# **Preconcentration and Determination of Chromium Species Using Octadecyl Silica Membrane Disks and Ammonium Pyrrolidine Dithiocarbamate**

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> A novel and selective method for the fast determination of trace amounts of chromium species in water samples has been developed. The procedure is based on the selective formation of chromium ammonium pyrrolidine dithiocarbamate complexes at different pH in the presence of Mn(II) as an enhancement agent of chromium signals followed by elution with organic eluents and determination by atomic absorption spectrometry. The maximum capacity of the employed disks was found to be  $402 \pm 3$  µg and  $397 \pm 2 \,\mu$ g for Cr(III) and Cr(VI), respectively. The detection limit of the proposed method is 52 and 46 ng  $L^1$  for Cr(III) and Cr(VI), respectively. The proposed method was successfully applied for determination of chromium species Cr(III) and Cr(VI) in different water samples.

> **Key Words: Solid phase extraction, Ammonium pyrrolidine dithiocarbamate, Flame atomic absorption spectrometry, Octadecyl silica membrane disk.**

### **INTRODUCTION**

Toxicological studies have proved that the degree of toxicity of an element directly depends on the species in which it is present. Cr(III) is considered as an essential micronutrient for humans and mammals in order to maintain glucose metabolism, where as Cr(VI) is a potentially carcinogenic agent<sup>1</sup>. The significant drawbacks of  $Cr(VI)$  are breathing disturbances, liver and digestion malfunctions, dermal corrosion and skin allergies<sup>2</sup>. Therefore, It is necessary to control the level of chromium in industrial effluent, natural and drinking waters. Speciation of chromium in environmental samples is of prime importance.

There are numerous methods and techniques concerning chromium speciation and determination including liquid-liquid extraction after complex formation<sup>3,4</sup>, solid-liquid extraction<sup>5-8</sup>, LC-AAS<sup>9,10</sup>, FIA-AAS<sup>11,12</sup>, spectrophotometric<sup>12,13</sup>, ICP-AES<sup>14</sup> and NAA<sup>15</sup>. However, some of these techniques are time consuming and have elaborate sample preparation steps and low enrichment factors.

Solid phase extraction (SPE) methods are the best alternatives for traditional classic methods due to selective removal of trace amounts of metal ions from their matrices. SPE determinations can be carried out on different efficient ways. One of the most appropriate performation features of SPE is achieved by using octadecyl silica membrane disks. SPE reduce the use of toxic solvent, disposal costs and extraction time<sup>16,17</sup>. The octadecyl silica membrane disks involves shorter sample processing time and decreased plugging due to the large cross-sectional area of the disk and small pressure drop which allows higher flow-rates; reduced channeling resulting from the use of sorbent with smaller particle size and a greater mechanical stability of the sorbent bed $18$ .

In our previous attempts, we modified SPE membrane disks with suitable compounds for selective determination of chromium<sup>19-21</sup> and lead<sup>22</sup>. Meanwhile, other investigators have successfully utilized these sorbents for quantitative extraction and monitoring trace amounts of lead<sup>23-25</sup>, copper<sup>26-28</sup>, silver<sup>29,30</sup>, mercury<sup>31,32</sup>, cadmium<sup>33</sup>, palladium<sup>34</sup>, Ce<sup>35</sup> and UO<sub>2</sub><sup>36</sup>.

With chelating agent, such as ammonium pyrrolidine dithiocarbamate (APDC),  $Cr(III)$  and  $Cr(VI)$  can form different complexes<sup>37-39</sup> (Fig. 1a-b). As a result of the reaction of Cr(III) and APDC one product can be found *tris*[ammonium pyrrolidine dithioato-S,S']Cr(III). Cr(VI) is reduced by dithiocarbamates to Cr(III) which leads to two different Cr complexes<sup>38</sup>. In the case of APDC, these compounds are *bis*[pyrrolidine dithioato-S,S']- [pyrrolidineperoxydithioato, $O.S$ ]Cr(III)<sup>37,40</sup>.



Fig. 1a. Cr(III) and APDC *tris*[pyrrolidine-S,S']Cr(III); 1(b) Reaction product of Cr(VI) and APDC *bis*[pyrrolidine dithioato-S,S']-[pyrrolidineperoxydithioato-O,S]Cr(III)

The aim of this work was to develop a simple, highly sensitive and efficient method for the selective extraction and concentration of trace amounts of Cr(III) and Cr(VI) ions from aqueous media by means of pH adjustment using octadecyl silica membrane disks. The present article describes the preconcentration of APDC complexes of Cr(III) and Cr(VI) on octadecyl silica disk and their determination by off-line flame atomic absorption spectrometry (FAAS).

# **EXPERIMENTAL**

All reagents were of analytical grade (from Merck). APDC (0.01 % w/ v) was prepared in deionized water. Organic eluents used were of HPLC grade. Standard solutions of Cr(III) and Cr(VI) were prepared by appropriate dilution of a 1000  $\mu$ g mL<sup>-1</sup> stock solutions made from chromium(III) chloride and potassium dichromate, respectively.

Acidic buffer ( $pH = 4.5$ ) was prepared by mixing 100, 200 g CH3COONa·2H2O and 150, 30 mL glacial acetic acid in 1 L distilled water, respectively. The 0.45 mm nitro-cellulose membrane filters used for the wastewater samples filtration, were obtained from Schleicher and Schuell, Germany (REF-NO: 10404012) and 47 mm diameter solid phase extraction disks used for extraction and preconcentration of chromium from waste water samples, were supplied by Supelco ENVI-DISKTM (Cat.NO.57171).

A Varian SpectrAA model 200 atomic-absorption spectrometer was used for determinations. The operational characteristics of employed parameters are summarized in Table-1.

<b>DETERMINATION</b>			
Parameters	Chromium		
Hollow cathode lamp current (mA)	7.0		
Wavelength (nm)	357.9		
Slit width (nm)	0.2		
Background correction	Deuterium lamp		
Measurement mode	integrate		
Air flow rate $(L min^{-1})$	13.50		
Acetylene flow rate $(L \text{ min}^{-1})$	2.90		

TABLE-1 INSTRUMENTAL PARAMETERS FOR CHROMIUM

A Metrohm model 691 digital pH meter equipped with a combined glass-calomel electrode was used for pH adjustments and a Millipore filtration set has been employed for solid phase extraction.

**Proposed procedure:** Extraction was performed with ENVI-DISK™ (47 mm diameter and 0.5 mm thick) containing octadecyl-bonded silica (80 µm particles, 6 nm pore size). Typical capacity of a disk for satisfac-

tory retention of compounds ranges from 10 to 20 mg. The disks were used in conjunction with a standard Millipore 47 mm filtration apparatus connected to a desktop vacuum pump. In order to remove potential interferences and to ensure optimal extraction of the analyte of interest, the disk cleaning and conditioning should be performed before its use. Thus, after placing the disk in the filtration apparatus, 10 mL of methanol was poured onto the disk and immediately drawn through it by applying a slight vacuum. After all of the solvent was passed through the disk, it was dried by passing air through it for a few minutes. The disk conditioning started by pouring 10 mL of methanol onto the disk. Immediately a low vacuum was applied and the solvent was drawn through the disk until the solvent surface almost reaches the surface of the disk. The disk should not be allowed to be dried. This is to ensure complete wetting of the disk with the organic solvent. It is preferable to leave extra methanol above the disk rather than to allow air to contact the surface of the disk. Immediately thereafter 20 mL of water was introduced onto the disk and was drawn through it. The disk was then dried under vacuum for 5 min or longer if necessary. This is especially important for the disks, which are used for the first time. This step pre-wets the surface of disk prior to the extraction of Cr(III) and Cr(VI) ions from water. Then, a sample solution containing  $Cr(III)$  and  $Cr(VI)$ , species while its pH was adjusted to 1.5 or 6, respectively was passed through the disk The sample solution also contains 0.5 mg  $L^{-1}$  of Mn(II) and APDC (0.01 % w/v). The flow-rate was 60-80 mL min-1. After the extraction, the disk was dried completely by passing air through it. The extracted chromium species was desorbed from the disk by 5 mL of acidified methanol at a flow rate of 5-10 mL min-1. The eluting solution was collected in a 23 mm  $\times$  200 mm test tube which was placed under the stem of the extraction funnel. The chromium contents were then determined by AAS using an external calibration curve.

### **Analysis of water samples**

**Determination of Cr(VI):** Cr(VI) content of the samples was determined by adjusting the pH of the solution to 1.5 and maintaining the Mn(II) concentration at  $0.5$  mg  $L^{-1}$  and preconcentration and elution procedure was followed as described above.

**Determination of Cr(III):** Cr(III) content of the samples was determined after adjusting the pH to 6 and maintaining the Mn(II) concentration at  $0.5$  mg  $L^{-1}$  and preconcentration and elution was performed as described above.

**Analysis of ground water samples:** The ground water samples were first passed through a 45 µm (Millipore). Nylon filters to remove suspended particles. The Cr(VI) and Cr(III) contents of ground water samples were

analyzed by adjusting pH to 1.5 and 6 by addition of 0.5 mg  $L^{-1}$  of Mn(II), respectively and subjecting to preconcentration and elution as described above.

# **RESULTS AND DISCUSSION**

## **Selection of optimum conditions for extraction of Cr(VI)**

**Effect of pH:** It was observed that, pyrrolidine dithiocarbamate complex of Cr(VI) is efficiently retained on a membrane disk in the 1-2 pH range. However, according to Fig. 2, there is significant decrease in retention efficiency at  $pH = 3$  and no sorption of  $Cr(VI)$  was noticed in the 4-8 pH range, but was observed that, APDC complex of Cr(III) is efficiently retained on a membrane disk in the 4-8 pH range. Hence, the pH of the sample solutions was adjusted to 1.5 by addition of dilute HCl throughout the experiments.



Fig. 2. Effect of pH on the recovery percentage of 50 mg  $L^{-1}$ of each of Cr(VI) and Cr(III)

**Optimization of APDC concentration:** In order to investigate the influence of the APDC concentration on the quantitative extraction of chromium. The concentration of the ligand in 50 mL portions of sample solutions each containing 10  $\mu$ g of Cr(VI) was maintained (0.005 % w/v to 1 % w/v). In all cases, the extraction of Cr(VI) was found to be quantitative (Table-2). Hence, subsequent SPE experiments were carried out with 0.01 % w/v of the ligand.

TABLE-2 OPTIMIZED CONDITIONS FOR DETERMINATION OF Cr(VI) AND Cr(III)

Parameters	Chromium(VI)	Chromium(III)
pH	$1 - 2$	4-9
PDC concentration ( $w/v$ %)	$0.005 - 1$	$0.05-1$
Eluent flow-rate $(mL min^{-1})$	40	4.0

**Effect of different eluents:** In order to select the most effective eluent for the quantitative extraction of the sorbed chromium species, after he extraction of 25 µg of chromium from 50 mL of water, the retained chromium species were stripped with various amounts of different solvents. The results are tabulated in Table-3. It is noteworthy that in all cases the concentrations of Mn(II) and APDC were 0.5 mg  $L^{-1}$  and 0.01 % w/v, respectively. Present experiments revealed that 10 mL of methanol acidified with 0.01 M HCl is able to remove the retained chromium(VI) quantitatively.

### TABLE-3

	Recovery $(\%)$			
Stripping solution	2mL	$5 \text{ mL}$	$10 \text{ mL}$	
	$Cr(III)$ /	$Cr(III)$ /	$Cr(III)$ /	
	Cr(VI)	Cr(VI)	Cr(VI)	
Methanol	$37.2(3.5)^{b}$	71.5(2.5)	100.0(1.8)	
	5.7(2.9)	16.3(1.6)	99.6(2.8)	
Acidified methanol <sup>c</sup>	42.1(1.5)	68.2(1.4)	98.9(1.8)	
	60.2(3.5)	74.3(2.8)	99.5(2.7)	
Ammonical methanol <sup>d</sup>	23.2(2.5)	36.2(3.3)	82.4(2.2)	
	41.2(3.5)	50.7(1.7)	95.3(3.6)	
Ethanol	40.5(1.5)	52.6(1.4)	93.5(2.1)	
	47.6(1.0)	50.2(2.5)	95.2(2.6)	
1-Propanol	32.5(2.1)	41.9(2.5)	75.6(2.5)	
	28.5(3.0)	47.2(1.5)	82.3(1.5)	
Formic acid(1M)	22.0(2.5)	35.8(3.0)	79.2(3.5)	
	21.1(2.3)	31.1(1.7)	80.5(2.2)	
Hydrochloric acid (3 M)	31.4(2.1)	46.5(2.9)	68.5(3.5)	
	34.2(27.0)	55.9(3.0)	75.2(1.5)	
Hydrochloric acid (1 M)	20.5(3.2)	36.7(1.8)	53.7(2.1)	
	35.6(2.1)	42.5(2.1)	44.3(2.8)	
Nitric acid (3 M)	32.6(1.5)	39.2(1.2)	57.6(3.0)	
	40.1(2.9)	47.7(3.2)	79.2(0.5)	
Nitric acid (1 M)	30.2(2.5)	65.5(2.4)	67.1(3.0)	
	35.6(1.5)	52.2(1.5)	62.2(0.7)	
$\textsuperscript{a}$ Thitial samples contained 25 µg of each Cr(III) and Cr(VI) in 50 mL water.				

EFFECT OF DIFFERENT ELUTING SOLVENTS ON PERCENTAGE RECOVERY OF CHROMIUM ADSORBED ON THE DISK<sup>a</sup>

<sup>b</sup>Values in parentheses are RSD based on five individual replicate analysis.

c Acidified solvents obtained by addition of 0.01 M HCl.

<sup>d</sup> Ammoniacal solvents obtained by addition of 0.1 M NH<sub>3</sub>.

### **Selection of optimum conditions for extraction of Cr(III)**

**Effect of pH:** The influence of the pH of aqueous samples for the recovery of 10 µg of Cr(III) from 50 mL of solution containing was studied in the pH range of 2.0-8.0. The pH was adjusted using 0.1 M of either hydrochloric acid or sodium hydroxide solutions. The results shown in Fig. 2 indicate that Cr(III) ion can be retained quantitatively by the membrane disk at  $pH = 6$ . Higher  $pH$  values ( $> 9$ ) were not tested because of the possibility of the hydrolysis of octadecyl silica in the disks<sup>35</sup>.

**Effect of APDC concentration:** The APDC concentration was varied from 0.01-1 % w/v during the sorption of Cr(III)-PDC complex on membrane disk in the presence of manganese(II). Extraction of 20  $\mu$ g Cr(III) from 50 mL aqueous solutions under the optimal experimental conditions was conducted by varying the amount of ligand from 0.005 to 1.00 % w/v. In all cases, the extraction of Cr(III) was found to be quantitative. The results are accorded in Table-2. Therefore, concentration of APDC was maintained at 0.01 % w/v throughout the experiments.

**Effect of Mn(II):** Presence of 1-2 mg  $L^{-1}$  of Mn(II) in the sample solution results a ten-fold enhancement of the flame AAS signal due to increased retention efficiency of Cr(III). Addition of 1 mg  $L^{-1}$  of Zn(II), Fe(II) and Fe(III) instead of Mn(II) resulted in two-three and five-fold enhancement of flame AAS signal of Cr(III), respectively. However, in the presence of 5 mg L<sup>-1</sup> of Mn(II) in the sample, the addition of 1 mg L<sup>-1</sup> of Zn(II), Fe(II) and Fe(III) have no influence. Therefore, an overall concentration of 5 mg  $L^{-1}$  of Mn(II) in the sample solution was selected in subsequent experiments.

There is Cr(VI) in anionic form but Cr(III) is cationic therefore the addition 1-2 mg  $L^{-1}$  of Mn(II) to sample solution was found to provide maximum signal for Cr(III) during sorption on  $C_{18}$  bonded silica gel in presence of pyrrolidine dithiocarbamate as the chelating agent. It was also noted that the addition of 1-2 mg  $L^{-1}$  of Mn(II) results in ten-fold enhancement in the flame AAS signal due to increased retention efficiency of Cr(III).

**Effect of eluent acidity:** Cr(VI) and Cr(III) complexes are eluted quantitatively only with acidified methanol  $(> 0.1 \text{ M} \text{ in } HCl \text{ or } HNO<sub>3</sub>)$ . Hence, acidified methanol (0.1 M HCl in methanol) was used to strip the chromium species sorbed on membrane disk in the presence of Mn(II) pyrrolidine dithiocarbamate chelate (Table-3).

**Effect of flow rates:** The effect of flow rates of the sample solutions on the retention and recovery of Cr(III) and Cr(VI) were investigated. It was found that in the range of 8-80 mL min<sup>-1</sup>, the retention of  $Cr(III)$  and Cr(VI) by the membrane disk is hardly affected by the sample solution flow rate. Similar results for the extraction of inorganic and organic materials by octadecyl silica membranes disks have already been reported<sup>36</sup> (Fig. 3).



Fig. 3. Effect of flow rates of the sample solutions on the recovery percentage of 50 mg  $L^{-1}$  of each of Cr(VI) and Cr(III)

The effect of flow rates of the stripping sample solutions on the retention and recovery of Cr(III) and Cr(VI) were investigated. On the other hand, quantitative stripping of Cr(III) and Cr(VI) ions form the disk was achieved at a flow rate range of 1-20 mL min-1, using 5 mL of methanol (Fig. 4).



Fig. 4. Effect of flow rates of the striping solutions using 5 mL methanol on the recovery percentage of 50 mg  $L^{-1}$  of each of Cr(VI) and Cr(III)

**Interfering ions:** The interferences due to coexisting ions on the determination of 0.5  $\mu$ g mL<sup>-1</sup> of Cr(VI) or Cr(III) were investigated under the optimum experimental conditions. It was concluded that Na<sup>+</sup> (50000 mg L<sup>-1</sup>), K<sup>+</sup> (1000 mg L<sup>-1</sup>), Mg<sup>2+</sup> (2500 mg L<sup>-1</sup>), Ca<sup>2+</sup> (1000 mg L<sup>-1</sup>), Fe<sup>2+</sup> (1000  $\mu$ g L<sup>-1</sup>), Fe<sup>3+</sup> (5000  $\mu$ g L<sup>-1</sup>), Cd<sup>2+</sup> (1000  $\mu$ g L<sup>-1</sup>), Pb<sup>2+</sup> (5000  $\mu$ g L<sup>-1</sup>),  $Cu^{2+}$  (5000 µg L<sup>-1</sup>), Ni<sup>2+</sup> (1000 µg L<sup>-1</sup>), Mn<sup>2+</sup> (5000 µg L<sup>-1</sup>) and Zn<sup>2+</sup> (10000  $\mu$ g L<sup>-1</sup>) do not interfere in determination of chromium (Table-4). Additionally, the counter anions influence on the extraction and subsequent cleanup and recovery of chromium species has been summarized in Table-4.

### TABLE-4

# PER CENT RECOVERY OF Cr(III) AND Cr(VI) FROM BINARY MIXTURES<sup>a</sup> and PER CENT RECOVERY OF Cr(III) AND Cr(VI) FROM THE MEMBRANE DISK IN THE PRESENCE OF 0.02 M OF DIFFERENT COUNTER IONS<sup>b</sup>



<sup>a</sup>Initial samples contained 25 µg of each Cr(III) and Cr(VI) in 50 mL water. <sup>b</sup>Initial samples contained 25 µg of each Cr(III) and Cr(VI) and 0.02 M of each counter ion in 50 mL.

c Values in parentheses are RSD based on five individual replicate analysis.

**Effect of presence of Cr(VI) on determination of Cr(III) and** *vice versa*: The influence of varying amounts of Cr(III) in Cr(VI) determination and of Cr(VI) in Cr(III) determination have been investigated and the results are presented in Table-5.

 According to the results, 5000 and 1000 fold amounts of Cr(III) and Cr(VI) do not interfere in the determination of as low as 0.5  $\mu$ g L<sup>-1</sup> of Cr(VI) and Cr(III), respectively.

**Analytical performance:** Different volumes (10 mL–2.5 L) of sample solutions containing  $10 \mu$ g of each Cr(III) and Cr(VI) were passed through the disks. Under the best conditions, the chromium contents quantitatively



TABLE-5 ANALYSIS OF SYNTHETIC MIXTURES OF Cr(VI) AND Cr(III)

were retained in all cases. Hence, by considering the final elution volume of 10 mL and the break through volume of about 2.5 L, an enrichment factor of about 250 is easily achieved. The maximum capacity of the disks was determined by passing 50 mL portion of sample solution containing 800 µg of Cr(III) and 0.1 M acetate buffer at (pH = 6) and 0.01 % w/v APDC, also by passing 50 mL portion of sample solution containing 800 ug of Cr(VI) and HCl at  $pH = 1.5$  and 0.01 % w/v APDC, followed by the determination of the retained metal ions in the eluting solution using FAAS and an external calibration graph. The maximal capacity of the disk obtained from three individual replicate measurements was  $402 \pm 3 \mu$ g Cr(III) and  $397 \pm 2 \mu$ g Cr(VI) on the disk, respectively.

**Analysis of real samples:** To assess the applicability of the method to real samples, it was applied to the extraction and determination of Cr(III) and Cr(VI) from different water samples. Tap water (5 min after operation, Tehran (20 December 2004), ground water (Varamin-Charm Shahre, 12 January 2005), snow water (Tehran, 14 January 2005), rain water (Tehran, 14 December 2005).The samples were analyzed and the results are listed in Table-6. The amounts of Cr(III) were found to be in the range of 2.01- 70.91  $\mu$ g L<sup>-1</sup> and Cr(VI) 0.10-0.86  $\mu$ g L<sup>-1</sup>. Cr(VI) is detected in water samples which have been preserved for less than 4 min in 0.01 M HCl (Table-6).

Real samples marked 1 to 5, were collected at the distances of 5, 100, 250, 350 and 500 metres of the total waste water of electroplating plants in the eastern parts of Karaj, respectively. The total results obtained by the recommended procedure and ICP-AES have compared in Table-7. The results obtained by the proposed method and ICP-AES are in agreement with each other. Moreover, as expected, the chromium concentration decreases as the distance from the electroplating plants increases.

**Determination of methods detection limit:** The limit of detection (LOD) of metal ions by this mthod disk was studied under optimal experimental conditions, calculated<sup>41</sup> as CLOD =  $K_b$ .  $S_b/m$  where  $K_b$  is a numeri-

# TABLE-6

# RECOVERY OF CHROMIUM ADDED TO 1000 mL Of Different Water Samples (Containing 0.1 M ACETATE AT pH = 6.0) FOR Cr(III) AND  $HCl AT pH = 1.5 FOR Cr(VI)$







 $b$ <sup>b</sup>Mean  $\pm$  standard deviations based on five individual replicate analyses.

"Tabulated F-value for  $(4,4)$  degrees of freedom at p  $(0.95)$  is 6.39.

 $d$ Tabulated T-value for 8 degrees of freedom at p (0.95) is 2.306.

cal factor of 3,  $S_b$  is the standard deviation of ten replicates blank measurements and m is the slop of calibration curve. The LOD obtained for Cr(III) and  $Cr(VI)$  were 52 and 46 ng  $L^{-1}$ , respectively. It is also noteworthy that detection limit for each preconcentrated sample should be divided to its enrichment factor.

**Calibration curve:** The standard stock solutions of Cr(III) and Cr(VI) (1000 ppm) were prepared by dissolving 0.4867 g of CrCl<sub>3</sub>·6H<sub>2</sub>O (95 %) and 0.2825 g of  $K_2Cr_2O_7$  (99.9 %) in deionized water in 100 mL volumetric flasks, respectively. Standard solutions of chromium were obtained by appropriate stepwise dilution of standard stock solutions.



Fig. 5. Calibration curve of Cr(III) and Cr(VI) measured by SPE-AAS

The calibration curve of  $Cr(III)$  and  $Cr(VI)$  solutions is represented in Fig. 5 and the related regression is  $Y = 0.0177X + 0.0022$  and  $Y = 0.072X$  $+ 0.0008$ , providing a correlation coefficient of R = 0.9981 and R = 0.9997, respectively.

# **Conclusion**

 The proposed SPE method possesses advantages such as easiness and considerable selectivity in comparison with the previously reported procedures for isolation and determination of chromium contents (Table-8). The maximum time taken for separation, preconcentration and monitoring of chromium species in 50 mL portions of water samples is at the most 10 min. In the suggested procedure, any risk of analyte loss and or shift in equilibrium between the species has been minimized. The reproducibility of the procedure is near 3 %. The proper preconcentration factor improves the LOD of the method by a factor of about 250. This procedure has the advantage of preconcentration of Cr(III) or Cr(VI) depending on the pH of the sample solution.

# TABLE-8 COMPARISON OF PUBLISHED RESULTS OF SEVERAL ON-LINE OR SEVERAL METHODS FOR DETERMINATION OF Cr(III) AND Cr(VI)



### **REFERENCES**

- 1. J.O. Nriagu and E. Nicboer, Chromium in the Natural and Human Environment, New York (1988).
- 2. V. Camel, *Spectrochim. Acta*, **58B**, 1177 (2003).
- 3. E.B. Gonzalez, J.B. Garcia, P.B. Barrera and A.B. Barrera, *Fresenius' J. Anal. Chem.*, **344**, 301 (1992).
- 4. K.S. Subramanian, *Anal. Chem.*, **60**, 11 (1988).
- 5. R.M.C. Romero, M.C.Y. Biurrun and M.P.B. Barrera, *Anal. Chim. Acta*, **327**, 37 (1996).
- 6. M.C. Pannain and R.E. Santelli, *Talanta*, **42**, 1609 (1995).
- 7. B. Pasullean, C.M. Davidson and D.L. John, *J. Anal. Atom. Spectrom.*, **10**, 241 (1995).
- 8. M.S. Jimenez, I. Martin, J.M. Mir and J.R. Castillo, *Atom. Spectroscopy*, **17**, 201 (1996).
- 9. A. Gaspar, J. Posta and R. Toth, *J. Anal. Atom. Spectrom.*, **11**, 1067 (1996).
- 10. J. Posta, A. Gaspar, R. Toth and L. Ombodi, *Fresenius J. Anal. Chem.*, **355**, 719 (1996).
- 11. H.P. Zhou, S.K. Zu and Z.L. Fang, *Spectroscoy*, **17**, 112 (1996).
- 12. L. Girard and J. Hubert, *Talanta*, **43**, 1965 (1996).
- 13. H.D. Revanasiddappa and T.N.K. Kumar, *Talanta*, **60**, 1 (2003).
- 14. S. Hirata, Y. Umezaki and M. Ikeda, *Anal. Chem.*, **58**, 2602 (1986).
- 15. R.R. Greenberg, R. Ziesler, H.M. Kingston and T.M. Sullivan, *Fresenius J. Anal. Chem.*, **332**, 562 (1988).
- 16. R.E. Majors, *LC-GC*, **4**, 972 (1989).
- 17. D.F. Hangen, C.G. Markell, G.A. Scmitt and D.D. Blevins, *Anal. Chim. Acta*, **236**, 157 (1990).
- 18. C.F. Poole, S.K. Poole, D.S. Seibert and C.M. Chapman, *J. Chromatogr. B.*, **689**, 245 (1997).
- 19. M. Moghimi, Ph.D. Thesis, Islamic Azad University, Science and Research Campus, Unpublished Results, Tehran, Iran (2006).
- 20. M.S. Tehrani, A.A. Ebrahimi and F. Rastegrar, *Ann. Chim. (Rome)*, **94**, 429 (2004).
- 21. T.P. Rao, S. Karthikeyan, B. Vijayalakshmi and C.S.P. Iyer, *Anal. Chim. Acta*, **369**, 69 (1998).
- 22. M.M. Hosseini, K. Zare and M.S. Tehrani, *J. Phys. Theo. Chem. I.A.U. Iran*, **1**, 149 (2004).
- 23. O.R. Hashemi, F. Raoufi, M.R. Ganjali, A. Moghimi, M. Kargar-Razi, H. Aghabozorg and M. Shamsipur, *Anal. Sci.*, **16**, 1221 (2000).
- 24. M. Shamsipur, F. Raoufi and H. Sharghi, *Talanta*, **52**, 637 (2000).
- 25. O.R. Hashemi, M. Kargar-Razi, F. Raoufi, A. Moghimi, H. Aghabozorg and M.R. Ganjali, *Microchem. J.*, **69**, 1 (2001).
- 26. M. Shamsipur, A. Avanes, M.K. Raoufi, H. Sharghi and G. Aghapour, *Talanta*, **54**, 863 (2001).
- 27. M. Shamsipur, A.R. Ghiasvand, H. Sharghi and H. Naeimi, *Anal. Chim. Acta*, **408**, 271 (2000).
- 28. M.R. Ganjali, M.R. Pourjavid, L.H. Babaei and M.S. Niasari, *Quim. Nova*, **27**, 213 (2004).
- 29. M. Shamsipur, M. Javanbakht, Z. Ghasemi, M.R. Ganjali, V. Lippolis and A. Garau, *Sep. Purif. Technol.*, **28**, 141 (2002).
- 30. L.H. Babaei, Z. Ghasemi, F. Darciche, M. Shamsipur, F. Raoufi and M.R. Ganjali, *Anal. Sci.*, **17**, 1305 (2001).
- 31. M. Shamsipur, A. Shokrollahi, H. Sharghi and M.M. Eskandari, *J. Hazard. Mater.*, **117**, 129 (2005).
- 32. Z. Chen, G. Yang, Q. Hu, J. Yin and Q. Su, *J. Chin. Chem. Soc.*, **51**, 297 (2004).
- 33. H. Cesur and B. Bati, *Turk. J. Chem.*, **26**, 29 (2002).
- 34. K. Farhadi and G. Teimouri, *Talanta*, **65**, 925 (2005).
- 35. M. Shamsipur, M. Yousefi, Z. Ghasemi, L.H. Babaei and M.R. Ganjali, *Sep. Sci. Technol.*, **37**, 3525 (2002).
- 36. M. Shamsipur, A.R. Ghiasvand and Y. Yamini, *Anal. Chem.*, **71**, 4892 (1999).
- 37. C.M. Andrle, M. Linscheid and J.A.C. Broekaert, Unpublished Work (1995).
- 38. J.M. Hope, R.L. Martin, D. Taylor and A.H. White, *J. Chem. Soc., Chem Commun.*, 99 (1977).
- 39. C.M. Andrle, N. Jakubowski and J.A.C. Broekaert, *Spectrochim. Acta*, **52B**, 189 (1997).
- 40. Z.B. Alfassi and C.M. Wai, Preconcentration Techniques for Trace Elements, CRC Press, Boca, Raton Ann Arbor, p. 113 (1990).
- 41. D.J. Ingle, S.R. Crouch, Spectrochemical Analysis, Hall Engle Wood Cliff, NJ (1987).
- 42. V. Ghoulipour and S.W. Husain, *Acta Chromatogr.*, **12**, 170 (2002).

(*Received*: 11 November 2006; *Accepted*: 14 September 2007)AJC-5842