds⁴. Aluminum ion toxicity is perhaps the most important cause of reduced plant growth. Flyash is basic in nature and a good source of NPK alongwith Cu, Zn, Mn, Mo and Fe. In the present work, soil of Arpa irrigation project area has been used for amelioration of soil acidity.

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EXPERIMENTAL

Flyash from NTPC Korba district of Chhattisgarh state is used to ameliorate the acidic, soil of Arpa irrigation project area. Soil, flyash and their various combinations were analyzed for their physico-chemical properties. Soil and flyash were mixed in different proportions and homogenized by grinding and seiving. Samples of different combinations were analyzed by the methods suggested by Hesse⁵.

About 4 kg sample of each combination were taken in pots. The details of each combination have been shown in the Table-1. The properties of flyash are summarized in Table-2.

TABLE-1 SOIL AND FLYASH COMBINATIONS

S. No.	Details of combinations				
1.	Plain soil	Α			
2.	90 % w/w soil + 10 % w/w flyash + NPK (400 : 600 : 100)	В			
3.	80 % w/w soil + 20 % w/w flyash + NPK (400 : 600 : 100)	С			
4.	70 % w/w soil + 30 % w/w flyash + NPK (400 : 600 : 100)	D			
	+ indole acetic acid + giberellic acid (0.001 M each)				
	sprayed periodically.				

TABLE-2 PROPERTIES OF ELVASH

PROPERTIES OF FLYASH					
Properties of flyash		Properties of flyash			
рН	8.795	$Fe_2O_3(\%)$	7		
Electrical conductivity (m mhos/cm)	0.172	$P_2O_5(\%)$	0.20		
$SiO_2(\%)$	62	$SO_3(\%)$	0.25		
$Al_2O_3(\%)$	18	CaO (%)	1.62		

The physico-chemical properties of different combinations are shown in the Table-3.

TABLE-3											
PE	PHYSICO-CHEMICAL PROPERTIES OF DIFFERENT COMBINATIONS										
		Electrical	Oxides and N conc. (%)				Trace elements				
Symbol	pН	conductivity	conc. (ppm)								
		(m mhos/cm)	SiO ₂	Al_2O_3	P_2O_5	CaO	Ν	Mn	Fe	Zn	Mo
А	6.25	0.082	77.00	15.00	0.15	1.19	0.006	47	40	54	3.5
В	6.40	0.085	77.50	15.50	0.18	1.49	0.008	48	42	60	3.9
С	6.52	0.090	77.55	16.00	0.20	1.50	0.006	50	42	55	4.5
D	6.88	0.120	78.00	17.00	0.22	1.60	0.005	52	45	60	4.6

Preparation of protein hydrolyzate

Oil was first removed from seeds of ground nut (*A. hypogaea* L.) using soxhlet extractor, using normal hexane as solvent. Proteins were isolated by repeated exhaustive extraction of defatted, decarbohydrated seeds, with

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cold, alkaline 10 % brine solution. The proteins were precipitated from the filtrate by addition of 10 % CH₃COOH and heating the solution at 80°C for 2 h.

Isolated protein (0.5 g) was refluxed with a mixture of 100 mL 6 N HCl and 90 % HCOOH for 10 h. Excess of acid was removed by evaporation. Volume was reduced to 5 mL and then further diluted with 25 mL of distilled water and filtered. Filtrate was vacuum dried and stored in a brown bottle under nitrogen atmosphere and sent for HPLC analysis⁶. Plant growth parameters have been displayed in Table-4.

PLANT GROWTH PARAMETERS OF GROUND NUT (A. hypogaea L.)						
	Root	Shoot	Leaf	Total chlorophyll	Vield per	Protein
Symbol	length	length	area	content (mg/g)	plant (g)	concentrate
	(cm)	(cm)	(cm^2)	content (Ing/g)	plant (g)	(%)
А	12	55	2090	1.70	15	55
В	14	60	2950	1.80	16	60
С	15	65	3550	1.90	17	65
D	17	70	4500	2.50	19	70

TABLE-4 PLANT GROWTH PARAMETERS OF GROUND NUT (A. hypogaga

HPLC studies of seeds of ground nut (*A. hypogaea* L.) from the plain soil (A) and harmone treated soil (D) show the presence of phenyl alanine, methionine, threonine, leucine and valine. This has been further confirmed by the molecular interactions of L-amino acids and iodine in 50 % aqueous ethanol solvent system as shown in the Table-5.

TABLE-5
ELECTRONIC ABSORPTION SPECTRA OF L-AMINO ACIDS-IODINE
SYSTEM IN 50 % AQUEOUS ETHANOL SOLUTION

	a	
S.	System of L-amino acid	
	and iodine in 50 %	λ_{\max} (absorbance)
N.	aqueous ethanol solution	
1.	Leucine	212 (0.6302), 297 (0.3590), 359 (0.3689)
2.	Phenyl alanine	212 (0.4558), 294 (0.2340), 352 (0.2352)
3.	Valine	212 (0.5870), 290 (0.1800), 358 (0.120)
4.	Threonine	212.4 (1.095), 291 (0.3400), 358 (0.190)
5.	Methionine	215 (0.345), 287 (0.360)

RESULTS AND DISCUSSION

The ground nut is unique in one aspect of nutrition. The seed develops *via* nutrients it gathers directly from the soil rather than those transported from roots to shoots then to seeds. Acidic soil requires liming for better crop yield and better seed kernel developments. In the case (D) the maximum yield is due to better calcium absorption by the pod⁷. Phosphorus

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may be the most deficient element on a global scale⁸. Molybdenum deficiency is more common in acid soils while molybdenum is required to increase nitrogen, methionine and other amino acids contents. It is essential for nodule formation⁹. Zinc deficiency occurs in acidic soil¹⁰.

Application of plant hormones indole acetic acid and giberellic acid are important to induce enzyme activities. Cell elongation, accompanied by increased DNA synthesis is due to GA application¹¹. Protein synthesis requires auxin induced cell enlargement¹².

Conclusively in the present work, the use of flyash has ameliorated soil acidity and help in availability of trace elements to the crop which has resulted increased plant growth parameters, protein and essential amino acid contents.

REFERENCES

- N.A. Kachot and D.D. Malviya, Integrated Nutrient Management for Improving Yield 1. and Quality of Ground Nut. Extended Summaries, 2nd International Agro. Congress, New Delhi, Vol. I, pp. 274-75 (2002).
- 2. V.M. Shorrocks, Micronutrient problems in South East Asia, Fertilizer International, Vol. 143 pp. 9, 11, 43, 45, 57 (1981).
- 3. G. Singh, Utilization of Flyash in Agriculture Report. No. T.R./CFRI/107/97, CFRI, Dhanbad, India (1997).
- 4. G.J. Gascho and J.G. Davis, in ed.: J. Smart, Mineral Nutrition in the Ground Nut Crop - A Scientific Basis for Improvement, Chapman & Hall, London, p. 214 (1994).
- 5. P.R. Hesse, ATB of Soil Chemical Analysis, A CBS Publication, Delhi, India (2001).
- N. Raman and J. Muthumuniassamy, Asian J. Chem., 12, 925 (2000). 6.
- W.K. Robertson, H.W. Lundy and L.G. Thompson, Proc. Soil Crop. Sci. Soc., 25, 335 7. (1965).
- 8. B.H. Katarki and A.L. Banahatti, Indian Oil Seed J., 9, 50 (1975).
- 9. F.C. Bosewell, P.E. Anderson and L.F. Welch, Res. Bull., 9 (1967).
- 10. G.U. Malewar and B.S. Indulkar, Indian J. Agron., 36, 5 (1991).
- 11. J. Nitsan and A. Long, Plant Physiol., 41, 956 (1966).
- 12. L.D. Nooden and K.V. Thimann, Proc. Natl. Acad Sci., 50, 194 (1963).

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