

NOTE**Chemical Study of Mucilage Obtained from Medicinal Plant *Cassia rengifera***

P.K. SINGH*, PRASHANT SINGH, PRIYANKA SINGH and KASIM HASAN ALI
*Chemical Laboratory of Carbohydrate Research, Department of Chemistry
Kamla Nehru Institute of Physical & Social Science, Sultanpur-228 001, India*

A water-soluble and non-ionic D-galactomannan has been isolated from the seeds of *C. rengifera* of Indian origin, containing D-galactose and D-mannose in 3:7 molar ratio. Acid catalyzed fragmentation, periodate oxidation, methylation and enzymic hydrolysis showed that the mucilage has a branched structure consisting of a linear chain of β -D-(1 \rightarrow 4) linked mannopyranosyl units, some of which are substituted at 1 \rightarrow 6 by α -D-galactopyranosyl units, glycosidically. This galactomannan have similarity with ghee guar plants.

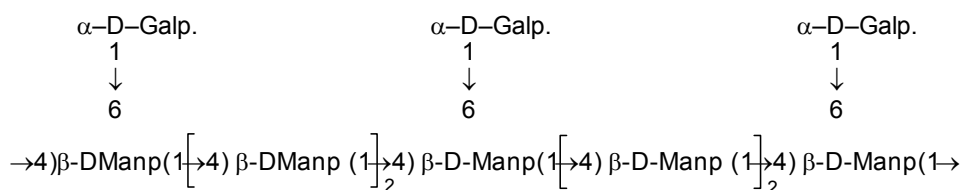
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A water soluble and non-ionic D-galactomannan has been isolated from the seed of *Cassia rengifera* of Indian origin¹. Polysaccharide was conveniently extracted from the crushed, defatted and decolorized seeds by extracting with 1 % aqueous acetic acid and by repeated precipitation² from its solution therein with ethanol. It was purified and tested for homogeneity by usual methods. The white amorphous polysaccharide had $[\alpha]_D^{25} + 68^\circ$ (in water), an ash content³ of (0.3 %) and a negligible percentage of methoxy, acetyl and uronic acid contents.

After complete acid-hydrolysis the polysaccharide yielded (D-galactose and D-mannose in 3:7 molar ratio). Graded acid hydrolysis resulted in the preferential removal of α -linked D-galactose units⁴ on the periphery as end groups. To determine the position of linkages between the building units of the galactomannan⁵, it was exhaustively methylated by Haworth-Purdie method^{6,7}, to afford a brown, semisolid glassy mass and had $[\alpha]_D^{25} + 41^\circ$ (chloroform). Hydrolysis of the methylated seed-gum gave 2,3,4,6-tetra-O-methyl-D-galactose⁸ (3 mol), 2,3,6-tri-O-methyl-D-mannose⁹ (4 mol) and 2,3-di-O-methyl-D mannose¹⁰ (3 mol).

The identity of these methylated monosaccharides¹¹ was established on the basis of their R_{TMG} values, optical rotations and crystalline derivatives. The percentage of end groups calculated from methylation studies was 29.9 %. Oxidation of the mucilage with sodium metaperiodate consumed 845 mM of the oxidant with the liberation of 183 mM of formic acid per 100 g of the polysaccharide indicating 29.5 % end-groups (*cf.* methylation).

Acid catalyzed partial hydrolysis of the mucilage gave, two disaccharides: α -D-Galp (1 \rightarrow 6)-D-Manp, β -D-Manp (1 \rightarrow 4)-D-Manp¹² and two trisaccharides α -D-Galp (1 \rightarrow 6)- β -D-Manp (1 \rightarrow 4)-D-Manp¹³ and β -D-Manp (1 \rightarrow 4)- β -D-Manp along with the component sugars. All of the oligosaccharides were characterized. These results corroborated the earlier findings. The foregoing data accord with the following structure.



Paper chromatography¹⁴ was conducted on Whatmann filter paper no. 1 and 3 mm papers by descending technique using the following systems (v/v). A-1-butanol-ethanol-water (5:1:4), B-1-butanol-isopropanol-water¹⁵ (11:6:3), C-ethylacetate-pyridine-water (2:1:2).

Solutions were concentrated at diminished pressure and at low temperature. All residues were dried *in vacuo* over anhydrous CaCl₂, melting points are uncorrected and $[\alpha]_D$ values are for equilibria.

REFERENCES

1. R.N. Chopra, In Glossary of Indian Medicinal Plants, CSIR, New Delhi, India p. 81 (1956).
2. A.M. Unrau and Y.M. Choy, *Can. J. Chem.*, **48**, 1128 (1970).
3. E.L. Hirst and J.K.N. Jones, *Discuss. Faraday Soc.*, **7**, 268 (1949).
4. S.A.J. Rizvi, P.C. Gupta and R.K. Kaul, *Planta Med.*, **19**, 222 (1971).
5. H. Meier, *Acta Chem. Scand.*, **14**, 749 (1960).
6. W.N. Haworth, *J. Chem. Soc.*, **107**, 8 (1915).
7. T. Purdie and J.C. Irvine, *J. Chem. Soc.*, **83**, 1021 (1903).
8. A.M. Staub, in ed.: R.L. Whistler, *Methods Carbohydr. Chem.*, Academic Press Inc., Vol. 5, p. 5 (1965).
9. H.C. Srivastava and P.P. Singh, *Carbohydr. Res.*, **4**, 326 (1967).
10. R.L. Whistler and G.E. Lauterbach, *Arch. Biochem. Biophys.*, **77**, 289 (1958).
11. H.O. Pouvang, H. Kiessling, B. Lindberg and J. McKay, *Acta Chem. Scand.*, **16**, 615 (1962).
12. A.B. Foster, in ed.: M.L. Wolfrom, in *Adv. Carbohydr. Chem.*, Academic Press, New York, p. 81 (1957).
13. A.S. Cerezo, *J. Org. Chem.*, **30**, 924 (1965).
14. S.N. Khanna and P.C. Gupta, *Phytochemistry*, **6**, 605 (1967).
15. F. Smith and R. Montgomery, in ed.: W.A.R. Hamor, *The Chemistry of Plant Gums and Mucilage*, New York, p. 134 (1959).

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