

Oxidation of the 5'-Position of Adenosine Analogues

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The yield for the oxidation of cordycepin was almost quantitative in reproducible runs. This method for adenosine and its analogues is far more attractive than methodologies using 1-(3-dimethylaminopropyl)-3-ethylcarbodiimide (EDC·HCl) and DMSO, dicyclohexyl carbodiimide (DCC) and dichloroacetic acid (DCAA) in DMSO, tetra-N-propylammonium perruthenate (TPAP) and N-methylmorpholine N-oxide (NMO), CrO₃ in pyridine, DMSO with trichloroacetic anhydride or oxidations which proceed through either the oxime or the 1,3-diphenylimidazolidine derivative to furnish the aldehyde.

Key Words: 5'-Hydroxyl oxidation, Periodinane, Purine nucleosides.

INTRODUCTION

Purine and pyrimidine nucleosides bearing functionalities at the 4' α -position have recently been shown to possess antiviral activity¹⁻⁴. Nucleosides such as 4'-cyano-, 4'-azido-, 4'-ethynylthymidine and 4'-ethynyl-2'-deoxycytidine are reported to exhibit potent anti-HIV activity. We have synthesized 4'-C-ethynyl-D-arabinofuranosyl and 4'-C-ethynyl-2'-deoxy-D-ribofuranosyl nucleosides which were shown to act as inhibitors of HIV reverse transcriptase through their triphosphates. Cordycepin (3'-deoxyadenosine) is reported to possess antiviral activity against several RNA viruses⁵. The molecular basis of this antiviral activity is believed to be the inhibition of viral RNA polymerases by cordycepin 5'-triphosphate.

In an effort to synthesize novel cordycepin analogues with functionalized substitution at the 4'-position, we have focused on the utilization of the carboxaldehyde (**1**) as the key intermediate for providing entry to this class of compounds. There are a few examples in the literature of nucleoside

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5'-carboxaldehydes, access to these key intermediates in sugar-modified nucleoside synthesis is exceedingly difficult. This is particularly the case for adenosine analogues. For example, Rosenberg and co-workers⁶ noted that oxidation of 2'-deoxy-3'-*o*-*tert*-butyldiphenylsilyl nucleosides and 2',3'-*o*-isopropylidene derivatives of nucleosides worked well under the Swern conditions [DMSO/COCl₂] in most cases except for adenosine derivatives. An alternate route to adenosine 5'-carboxaldehyde proceeds through the 1,3-diphenylimidazolidine or oxime derivatives⁷. However, this procedure is somewhat cumbersome and the yields are modest. Because adenosine 5'-carboxaldehyde derivatives represent attractive intermediates from which many novel adenosine targets can readily be synthesized, we developed a highly efficient procedure for this oxidation and our report communicates these results.

EXPERIMENTAL

¹H and ¹³C NMR spectra were recorded on Varian Mercury Plus 400 MHz or Varian Inova 500 MHz NMR spectrometers. High resolution ESI or FAB mass spectral data were obtained through the Nebraska Center for Mass Spectrometry. Column chromatographic separations were carried out using 230-400 mesh silica gel.

2',5'-Di-*o*-(*tert*-butyldimethylsilyl)-3'-deoxyadenosine (3): Adenosine (5.0 g, 18.73 mmol, 1.0 eq) was dissolved in dry pyridine (40 mL). *tert*-Butyldimethylsilyl chloride (TBDMSCl) (8.46 g, 56.19 mmol, 3.0 eq) was added in a single portion. The reaction stirred room temperature for 48 h and was then concentrated *in vacuo*. The residue was partitioned between 150 mL ice-cold 5 % HCl (aq) and 150 mL of CH₂Cl₂. The organic layer was washed with NaHCO₃ (aq) (75 mL) and brine (75 mL), dried over MgSO₄ and concentrated. The residue was purified by flash column chromatography with a gradient of hexane/EtOAc (4:1 to 2:1 to 3:2 to 1:1 to 1:4 to 100 % EtOAc). Two major fractions were collected which corresponded to the desired 2',5'-di-*o*-(*tert*-butyldimethylsilyl) (2',5'-di-OTBDMS) derivative and the 3',5'-di-OTBDMS isomer produced, respectively, in a ratio of 6:4. The 3',5'-di-OTBDMS derivative was isomerized by stirring with 100 mL of MeOH and 2.5 mL of NEt₃ (2.5 % v/v) at room temperature for 3 h. The reaction mixture was concentrated and purified by flash column chromatography. 2',5'-Di-*o*-(*tert*-butyldimethylsilyl)-3'-deoxyadenosine was obtained in 61 % (5.69 g) overall yield as a white solid. ¹H (CDCl₃, 500 MHz): δ 8.34 (s, 1H), 8.22 (s, 1H), 6.16 (bs, 2H, -NH₂), 6.12 (d, 1H, *J* = 5.0 Hz), 4.64 (t, 1H, *J* = 4.5 Hz), 4.29 (t, 1H, *J* = 4.0 Hz), 4.21 (q, 1H, *J* = 2.5, 5.5 Hz), 4.01 (dd, 1H, *J* = 2.0, 11.5 Hz), 3.85 (dd, 1H, *J* = 2.0, 11.5 Hz), 2.94 (bs, 1H, OH), 0.95 (s, 9H), 0.83 (s, 9H), 0.14 (d, 6H, *J* = 8.5 Hz), -0.09 (d, 6H, *J* = 28 Hz). ¹³C NMR (CDCl₃, 125 MHz): δ 155.7, 153.1,

150.1, 139.0, 119.9, 88.1, 85.4, 76.9, 71.4, 63.3, 26.2, 25.8, 18.7, 18.1, -4.8, -5.3. HRMS (M + H) calcd. for C₂₂H₄₂N₅O₄Si₂ 496.27, found 496.27.

The 2',5'-di-OTBDMS adenosine derivative prepared above (5.69 g, 11.49 mmol, 1.0 eq) was dissolved in dry CH₂Cl₂ (115 mL) and purged with N₂. Next, 4-dimethylaminopyridine (5.61 g, 45.98 mmol, 4.0 eq) was added in a single portion followed by phenoxythiocarbonyl chloride (3.2 mL, 22.99 mmol, 2.0 eq). The reaction mixture was stirred for 18 h at room temperature and was diluted with CH₂Cl₂ (200 mL). The organic phase was washed with water, 1 N HCl (aq), H₂O, NaHCO₃ (aq) and NaCl (aq) (200 mL each) and then dried over MgSO₄ and concentrated. The residue was purified by flash column chromatography using a solvent gradient of 4:1 to 2:1 to 1:1 to 3:7 hexane/EtOAc. The product was obtained as an orange foam in 60 % yield (4.33 g). ¹H (CDCl₃, 500 MHz): δ 8.52 (s, 1H), 8.37 (s, 1H), 7.58 (t, 2H, *J* = 8.0 Hz), 7.45 (t, 1H, *J* = 7.0 Hz), 7.25 (d, 2H, *J* = 8.5 Hz), 6.70 (s, 2H, -NH₂), 6.37 (d, 1H, *J* = 5.5 Hz), 6.01 (dd, 1H, *J* = 3.5, 5.0 Hz), 5.12 (t, 1H, *J* = 5.0 Hz), 4.66 (q, 1H, *J* = 2.5, 5.5 Hz), 4.22 (dd, 1H, *J* = 2.5, 11.5 Hz), 4.12 (dd, 1H, *J* = 2.5, 11.5 Hz), 1.12 (s, 9H), 0.98 (s, 9H), 0.32 (d, 6H, *J* = 5.0 Hz), 0.088 (d, 6H, *J* = 88.5 Hz). ¹³C NMR (CDCl₃, 125 MHz): δ 194., 155.9, 153.5, 153.2, 150.2, 138.6, 129.7, 126.9, 121.9, 119.9, 88.1, 82.8, 81.2, 74.9, 62.9, 26.3, 25.6, 18.6, 18.0, -5.0, -5.2. HRMS (M + H) calcd. for C₂₉H₄₆N₅O₅SSi₂ 632.27, found 632.27.

The 2',5'-di-OTBDMS-3'-phenoxythiocarbonyl adenosine derivative prepared above (4.33 g, 6.86 mmol, 1.0 eq) was dissolved in dry toluene (43 mL) in a 250 mL 3-neck round bottom flask and brought to reflux under N₂. A solution of toluene (21.2 mL), AIBN (225 mg, 1.37 mmol, 0.20 eq) and Bu₃SnH (2.8 mL, 10.29 mmol, 1.5 eq) was added *via* syringe over 0.5 h. The reaction mixture was heated under reflux for 6 h and then concentrated. The crude material was subjected to column chromatography using an EtOAc/hexane gradient of 6:4 to 7:3. The product was obtained as an off-white amorphous powder in 85 % yield (2.81 g). ¹H (CDCl₃, 500 MHz): δ 8.25 (s, 1H), 8.24 (s, 1H), 7.28 (s, 2H, -NH₂), 5.95 (s, 1H), 4.55-4.57 (m, 1H), 4.45-4.49 (m, 1H), 4.02 (dd, 1H, *J* = 2.0, 11.5 Hz), 3.67 (dd, 1H, *J* = 2.0, 11.5 Hz), 1.74-2.22 (m, 2H), 0.81 (s, 9H), 0.80 (s, 9H), 0.03 (d, 6H, *J* = 23.5 Hz), -0.001 (s, 6H). ¹³C NMR (CDCl₃, 125 MHz): δ 156.1, 152.6, 149.1, 138.4, 119.9, 91.7, 81.0, 76.0, 63.6, 33.6, 25.9, 25.6, 18.4, 18.0, -5.0, -5.6. HRMS (M + H) calcd. for C₂₂H₄₂N₅O₃Si₂ 480.28, found 480.28.

N⁶-Benzoyl-2'-*o*-(*tert*-butyldimethylsilyl)-3'-deoxyadenosine (4):

The 2',5'-di-OTBDMS-3'-deoxyadenosine **3** (2.54 g, 5.30 mmol, 1.0 eq) was dissolved in dry pyridine (72 mL) and stirred at room temperature under N₂. To this solution was added benzoyl chloride (2.5 mL, 21.21 mmol, 4.0 eq) and the reaction continued to stir for 18 h. The reaction was cooled

to 0 °C in an ice bath and NH₄OH (28 % aq) was slowly added (20 mL). The mixture stirred for 0.5 h at 0 °C. The solvent was removed *in vacuo* and the residue dissolved in EtOAc (350 mL). The organic layer was washed with water, NaHCO₃(aq) and brine (175 mL each) and dried over Na₂SO₄ and concentrated. The crude product was subjected to flash column chromatography using a 7 cm diameter column and 12 cm of silica gel. A solvent gradient of 7:3 hexane/EtOAc was used to elute the product. The N⁶-benzoylated product was isolated in 95 % (2.61 g) yield as a foam. ¹H (CDCl₃, 500 MHz): δ 9.33, (bs, 1H, -NH), 8.78 (s, 1H), 8.54 (s, 1H), 8.02 (d, 2H, *J* = 8.0 Hz, ArH), 7.54-7.60 (m, 1H, ArH), 7.48 (t, 2H, *J* = 8.0 Hz, ArH), 6.08 (d, 1H, *J* = 1.0 Hz), 4.62-4.66 (m, 1H), 4.56-4.61 (m, 1H), 4.13 (dd, 1H, *J* = 2.5, 12.0 Hz), 3.78 (dd, 1H, *J* = 2.5, 12.0 Hz), 2.10-2.28 (m, 1H), 1.88 (dd, 1H, *J* = 2.0, 6.0 Hz), 0.93 (s, 9H) 0.90 (s, 9H), 0.14 (d, 6H, *J* = 10.5 Hz), 0.10 (d, 6H, *J* = 10.5 Hz). ¹³C NMR (CDCl₃, 125 MHz): δ 165.0, 152.6, 151.3, 149.5, 141.6, 134.1, 132.7, 128.9, 128.0, 123.5, 92.2, 81.6, 77.5, 63.9, 33.9, 26.2, 25.8, 18.7, 18.1, -4.6, -4.9, -5.2, -5.3. HRMS (M + H) calcd. for C₂₉H₄₆N₅O₄Si₂ 584.30, found 584.30.

The N⁶-benzoyl-2',5'-di-OTBDMS-3'-deoxyadenosine prepared above (2.38 g, 4.59 mmol, 1.0 eq) was dissolved in THF (54 mL). The solution was cooled to 0 °C with a FLEX-COOL machine (EtOH bath). To this solution was carefully added a mixture of TFA (13.4 mL, 174.26 mmol, 38 eq) and water (13.4 mL). This reaction mixture was stirred at 0 °C for 24 h. The reaction mixture was neutralized with NaHCO₃ (aq) (200 mL) very carefully while stirring at 0 °C. The mixture was diluted with EtOAc (200 mL), washed with brine (100 mL) and dried over Na₂SO₄. The organic phase was concentrated *in vacuo* to give 1.84 g (96 % yield) of a foamy product. ¹H (CDCl₃, 500 MHz): δ 9.75 (bs, 1H, -NH), 8.89 (s, 1H), 8.41 (s, 1H), 8.18 (d, 2H, *J* = 8.52 Hz, ArH), 7.72 (t, 1H, *J* = 7.5 Hz, ArH), 7.60-7.75 (m, 2H, ArH), 5.95 (d, 1H, *J* = 4.0 Hz), 5.02-5.10 (m, 1H), 4.64-4.70 (m, 1H), 4.17 (d, 1H, *J* = 13.0 Hz), 3.74 (d, 1H), 2.56-2.66 (m, 1H), 2.18-2.28 (m, 1H), 0.96 (s, 9H), 0.09 (s, 3H), 0.002 (d, 3H, *J* = 2.0 Hz). ¹³C NMR (CDCl₃, 125 MHz): δ 165.1, 152.3, 150.8, 150.0, 142.5, 133.7, 132.8, 128.8, 128.1, 123.9, 93.2, 81.0, 75.2, 63.9, 34.4, 25.6, 17.9, -5.0, -5.1. HRMS (M + H) calcd. for C₂₃H₃₂N₅O₄Si 470.22, found 470.22.

5-(6-N-Benzoylamino-9H-purin-9-yl)-tetrahydro-4'-o-(tert-butyl-dimethylsilyl)furan-2-carb-aldehyde (1): The N⁶-benzoylated-2'-OTBDMS-3'-deoxyadenosine derivative prepared above (1.77 g, 3.77 mmol, 1.0 eq) was dissolved in dry CH₂Cl₂ (35 mL) and stirred at 0 °C in an ice bath. To this solution was added Dess-Martin periodinane (2.40 g, 5.66 mmol, 1.5 eq) in one portion and the reaction stirred at 0 °C for 1 h. The reaction was quenched at 0 °C by stirring with a solution of Na₂S₂O₃ (6.8 g in 40 mL water) and NaHCO₃ (saturated, aq, 40 mL) for 10 min to destroy

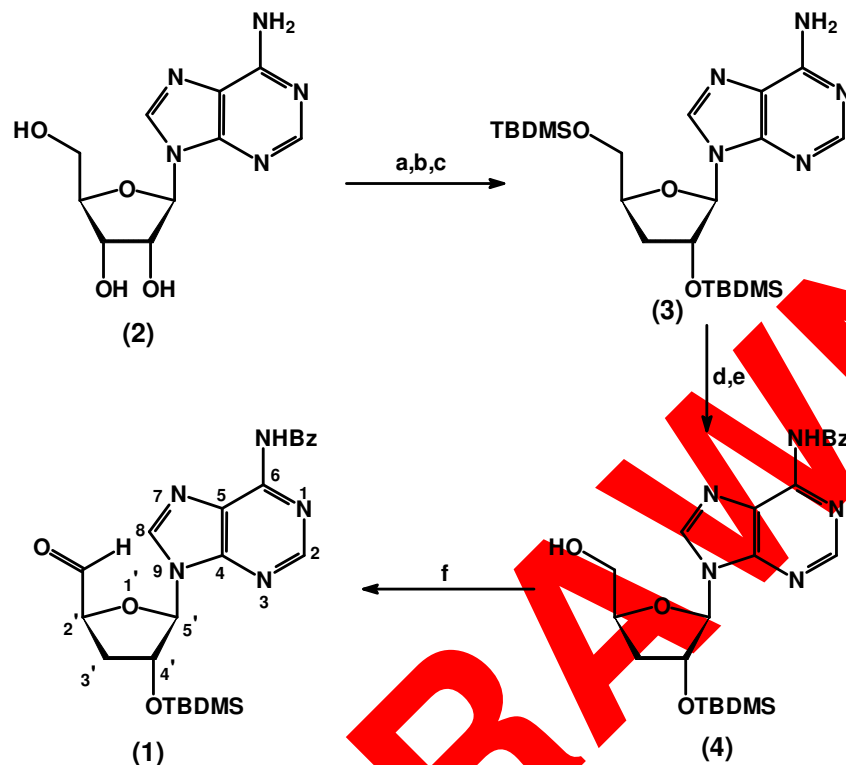
any unreacted Dess-Martin reagent. The reaction mixture was poured into a separatory funnel and extracted with EtOAc (3 × 80 mL). The organic layers were pooled and washed with brine (80 mL), dried over MgSO₄ and concentrated to give 1.73 g (98 %) of almost pure (> 98 %) product which was isolated as a foam. ¹H (CDCl₃, 500 MHz): δ 9.82 (s, 1H, -CHO), 9.03 (bs, 1H, -NH), 8.71 (s, 1H), 8.21 (s, 1H), 7.97 (d, 2H, *J* = 7.5 Hz, ArH), 7.40-7.60 (m, 3H, ArH), 5.98 (d, 1H, *J* = 2.0 Hz), 4.95-5.00 (m, 1H), 4.82 (t, 1H, *J* = 8.5 Hz), 2.23-2.42 (m, 2H), 0.83 (s, 9H), 0.03 (d, 3H, *J* = 4.5 Hz), 0.00 (s, 3H). ¹³C NMR (CDCl₃, 125 MHz) δ 199.9, 164.7, 152.4, 151.2, 149.8, 141.6, 133.5, 132.8, 128.8, 128.1, 124.0, 93.6, 84.2, 75.5, 35.2, 25.6, 17.9, -4.8, -5.0. HRMS (M + H) calcd. for C₂₃H₃₀N₅O₄Si 468.20, found 468.20.

RESULTS AND DISCUSSION

Our successful Dess-Martin oxidation of adenosine or its derivatives (*e.g.*, cordycepin) depends on a judicious selection of protecting groups for both the hydroxyl groups and the N⁶-position of the nucleobase. For example, for the synthesis of 3'-deoxyadenosine 5'-carboxaldehyde intermediate (**1**), we devised an efficient protection-deprotection strategy as shown in **Scheme-I**. Adenosine (**2**) was converted in three steps to compound **3**^{8,9} by selective 2',5'-disilylation (61 % yield), followed by treatment with phenyl chlorothionoformate and 4-dimethylaminopyridine (DMAP) in dichloromethane (60 % yield)¹⁰ and subsequent Barton radical deoxygenation reaction at C-3 (85 % yield)¹¹. Compound **3** can be deprotected to cordycepin and this approach represents an excellent method to produce gram quantities of this antiviral compound.

It was found that the order of the next two steps was very important in terms of yield. The N⁶-position of intermediate **3** was first protected using benzoyl chloride in pyridine (95 % yield)¹². Selective 5'-desilylation was subsequently accomplished using TFA-H₂O-THF (1:1:4) (96 % yield)¹³. Finally, Dess-Martin periodinane was used to oxidize the 5'-hydroxyl group to the aldehyde **1** (98 % yield)¹⁴. Compound **1** can be deprotected in almost quantitative yield. Although several other oxidation conditions were explored, including EDC·HCl and DMSO¹⁵, DCC and DCAA in DMSO¹⁶, TPAP and NMO¹⁷, CrO₃ in pyridine and DMSO with trichloroacetic anhydride¹⁸, only the Dess-Martin periodinane oxidation in CH₂Cl₂ proceeded with high efficiency.

In summary, N⁶-benzoyl-2'-O-(*tert*-butyldimethylsilyl)-3'-deoxy-4'-formyladenosine **1** can be synthesized in an efficient and facile manner using a judicious protecting group strategy and the Dess-Martin periodinane for the oxidation step. The more robust benzamide and *tert*-butyldimethyl silyl ether protections are far superior for this oxidation than the acetyl protecting group. The aldehyde produced is a key intermediate is useful in the synthesis of many 4'-substituted adenosine nucleosides of antiviral interest.



Scheme-I: Reagents and conditions: (a) TBDMSCl, pyridine, room temperature, 48 h; (b) PhOC(S)Cl, DMAP, CH₂Cl₂, room temperature, 6 h; (c) *n*-Bu₃SnH, AIBN, toluene, reflux, 4 h; (d) 1. BzCl, pyridine, room temperature, 2 h. 2. NH₂OH (28 %, aq.), 0 °C, 0.5 h; (e) TFA-H₂O (1:1), THF, 0 °C, 5 h; (f) Dess-Martin periodinane, CH₂Cl₂, 0 °C, 2 h.

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