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# Growth and Chemical Composition of Cherry Rootstock Gisela 5 Cultured Under Different Bicarbonate Concentrations *in vitro*

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In vitro response of Gisela 5 cherry rootstock to increasing concentrations of HCO<sub>3</sub><sup>-</sup> (0, 5, 10, 20, 30, 40, 50 mM) in MS medium was investigated. With increasing concentration of HCO<sub>3</sub><sup>-</sup>, dry biomass and shoot length were negatively affected. At 50 mM HCO<sub>3</sub><sup>-</sup> treatment, the reduction of dry biomass production was 48 % whereas shoots length exhibited 4 %. Leaf injury symptoms were more severe at HCO<sub>3</sub><sup>-</sup> levels  $\geq$  20 mM. Viability loss was monitored in explants by increasing HCO<sub>3</sub><sup>-</sup> levels. Chlorophyll content (SPAD units) of leaves declined with increasing HCO<sub>3</sub><sup>-</sup> concentrations of the culture medium. As for ion concentration, increasing HCO<sub>3</sub><sup>-</sup> levels of the culture medium caused K and Ca concentrations increased. When HCO<sub>3</sub><sup>-</sup> level increased, the content of Fe did not directly affected. Cu and Zn concentrations in the explants were not significantly changed by HCO<sub>3</sub><sup>-</sup> levels.

Key Words: Bicarbonate, Cherry, Tissue culture.

# **INTRODUCTION**

High concentrations of bicarbonate ion in soil solution and around plant roots caused by excessive amounts of calcareous materials is one of the main nutritional problems of fruit orchards. Heavy soil texture, soil compaction, high moisture content, deficient drainage, a high level of microbial activity, as well as a high rate of root respiration would increase the partial pressure of  $CO_2$  in soil which, in turn, would mean high concentrations of  $HCO_3^-$  in soil solution<sup>1</sup>. On the other hand, high concentrations of bicarbonate ion in irrigation water are also considered as one of the common factors in increasing the bicarbonate content of soil solutions<sup>2</sup>.

The physiological problems caused by bicarbonate ion are quite complicated<sup>1,3,4</sup>. Researchers have presented various mechanisms to explain such problems. One of the obvious effects of excessively high bicarbonate ion concentrations in the root zone is the problem in translocation of Vol. 19, No. 4 (2007)

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calcium ions from plant roots to shoots and its accumulation in the roots with precipitation as CaCO<sub>3</sub>. Production and accumulation of organic acids within the roots would be another adverse physiological effect of too much bicarbonate ion in the root zone<sup>1</sup>. Similarly, high concentrations of bicarbonate in the rhizosphere are associated with high pH levels in cytoplasm, high rates of root respiration, cytochrome oxidase enzyme activity and reduction in the chlorophyll content of the leaves<sup>1</sup>. Various researchers<sup>5-7</sup> working with differing concentrations of bicarbonate ion in solution culture found a positive and significant correlation between the extent of leaf chlorosis and bicarbonate ion concentrations. On the other hand, the findings<sup>2</sup> make it imperative to investigate the concentrations of bicarbonate ion in irrigation water that can cause serious nutritional problems in agricultural crops. Shahabi et al.1 investigated the effects of varying concentrations of bicarbonate in the irrigation water on nutritional disorders of some apple cultivars, grown under greenhouse conditions. They concluded that the increasing levels of bicarbonate ion increased the leaf concentrations of N, P and K, but lowered the concentrations of Fe, Mg and Mn and did not affect Ca, Cu, or Zn levels. Brancadore et al.8 investigated the resistance of various peach and grape varieties to bicarbonate ion in solution culture. They concluded that bicarbonate ion caused Fe chlorosis and thus that resistance to Fe chlorosis depends on the variety and also the genetic factors of the plants. In the case of the Mazani variety grown under conditions of high bicarbonate ion concentrations in the rhizosphere, the roots had become more resistant to bicarbonate's adverse effects than the roots developed in a rhizosphere containing Fe but no bicarbonate ion.

Gisela 5 cherry rootstock is among the best dwarfing, precocious and productive rootstocks for modern, intensive sweet cherry growing. Recently it has replaced *Prunus avium* and *Prunus mahaleb* rootstock in cherry growing area<sup>9,10</sup>. Despite its thorough testing in many different scientific investigations, the studies on the responses of Gisela 5 to environmental stresses are limited and the responses of bicarbonate ion remain unclear.

Different techniques and approaches have been developed, alone or in sequence, from the most conventional, as pot or hydroponic culture, to more recent approaches, as *in vitro* culture and molecular investigations<sup>11</sup>. Tissue culture can be an attractive system to unravel the physiological and biochemical aspects undergoing the different genotypic responses *vs.* bicarbonate enriched conditions<sup>11-13</sup>. The objective of this study was to determine the effect of HCO<sub>3</sub><sup>-</sup> on dry weight, shoot length, chlorophyll content and chemical composition of *in vitro* produced shoots of Gisela 5 rootstock. The understanding of the physiological and biochemical responses to bicarbonate ion stress in this rootstock can be important for the cherry growing area affected bicarbonate ion.

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# **EXPERIMENTAL**

**Plant material and growth conditions:** Shoot-tips from healthy shoots of 3-year- old plants of Gisela 5 were established on MS (M 5519, Sigma Chemical Co., St. Louis, Missouri, USA) medium supplemented with sucrose (30 g L<sup>-1</sup>) and solidified with agar (7g L<sup>-1</sup>) and media pH adjusted to 5.8 before autoclaving (for 20 min at 121°C). The explants were established and proliferated on the above medium containing 1.0 mg L<sup>-1</sup> 6-benzylaminopurine (BAP), 0.1 mg L<sup>-1</sup> indole-3 butyric acid (IBA) and 0.1 mg L<sup>-1</sup> gibberellic acid (GA<sub>3</sub>). The proliferated cultures were subcultured at 4 week intervals and all cultures were maintained at  $25 \pm 1^{\circ}$ C under a 16 h photoperiod with white light (51 µmol m<sup>-2</sup> s<sup>-1</sup>).

**Bicarbonate treatments:** For  $HCO_3^-$  treatment (supplied as NaHCO<sub>3</sub>), three-node-shoot explants were exposed to increasing concentrations of NaHCO<sub>3</sub> (0, 5, 10, 20, 30, 40 and 50 mM) on MS solid medium. According to NaHCO<sub>3</sub> concentrations, pH of media was 5.80, 6.95, 7.20, 7.47, 7.70, 7.72, 7.74, respectively. After 4 weeks, the explants were collected and washed for 2 min with distilled water to remove medium, dried on filter paper and either used or dried at 70°C for later use.

**Measurements:** The growth response of Gisela 5 to  $HCO_3^-$  was measured in terms of both dry weight and shoot length. Dry matter of explants was obtained by heated at 70°C for 24 h. Data are expressed as per cent of control at each level of  $HCO_3^-$ .

The relative chlorophyll content was measured with a portable leaf chlorophyll meter (SPAD 502, Minolta Co. Ltd., Japan) and the data represented were the means of one reading from each explant of each replicate.

Explants were also scored for visible symptoms of bicarbonate injury on a scale, ranging from 1 to 5 (1, no symptoms; 2, light green colour; 3, green shoot-tip and browning of leaf edge; 4, intensive brown leaf; 5, dead). Following this, bicarbonate injury index (BI Index) was calculated according to the following formula:

BI Index =  $\Sigma$  (n<sub>i</sub> × i)/N, where n<sub>i</sub> is the number of explants receiving the mark i (from 1 to 5) and N is the total number of explants in each bicarbonate concentration.

To determine the mineral composition, 0.3-0.4 g of previously dried, ground and homogenized plant material was placed in a platinum crucible. Crucible was covered during the ashing as a precaution against contamination. Dry ashing was accomplished in a muffle furnace at 550°C for 6 h. After a 10 min cooling period, the ash was dissolved in 2 mL concentrated HNO<sub>3</sub> (65 %) and heated on a hot plate for 10 min. The digested material was transferred into a volumetric flask and made up to 25 mL with distilled water, which was used for rinsing the platinum crucible. The digested material was diluted to 25 mL with the rinsing solutions prior to analyses.

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The concentration of Na, Mg, K, Mn, Ca, Cu, Zn and Fe in the explants were determined by atomic absorption spectrophotometer (Unicam, 929 Model AA).

**Statistics:** The experiments were set up in a completely randomized design. There were four explants in each 250 mL jar (containing 40 mL medium), 3 jars in each replicate and 3 replicates in each treatment. Analysis of variance was performed on the data and significant differences among treatment means were calculated by LSD test at p < 0.05.

# **RESULTS AND DISCUSSION**

Table-1 shows both growth parameters (dry biomass and shoot length) were differently affected by the increasing bicarbonate levels. Dry biomass was markedly reduced in the 50 mM HCO<sub>3</sub><sup>-</sup> treatment (shoot dry weight of bicarbonate treated explants was about 48 % of that control plants), while shoot length was slightly decreased. Increases in the concentration of HCO<sub>3</sub><sup>-</sup> in the nutrient medium decreased dry weight of shoots in peach<sup>14,15</sup>, barley, sorghum and maize<sup>6</sup>, tobacco<sup>16</sup>, tomato<sup>17</sup> and rice<sup>18</sup>. Whereas, shoot growth was not affected by bicarbonate in lupins<sup>3</sup>. On the other hand, Zribi and Gharsalli<sup>19</sup> reported that the growth response of pea plants to the bicarbonate treatments differed according to the cultivar. Similar results were also reported for grapevine cultivars<sup>20,21</sup>.

CHLOROPHYLL CONTENTS AND INJURY OF GISELA 5 GROWN in vitro												
NaHCO <sub>3</sub> (mM)	pH of Media	Dry Biomass Production (% of control)	Shoot length (% of control)	Relative chlorophyll content (SPAD)	Leaf Injury Index							
0	5.83	100.00 a*	100.00 a	41.74 a	1.00 d							
5	6.95	105.00 a	96.87 b	39.30 a	1.27 d							
10	7.20	77.66 b	96.76 b	18.49 b	3.05 c							
20	7.47	61.53 cd	96.25 b	10.29 bc	4.19 b							
30	7.70	60.20 cd	95.62 b	10.16 bc	4.27 b							
40	7.72	65.69 c	95.41 b	4.46 c	4.38 ab							
50	7.74	52.22 d	95.40 b	0.49 c	4.88 a							

TABLE-1

EFFECT OF HCO<sub>3</sub><sup>-</sup> CONCENTRATIONS ON THE GROWTH,

\*Values not associated with the same are significantly different (p < 0.05).

Chlorophyll contents of leaves declined as HCO<sub>3</sub><sup>-</sup> concentration of the culture medium increased from 0 to 50 mM (Table-1). Marginal chlorosis was noted on the leaves when HCO<sub>3</sub><sup>-</sup> concentration was above 10 mM, which later resulted in brown and dead leaves. Shahabi *et al.*<sup>1</sup> reported that different bicarbonate levels in the irrigation water significantly reduced

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chlorophyll development in seedlings of apple cultivars. Ksouri *et al.*<sup>21</sup> found that chlorophyll concentration of leaves of grapevine cultivated on bicarbonate-enriched medium decreased. Similar results were also found in peach rootstocks<sup>11,14</sup>.

Leaf injury developed only when  $HCO_3^-$  levels reached 10mM. As the  $HCO_3^-$  level increased from 10 to 50 mM, the severity of the injury increased. Table-1 indicates that 10 mM  $HCO_3^-$  treatment induced green-shoot tip and browning of leaf edge, whereas above 10 mM  $HCO_3^-$  stimulated intensive brown leaves. The burned appearance suggested that, in this case, the leaf damage may have been caused with  $HCO_3^-$  toxicity, as observed in tobacco plants<sup>16</sup>. Coulombe<sup>22</sup> and Mengel and Geurtzen<sup>23</sup> reported that leaf injury in the NaHCO<sub>3</sub> experiment may be attributed to toxicity of  $HCO_3^-$ .

Table-2 shows the ion concentration of the explants treated with increasing bicarbonate concentrations in the culture medium. K concentration declined markedly as  $HCO_3^-$  concentration increased. Similar findings were observed in peach rootstock<sup>14</sup>. K is an essential cation for plants and is required for the regulation of ion transport and osmotic regulation<sup>24</sup>. Ca concentration increased initially as  $HCO_3^-$  levels increased, but decreased again at the two highest levels of  $HCO_3^-$ . Decreasing Ca concentration with increasing  $HCO_3^-$  level was reported in peach, lettuce and celery<sup>14,24,25</sup>. As mentioned, bicarbonate effects may have appeared as decreases in the rates of absorption, as translocation of nutrients, or as reduced activities of those elements.

TABLE-2 EFFECTS OF HCO<sub>3</sub><sup>-</sup> ON ION CONCENTRATIONS (mg/g) OF GISELA 5 GROWN *in vitro* 

NaHCO <sub>3</sub> (mM)	K	Ca	Na	Mg	Mn	Fe	Cu	Zn			
0	31.75 a*	5.91 d	1.20 e	1.38 c	0.121 b	0.231 b	0.008	0.095			
5	27.94 a	6.01 d	1.33 de	1.23 c	0.137 ab	0.231 b	0.006	0.074			
10	18.79 b	6.66 cd	1.99 d	1.49 c	0.129 b	0.339 a	0.008	0.120			
20	17.10 bc	8.47 ab	3.02 c	1.87 b	0.157 a	0.198 b	0.007	0.127			
30	15.44 bc	9.72 a	4.32 b	2.29 a	0.155 a	0.217 b	0.005	0.096			
40	16.50 bc	7.80 bc	4.70 b	2.09 ab	0.148 ab	0.217 b	0.004	0.079			
50	14.10 c	6.98 cd	5.65 a	1.95 ab	0.159 a	0.339 a	0.005	0.103			
<b>VV</b> 1		• • • •	1 .1		· C'	1.00		171			

\*Values not associated with the same are significantly different (p < 0.05).

Increased HCO<sub>3</sub><sup>-</sup> caused significant increases in the Na, Mg, Mn content of the explants (Table-2). Increase in Na contents of the explants may be attributed to the increases in NaHCO<sub>3</sub> concentration of the medium. Similar results were found by Bie *et al.*<sup>24</sup>. Shi *et al.*<sup>14</sup> and Pearce *et al.*<sup>26</sup> demonstrated that excess HCO<sub>3</sub><sup>-</sup> did not change Mg content in peach

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and tobacco leaves. Cinelli and Viti<sup>27</sup> found that so high bicarbonate conditions did not affect foliar Mn content in the Myrobalan clones and GF 677 rootstock. But, Shahabi *et al.*<sup>1</sup> and Shi *et al.*<sup>14</sup> reported that increased HCO<sub>3</sub><sup>-</sup> concentrations resulted in lower concentrations of Mn in leaves. The results of the present study do not agree with the above research findings. Higher Mn concentrations are usually found in chlorotic plants grown in calcareous soils<sup>28,29</sup> and Mn has been reported to inactivate Fe metabolic activity by decreasing Fe<sup>2+</sup> concentrations and resulting in lower chlorophyll concentrations<sup>30</sup>.

Fe concentration of the explants did not decrease linearly as the HCO<sub>3</sub><sup>-</sup> level increased (Table-2). The higher Fe concentrations in the 10 mM and 50 mM HCO<sub>3</sub><sup>-</sup> were an unexpected results since Fe content was generally reported to decrease with increasing bicarbonate concentrations. Many researchers<sup>8,14,31</sup> reported that high HCO<sub>3</sub><sup>-</sup> ion concentration makes Fe insoluble or hampers Fe uptake, possible also affecting translocation of this element within the plant itself. Brown and Jolley<sup>32</sup> reported that there was no correlation between Fe chlorosis in grapes and high bicarbonate concentrations. Pearce *et al.*<sup>26</sup> demonstrated that Fe concentration in the tobacco seedlings was not significantly affected by HCO<sub>3</sub><sup>-</sup>. Cu and Zn concentrations in the explants were not significantly affected by HCO<sub>3</sub><sup>-</sup>

In conclusion, in Gisela 5 rootstock, tissue culture technique was used as a rapid screening method to identify growth rate, chlorophyll content and ion accumulation associated with bicarbonate enriched conditions. Affected by  $HCO_3^-$  Gisela 5 explants grown *in vitro* showed the typical necrosis within few days after the beginning of the experiments. Necrosis started old leaves as yellowing and evolved in a complete browning of the entire shoot. Viability loss was observed in Gisela 5 due to  $HCO_3^-$ . By increasing  $HCO_3^-$  concentrations of the culture medium Ca and K concentrations of explants decreased, whereas Na, Mg and Mn concentrations increased. This is the first report documenting some physiological and biochemical responses of Gisela 5 to  $HCO_3^-$ . Further research on this subject should be conducted to clarify the effects of  $HCO_3^-$ .

## REFERENCES

- 1. A. Shahabi, M.J. Malakouti and E. Fallahi, J. Plant Nutr., 28, 1663 (2005).
- 2. F. Dehghani, F. Alaee Yazdi and M.J. Malakouti, Technical Note No. 206, Agricultural Education Publications Karaj (2002).
- 3. E. Peiter, F. Yan and S. Schubert, J. Plant Nutr. Soil Sci., 164, 165 (2001).
- 4. P.M. Kopittke and N.W. Menzies, Plant Soil, 266, 343 (2004).
- 5. E. Alcantara, F.J. Romera and D.L. Guordian, J. Plant Nutr., 11, 65 (1988).
- 6. A.R. Alhendawi, V. Römheld, E.A. Kirby and H. Marschner, *J. Plant Nutr.*, **20**, 1731 (1997).

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- 7. H.U. Kosegarten, B. Hoffman and K. Mengel, Plant Physiol., 121, 1069 (1999).
- 8. L. Brancadore, G. Tamai, B. Zocchi and O. Failla, Acta Hort., 564, 359 (2001).
- 9. E. Exadaktylou and T. Thomidis, Sci. Hort., 106, 125 (2005).
- 10. S. Franken-Bembenek, Acta Hort., 667, 167 (2005).
- 11. F. Cinelli, M. Fisichella and R. Muleo, J. Plant Nutr., 26, 2277 (2003).
- 12. R. Muleo, F. Cinelli and R. Viti, J. Plant Nutr., 18, 91 (1995).
- 13. L. Lombardi, L. Sebastiani and C. Vitagliano, J. Plant Nutr., 26, 2149 (2003).
- 14. Y. Shi, D.H Byrne, R.H. Loeppert and D.W. Reed, J. Plant Nutr., 16, 1675 (1993).
- M.D. De La Guardia, A. Felipe, E. Alcantara, J.M. Fournier and F.J. Romera, Iron Nutrition in Soils and Plants, Academic Publishers, pp. 201-205 (1995).
- 16. R.C. Pearce, Y. Li and L.P. Bush, J. Plant Nutr., 22, 1069 (1999).
- 17. J.Bialczyk, Z.Lechowski and A.Libik, J. Plant Nutr., 27, 1687 (2004).
- 18. X. Yang, R. Hajiboland and V. Römheld, J. Plant Nutr., 26, 399 (2003).
- 19. K. Zribi and M. Gharsalli, J. Plant Nutr., 25, 2143 (2002).
- 20. L. Bavaresco, E.Giachino and S.Pezzutto, J. Plant Nutr., 26, 1451 (2003).
- 21. R. Ksouri, M. Gharsalli and M. Lachaal, J. Plant Physiol., 162, 335 (2005).
- 22. B.A. Coulombe, R.L. Chaney and W.J. Wiehold, J. Plant Nutr., 7, 411 (1984).
- 23. K. Mengel and G. Geurtzen, J. Plant Nutr., 9, 161 (1986).
- 24. Z. Bie, T. Ito and Y. Shinohara, Sci. Hort., 99, 215 (2004).
- 25. N. Tremblay, J. Masson and A. Gosselin, Acta Hort., 238, 119 (1989).
- 26. R.C. Pearce, Y. Li and L.P. Bush, J. Plant Nutr., 22, 1079 (1999).
- 27. F. Cinelli and R.Viti, J. Plant Nutr., 1, 65 (1995).
- 28. P.F. White and A.D. Robson, Plant Soil, 125, 39 (1990).
- 29. F.J. Romera, E. Alcantara and M.D Guardia, Plant Soil, 130, 115 (1991).
- 30. T. Zaharieva, J. Plant Nutr., 9, 3 (1986).
- 31. F. Cinelli, J. Plant Nutr., 18, 77 (1995).
- 32. J.C. Brown and V.D. Jolley, J. Plant Nutr., 9, 175 (1986).

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