



Phenolic Compounds from the Fruits of *Ziziphus jujuba* Mill

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A new phenolic compound [2-hydroxy-1-(3'-hydroxy,4'-methoxy)-pentane-1,4-dione] (**1**), together with five known compounds (**2-4**) *i.e.*, 4-hydroxy-3-methoxy-phenyl-1-O-(6'-O-galloyl)- β -D-glucopyranoside (**2**), batatasin(III) (**3**), D-glucopyranoside (**4**), glucopyranosylcaffeic acid (**5**) and vanillic acid (**6**) were isolated from the fruits of *Ziziphus jujuba* Mill. The structure of compound **1** was elucidated on the basis of extensive NMR and MS means. The antioxidant activity of (**1**) was evaluated and it showed antioxidant activity with an IC₅₀ value of 2.92 μ g/mL.

Key Words: Phenolic compounds, Fruit, *Ziziphus jujuba* Mill, Antioxidant activity.

INTRODUCTION

Ziziphus jujuba Mill belongs to the genus *Ziziphus* (Rhamnaceae) and originated in China and call as Da Zao which is cultivated for more than 4000 years and where there are over 400 cultivars¹⁻³. The fruit of *Ziziphus jujuba* Mill is highly nutritional value and good tasting. In addition to fruit utilization, it has been used as a traditional Chinese medicine for a long time, which shows various beneficial activities including antioxidant, antihepatitis, anxiolytic, antitumor, *etc.*⁴⁻⁶. The extract of Da Zao has also been used as cigarette additives^{7,8}.

Previous phytochemical research on *Ziziphus jujuba* has revealed that flavones^{9,10}, triterpene acid^{11,12}, alkaloid^{13,14} as well as polysaccharides¹⁵ are major principles isolated from the plant of this plant. With the aim of continuing efforts to evaluate the quality of cigarette additives and to identify bioactive natural products from the plants, a chemical investigation on the fruits of *Ziziphus jujuba* Mill indigenous to the Hanzhong Prefecture of Shangxi Province of China was carried out. A new phenolic compound [2-hydroxy-1-(3'-hydroxy,4'-methoxy)-pentane-1,4-dione] (**1**), together with five known one *i.e.*, 4-hydroxy-3-methoxy-phenyl-1-O-(6'-O-galloyl)- β -D-glucopyranoside (**2**)¹⁶, batatasin(III) (**3**)¹⁷, D-glucopyranoside (**4**)¹⁸, glucopyranosylcaffeic acid (**5**)¹⁸ and vanillic acid (**6**)¹⁹ (Fig. 1) were separated from this plant. In addition, the antioxidant activity of (**1**) were evaluated. Described in this paper are their structure elucidation and biological activities.

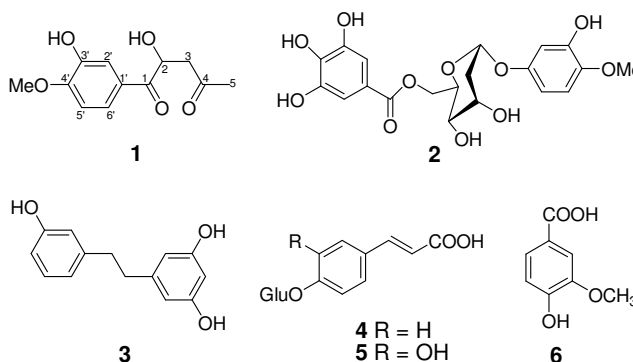


Fig. 1. Structure of phenolic compounds from the *Ziziphus jujube* Mill

EXPERIMENTAL

Optical rotation was measured in Horiba SEPA-300 high sensitive polarimeter. IR spectra were obtained in KBr disc on a Bio-Rad Wininfrared spectrophotometer. ESI-MS were measured on a VG Auto Spec-3000 MS spectrometer. ¹H, ¹³C and 2D NMR spectra were recorded on Bruker DRX-500 instruments with TMS as internal standard. Column chromatography was performed on silica gel (200-300 mesh) or on silica gel H (10-40 mm, Qingdao Marine Chemical Inc., China). On second separate used Agilent 1100 HPLC equipped with ZORBAX-C₁₈ (9.4 \times 250 nm, 5.0 mm) column and DAD detector.

The fruit of *Ziziphus jujuba* Mill was collected in Hanzhong Prefecture, Shangxi Province, P.R. China, in

February 2009 and identified by Prof. N. Yuan. A voucher specimen (No. YNNi 09-2-02) was deposited in our laboratory.

Extraction and isolation: The air-dried and powdered fruit of *Ziziphus jujuba* Mill. (1.5 kg) were extracted with 70 % aqueous ethanol (3.0 L × 3, 24 h each) at room temperature and the extract was concentrated under vacuum. The dried extract (118 g) was applied to silica-gel (200-300 mesh) column chromatography eluting with a CHCl₃-Me₂CO gradient system (9:1, 8:2, 7:3, 6:4, 5:5, 2:1) to give six fractions A-F. The separation of fraction E (CHCl₃-Me₂CO 1:1, 18.6 g) by silica-gel column chromatography eluted with CHCl₃-Me₂OH (9:1-1:2) yielded mixtures A1-A5. Fraction A2 (2.6 g) was subjected to preparative HPLC (40 % MeOH-H₂O, flow rate 12 mL/min) to give **1** (19.2 mg) and **3** (45 mg). Fraction A3 (1.8 g) was subjected to preparative HPLC (25 % MeOH-H₂O, flow rate 12 mL/min) to give **2** (35.6 mg), **6** (29.8 mg). Fraction A4 (1.2 g) was subjected to preparative HPLC (15 % MeOH-H₂O, flow rate 12 mL/min) to give **4** (18.6 mg), **5** (23.5 mg).

Antioxidant activity assay: Antioxidant activity was determined by the detection of the oxidative products with the 2',7'-dichlorofluorescein diacetate (DCFH) method reported previously²⁰.

RESULTS AND DISCUSSION

Compound **1** was obtained as a white amorphous solid with m.f. C₁₂H₁₄O₅ from HRESIMS (m/z: 239.0914 [M+H]⁺, calcd. 239.0919). IR absorptions at 3426, 1715 cm⁻¹ indicated the presence of hydroxyl and carbonyl groups, respectively. The ¹³C NMR spectrum of **1** (Table-1) along with analysis of the DEPT spectra displayed 12 carbon signals corresponding to 1-methyl (C, 31.2), 1-methylene (C, 48.4), 1-oxygenated methine (C, 71.2), 2-carbonyl carbons (C, 199.8, 207.6), 1-methoxyl carbon (C, 55.9) and 1-aromatic ring (C, 113.4, 115.9, 121.8, 125.2, 146.2, 152.8) with 3-aromatic protons (H, 6.84, 7.22, 7.46). The typical signals at (H, 6.84, d, *J* = 8.0), (H, 7.22, d, *J* = 8.0) and (H, 7.46, brs) observed in the ¹H NMR spectrum revealed the presence of 1, 2, 4-trisubstituted phenolic moiety, which was also supported by the HMBC correlations from H-2' (H, 7.46, brs) to C-3' (C, 146.2), C-4' (C, 152.8) and C-6' (C, 121.8), from H-5' (H, 6.84, d, *J* = 8.0) to C-1' (H, 125.2), C-3' (C, 146.2) and C-4' (C, 152.8) and from H-6' (7.22, d, *J* = 8.0) to C-2' (C, 115.9) and C-4' (C, 152.8). The side-chain of phenolic cycle was established by the HMBC correlations from H-2 (H, 5.82, dd, *J* = 4.6, 8.2) and H-3 (3.12, dd, *J* = 8.2, 15.8) to C-1 (C, 199.8), from H-2 (H, 5.82, dd, *J* = 4.6, 8.2), H-3 (H, 3.12, dd, *J* = 8.2, 15.8) and H-5 (H, 2.28 s) to C-4 (C, 207.6). The crosspeaks of H-2' (H, 7.46, brs) and H-6' (H, 7.22, d, *J* = 8.0) with C-1 (C, 199.8) (Fig. 2) helped in assigning the attachment of the side-chain to be at C-1'. The methoxyl groups located at C-4' was also supported by the HMBC of methoxyl proton (H, 3.79, s) to C-4'

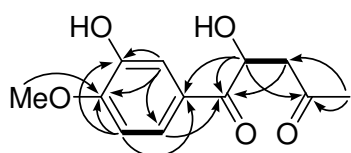


Fig. 2. Selected HBC (→) and 1H-1H COSY (---) of (**1**)

(C, 152.8). Thus, compound **1** was determined as 2-hydroxy-1-(3'-hydroxy, 4'-methoxy)-pentane-1, 4-dione.

The antioxidant activity of **1** was determined by the detection of the oxidative products with the 2',7'-dichlorofluorescein diacetate (DCFH) method reported previously²⁰. It shows antioxidant activity with an IC₅₀ value of 2.92 μg/mL. Compound **1** shows high antioxidant activity.

2-Hydroxy-1-(3'-hydroxy,4'-methoxy)-pentane-1,4-dione (**1**), C₁₂H₁₄O₅, white powder; [α]_D^{25.8} + 88.6 (c 0.028, CH₃OH); UV (CH₃OH), λ_{max} (log e) 206 (4.66), 248 (3.26), 288 (2.37), 326 (1.84) nm; IR (KBr, ν_{max}, cm⁻¹) 3426, 3027, 2926, 2855, 1715, 1655, 1527, 1485, 1382, 1026, 874, 762; ¹³C NMR and ¹H NMR data (CD₃OD, 500 MHz) see Table-1; positive ESIMS m/z 239 [M+H]⁺; HRESIMS m/z 239.0914 [M+H]⁺ (calcd. for C₁₂H₁₄O₅, 239.0919).

No.	δ _C (mult.)	δ _H (mult., <i>J</i> , Hz)
1	199.8 s	
2	71.2 d	5.82, dd, <i>J</i> = 4.6, 8.2
3	48.4 t	3.12, dd, <i>J</i> = 8.2, 15.8 3.33, dd, <i>J</i> = 5.2, 15.6
4	207.6 s	
5	31.2 s	2.28 s
1'	125.2 s	
2'	115.9 d	7.46, brs
3'	146.2 s	
4'	152.8 s	
5'	113.4 d	6.84, d, <i>J</i> = 8.0
6'	121.8 d	7.22, d, <i>J</i> = 8.0
OMe	55.9 q	3.79, s

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REFERENCES

- H.B. Yang, S.Q. An, O.J. Sun and Z.M. Shi, *J. Integr. Plant. Biol.*, **50**, 210 (2008).
- H. Zheng, H.F. Lu, Y.P. Zheng, H.Q. Lou and C.Q. Chen, *Int. J. Food. Eng.*, **101**, 402 (2010).
- P. Tetali, C. Waghchaure, P.G. Daswani, N.H. Antia and T.J. Birdi, *J. Ethnopharmacol.*, **123**, 229 (2009).
- H. Zhang, L. Jiang, S. Ye, Y.B. Ye and F.Z. Ren, *Food. Chem. Toxicol.*, **48**, 1461 (2010).
- M.W. Saif, F. Lansigan, S. Ruta, L. Lamb, M. Mezes and K. Elligers, *Phytomedicine*, **17**, 161 (2010).
- C.A. Wu, J.J. Wu, M.J. Tsai and R.Y. Chen, *J. Ethnopharmacol.*, **113**, 300 (2007).
- Z.Y. Zhu, X.C. Liao, G.Z. Li and P. Ji, *Chin. J. Agric. Sci. Bull.*, **12**, 1235 (2007).
- J.S. Zhang, X.C. Jia, Y. Dai, D. Bi Mai and W.Y. Zhang, *Chin. J. Tob. Sci. Tech.*, **26**, 476 (2003).
- Y. Tamaki, T. Konishi, M. Fukuta and M. Tako, *Food. Chem.*, **107**, 352 (2008).
- J.G. Jiang, X.J. Huang, J. Chen and Q.S. Lin, *Nat. Prod. Res.*, **21**, 310 (2007).

11. S. Guo, Y.P. Tang, J.A. Duan, S.L. Su and A.W. Ding, *Chin. Chem. Lett.*, **20**, 197 (2009).
12. X.H. Fang, J.F. Hao, H.Y. Zhou, L.X. Zhu and J.H. Wang, *Phytomedicine*, **17**, 75 (2010).
13. Y. Ma, H.S. Han, S.Y. Nam, Y.B. Kim and J.T. Hong, *J. Ethnopharmacol.*, **117**, 318 (2008).
14. H.S. Han, Y. Ma, J.S. Eun, R.H. Li, J.T. Hong and M.K. Lee, *Pharm. Biochem. Behav.*, **92**, 206 (2009).
15. Z.H. Zhao, J. Li, X.M. Wu, H. Dai, X.M. Gao and P.F. Tu, *Food. Res. Int.*, **39**, 917 (2006).
16. K. Ishi Maru, G.I. Nonaka and I. Nishioka, *Phytochemistry*, **26**, 1147 (1987).
17. X.Y. Guo, Y. Wang and N.L. Wang, *Chin. Trad. Herbal. Drugs*, **37**, 492 (2006).
18. C.B. Cui, Y. Tezuka and T. Kikuchi, *Chem. Pharm. Bull.*, **40**, 2035 (1992).
19. X.H. Pan, S. Li and Y. Li, *Nat. Prod. Res. Develop.*, **17**, 290 (2005).
20. S. Takamatsu, A.M. Galal, S.A. Ross, D. Ferreira, M.A. Elsohly, A.R. Ibrahim and F.S. El-Ferally, *Phytother. Res.*, **17**, 963 (2003).