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# Iridoid Compounds and Antimicrobial Activity of the Roots of *Tecoma stans* (L) Juss

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> The combined ethyl acetate and methanolic extracts of the roots of *Tecoma stans* gave two iridoid compounds, 5-deoxystantioside, plantarenaloside when subjected to chromatographic techniques. In addition to iridoid compounds luteolin, lupeol acetate and  $\beta$ -sitosterol were also isolated. The structures of these compounds were determined by spectral means (<sup>1</sup>H NMR, IR, UV, MS). Further the extracts and the isolates were subjected to antimicrobial studies, which showed moderate activity.

> Key Words: Iridoids, *Tecoma stans*, 5-Deoxystantioside, Plantarenaloside.

## **INTRODUCTION**

*Tecoma stans* (L) Juss (Bignoniaceae) is an ornamental tree, commonly known as yellow elder, distributed throughout the tropical parts of India. The roots of *T. stans* are used by the folk as a remedy for snake and rat bites, scorpion sting<sup>1,2</sup>. The plant was also reported to have antidiabetic and hypoglycemic activity<sup>3-7</sup>. Chemically the plant contains monoterpene alkaloids<sup>8,9</sup>, tecomine, tecostanine which were found to be responsible for the hypoglycemic activity<sup>10-14</sup>. In the present study two iridoid compounds, 5-deoxystantioside (TSR-IV), plantarenaloside (TSR-V) were isolated from the roots of *T. stans* apart from luteolin (TSR-III), lupeol acetate(TSR-II) and  $\beta$ -sitosterol(TSR-I).

### **EXPERIMENTAL**

UV Spectra were obtained on Systronics UV Spectrophotometer, IR spectra were recorded on BUCK Scientific-500 spectrophotometer using KBr pellets. Melting points were determined using Boeitus micro melting point apparatus and are uncorrected. The <sup>1</sup>H NMR spectra were taken on Bruker AM400 spectrophotometer with TMS as an internal standard. The mass spectra were taken on MAT-95 mass spectrophotometer. Column

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chromatography (CC) and TLC were carried out on silica gel (60-120 mesh, Acme) and Silica gel G (Acme), respectively. The visualization of TLC was done by spraying 5 % sulphuric acid reagent in methanol. All the solvents (Merck) used were distilled prior to use.

**Plant material:** The roots of *T. stans* were collected (2 Kg) from Andhra University campus, during December 2003 and authenticated by Dr. M. Venkaiah, taxonomist, Department of Botany, Andhra University, Visakhapatnam, a voucher specimen (SG/TS/12/2003) was deposited in the institution herbarium. The material was shade dried and coarsely powdered. About 1.5 Kg of the powder was extracted using soxhlet apparatus successively using petroleum ether (60-90 °C), ethyl acetate and methanol and concentrated under reduced pressure to get residues of 7.2, 12.3 and 15.4 g, respectively.

Isolation of compounds: The ethyl acetate and methanol residues of T. stans were combined as they were similar on TLC (chloroform:methanol, 9:1) and column chromatographed over silica gel (60-120 mesh, Acme) using gradient elution which yielded five compounds and were designated as TSR-I, TSR-II, TSR-III, TSR-IV and TSR-V. The compound TSR-I was obtained from 5 % ethyl acetate in petroleum ether which on crystallization gave colourless needles of m.p. 134-136 °C, showed positive Leibermann-Burchard test as green colour. Another compound TSR-II was obtained from 20 % ethyl acetate in petroleum ether which on crystallization gave colourless flakes of m.p. 214 °C and pink colour with Leibermann-Burchard test. The other compound TSR-III was obtained from 75 % ethyl acetate in petroleum ether which on crystallization gave yellow needles of m.p. 333-334 °C. On TLC a purple spot was observed under UV and intensified when exposed to ammonia, showed green colour with ferric chloride and orange colour with shinoda's test. The compound TSR-IV was obtained from 2 % methanol in ethyl acetate which on crystallization gave pale yellow flakes, m.p. 146-147 °C. It showed blue colour with Wieferring test. The compound TSR-V was obtained from 5 % methanol in ethyl acetate which on crystallization gave buff coloured amorphous powder. It showed blue colour with Wieferring test indicating iridoids.

### **RESULTS AND DISCUSSION**

Phytochemical examination of the roots of *T. stans* has been carried out. The combined root extacts of ethyl acetate and methanol on column chromatography afforded five compounds, TSR-I, TSR-II, TSR-III, TSR-IV and TSR-V. These compounds were identified as  $\beta$ -sitosterol, lupeol acetate, luteolin, 5-deoxystantioside and plantarenaloside by elemental analysis, spectal studies, m.m.p and co.TLC. The isolates 5-deoxystantioside and plantarenaloside were reported for the first time from the roots of *T. stans*.

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β-Sitosterol (TSR-I) colourless needles, m.p. 134-136 °C,  $[M]^+$  m/z: 414,  $[α]_D^{90}$  + 36° (chloroform), C<sub>29</sub>H<sub>50</sub>O, Found (%): C, 83.07; H, 12.7 Requires (%): C, 83.0; H, 12.2, IR (KBr, ν<sub>max</sub>, cm<sup>-1</sup>): 3440, 2970, 2880, 2050, 1470, 1385, 1055, <sup>1</sup>H NMR: δ 0.86, 1.25 (Me), 3.47 (1H, broad, 3-H) and 5.32 (1H, m, 5-H), the identity of TSR-I was confirmed by comparison with authentic sample through m.m.p and Co-TLC.

Lupeol acetate (TSR-II) colourless flakes, m.p. 214 °C,  $[M]^+$  m/z: 468,  $C_{32}H_{52}O_2$ , Found (%): C, 82.3; H, 10.8, Requires (%): C, 82.0; H, 11.2,  $[\alpha]_D{}^{30} + 37^\circ$  (C, 0.93, in chloroform), IR (KBr,  $v_{max}$ , cm<sup>-1</sup>): 2910, 2840, 1720, 1630, 1440, 1382, 1380, 1240, 1010, 970, 880, <sup>1</sup>H NMR (CDCl<sub>3</sub>, 90 MHz,  $\delta$ ):  $\delta$  0.76-1.02 (18H, 6XCH<sub>3</sub>), 1.65 (3H, s, CH<sub>3</sub>-C=CH<sub>2</sub>), 1.94 (3H, s, 3 $\beta$ OAc), 4.3 (1H, m, 3 $\alpha$ -H) 4.52 (2H, d, =CH<sub>2</sub>), the identity of compound **II** was confirmed by comparison with authentic sample through m.m.p and Co-TLC.

Luteolin (TSR-III) yellow needle, m.p. 333-334 °C,  $[M]^+$  m/z: 286,  $C_{15}H_{10}O_6$ ,  $\lambda_{max}$  MeoH n.m: 243 sh, 254, 266, 292 sh, 352, AlCl<sub>3</sub>: 264 sh, 328 sh, 402, AlCl<sub>3</sub>/HCl: 276 sh, 276, 296 sh, 356, 385, NaOAc: 269, 326 sh, 387, the identity of TSR-III was confirmed by comparison with authentic sample through m.m.p and Co-TLC.



TSR-I





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5-Deoxystantioside (TSR-IV) pale yellow flakes, m.p.146-147 °C,  $[M]^+$  m/z: 344, C<sub>16</sub>H<sub>24</sub>O<sub>8</sub>, Found (%): C, 55.4; H, 7.07, Requires (%): C, 55.8; H, 7.02  $[\alpha]_D^{30}$  -117°,  $\lambda_{max}$  MeOH n.m: 249, IR (KBr,  $\nu_{max}$ , cm<sup>-1</sup>): 3450, 2950, 2910, 2850, 1630, 1365, 1320, 1220, 1030, 900, 840, <sup>1</sup>H NMR (D<sub>2</sub>O, 90 MHz,  $\delta$ ):  $\delta$  9.18 (1H, s, H-11), 7.48 (1H, s, H-3), 5.54 (1H, d,  $J_{1.9}$  = 1.5 Hz, H-1), 4.88 (1H, d,  $J_{1.2}$  = 7.5 Hz, H-1), 3.00 (1H, m, H-5), 2.5-1.2 (6H, C-9, C-8, C-7.C-6), 1.15 (3H, d,  $J_{10.8}$  = 6.0 Hz MeOH). The data is well in accordance with the reported data.

Plantarenaloside (TSR-V) buff coloured amorphous powder,  $[M]^+ m/z$ : 360,  $C_{16}H_{24}O_9 \ [\alpha]_D^{25}$ -188.8°,  $\lambda_{max}$  MeOH nm: 241, IR (KBr,  $\nu_{max}$ , cm<sup>-1</sup>): 3450, 2950, 2910, 2850, 1630, 1365, 1320, 1220, 1030, 900, 840, <sup>1</sup>H NMR (D<sub>2</sub>O, 90 MHz,  $\delta$ ):  $\delta$  5.93 (H-1, s), 7.56 (H3, s), 0.90 (H10, d, 7.0), 9.23 (H-11, s), 4.83 (H-1<sup>1</sup>, d.7.5), D<sub>2</sub>O 97.3 (C-1), 165.5 (C-3), 125.0 (C-4), 72.9 (C-5), 38.3 (C-6), 32.2 (C-7), 34.2 (C-8), 51.7 (C-9), 15.9 (C-10), 194.6 (C-11), 99.4 (C-1<sup>1</sup>), 73.2 (C-2<sup>1</sup>), 76.1 (C-3<sup>1</sup>), 61.5 (C-6<sup>1</sup>). The data is well in accordance with the reported data.

	Zone of Inhibition											
Extracts	Gram +ve bacteria				Gram –ve bacteria				Fungi			
	B.s	B.p	S.a	S.p	E.c	Pr.v	P.a	Ps.v	A.n	C.a	S.c	R.o
Ethyl acetate extract (100 µg/mL)	13	10	17	12	14	13	12	11	09	10	12	09
Ethyl acetate extract (300 µg/mL)	17	15	20	16	19	18	15	20	14	12	15	11
Methanolic extract (100 µg/mL)	20	18	18	19	16	15	19	16	14	11	12	13
Methanolic extract (300 µg/mL)	25	23	21	24	21	22	24	22	17	15	15	16
Benzyl penicillin (10 µg/mL)	27	27	26	29	26	25	26	27	-	-	-	-
Ketaconazole (10 µg/mL)	-	-	-	-	-	-	-	-	23	20	20	21
TSR-IV (10 µg/mL)	19	17	16	18	15	14	17	16	14	12	15	13
TSR-V (10 µg/mL)	16	15	13	14	16	15	13	16	15	13	14	15
Ethyl acetate (Control)	-	-	-	-	-	-	-	-	-	-	-	-
Methanol (Control)	-	-	-	-	-	-	-	-	-	-	-	-

TABLE-1 ANTIMICROBIAL ACTIVITY OF THE TWO ISOLATES AND ETHYL ACETATE, METHANOLIC EXTRACTS OF THE ROOTS OF *T. stans* 

#Values are the average of triplicate, includes cup diameter (6 mm);

B.s = Bacillus subtilis; E.c = Escherichia coli; A.n = Aspergillus niger; B.p = Bacillus pumilis; P.a = Pseudomonas aeruginosa; C.a = Candida albicans; S.p = Streptococcus pyogens; Pr.v = Proteus vulgaris; S.c = Saccharomyces cerviceae; S.a = Staphylococcus aureus; Ps.v = Pseudomonas vulgaris; R.o = Rhizopus oryzae

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Antimicrobial activity of the two isolates and the root extract of T. stans: Antibacterial and antifungal activity were studied by agar cup plate method. Two concentrations (100 µg/mL, 300 µg/mL) of ethyl acetate, methanolic extracts of T. stans roots and 10 µg/mL concentration of TSR-IV and TSR-V were evaluated for their antibacterial and antifungal activity. Benzyl penicillin and ketaconazole were used as standards (10 µg/mL). Methanolic extract exhibited antibacterial activity against B. subtilis, S. pyogens, B. pumilis, Ps. vulgaris, Pr. vulgaris, P. aeruginosa and it was dose dependent. It also showed considerable activity on the tested fungi, S. cerviceae, R. oryzae. Whereas ethyl acetate extract exhibited moderate activity against S. aureus, E. coli, Pr. vulgaris and mild antifungal activity on the tested fungi. Among the compounds tested, the compound TSR-IV exhibited considerable activity against B. subtilis, S. pyogens, B. pumilis and Ps. vulgaris and showed moderate activity against other organisms. Whereas TSR-V exhibited moderate activity against B. subtilis, E. coli and Pr. vulgaris, mild activity against P. aeruginosa, S. pyogens and B. pumilis.

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