

Phytochemical Studies on *Parthenium hysterophorus* Linn.

P.R. SANTHI and P. ANBUSRINIVASAN*

Department of Chemistry, A.V.C. College (Autonomous), Mayiladuthurai-609 305, India

E-mail: panbusrinivas@gmail.com; panbu_srinivas@yahoo.co.in

Various fractions obtained from *Parthenium hysterophorus* contains antiinflammatory activities. The fraction obtained from diethyl ether was found to be a flavonol galangin. Aglycone and glucose fractions were also obtained during fractionation. All the fractions were characterized by UV-spectral analysis. The ethyl acetate fraction obtained was analyzed for its microbial activity, which showed a dose-dependent activity against heamolysis.

Key Words: Phytochemical, *Parthenium hysterophorus* Linn.

INTRODUCTION

Parthenium hysterophorus Linn. is a shrub found as weeds throughout Tamilnadu. It belongs to Asteraceae a sub order of compositae, contains 400 species¹. Crushed leaves of *P. hysterophorus* is used for snake bite and spider bite. The plant is also used for cholagogue². In present studies, the flowers of *P. hysterophorus* have been examined for their polyphenolic constituents and the results are presented.

EXPERIMENTAL

About 1 kg of fresh flowers of *P. hysterophorus* collected in and around A.V.C. College campus at Mannampandal during October were extracted with 95 % ethanol under reflux. Above 2 L of AnalaR grade ethanol is used for extraction. The alcoholic extract was concentrated *in vacuo* and the aqueous concentrate was fractionated with above 750 mL of benzene, 1 L of peroxide-free diethyl ether and 2 L of ethyl acetate. No crystalline solid could be recovered from benzene fraction.

The residue from diethyl ether fraction was taken up in ice-chest for a few days, when yellow needles were separated. It was identified as a flavonol-galangin. It had $\lambda_{\max}^{\text{MeOH}}$ nm 267, 305 sh, 359; + NaOMe 280, 327 sh, 412; + AlCl₃ 249, 273, 300 sh, 334, 412; + (AlCl₃ + HCl) 249, 274, 302 sh, 334, 412; NaOAc 275, 301 sh, 328 sh, 388, + (NaOAc + H₃BO₃) 267, 300 sh, 361.

It is soluble in organic solvents and sparingly soluble in hot water. It gave a golden yellow colour with NaOH, orange red colour with Mg-HCl

and violet colour with FeCl_3 . It responded Wilson's boric acid³, Horhammer-Hansel⁴ and Gibb's⁵ tests but did not respond to Molisch's test. R_f values as given in Table-1, which was identified as galangin. This was further confirmed by mixed paper chromatography and m.m.p. with an authentic sample of galangin, isolated from *Alpinia officinarum*.

TABLE-1
 R_f (X100) VALUES OF THE CONSTITUENTS OF THE FLOWERS OF
Parthenium hysterphorus
 (Whatmann filter paper No. 1, Ascending $30 \pm 2^\circ\text{C}$)

Compound	Developing solvents*							
	A	B	C	D	E	F	G	H
Glycoside obtained from ethyl acetate fraction	18	26	47	69	82	79	60	85
Aglycone from the diethyl ether fraction	8	-	9	21	56	96	92	71
Galangin (authentic)	8	-	10	22	56	95	94	72

*Solvent key; A= H_2O ; B = 5 % Acetic acid; C =15 % Acetic acid; D = 30 % Acetic acid; E = 60 % Acetic acid; F = *n*-BuOH:AcOH: H_2O = 4:1:5 (upper phase) (BAW); G = Water saturated phenol; H = AcOH:conc. HCl: H_2O = 30:3:10 (Forestal).

The fraction obtained from ethyl acetate was concentrated *in vacuo* and left in an ice for a week. A yellow solid separated was filtered and studied. It was recrystallized from methanol to obtain yellow crystals (Yield 0.005 %). It was identified as flavonol glucoside-galangin-3-O-glucoside. It is soluble in ethyl acetate, methanol and sparingly in water. It gave olive green with alcoholic Fe^{3+} , deep pink colour with Mg-HCl, yellow colour with NaOH and appeared deep purple under UV as well as on exposure to NH_3 . It did not respond Horhammer Hansel test but responded to the Wilson's boric acid Gibb's and Molisch's test. It had R_f as indicated in Table-1 and had $\lambda_{\text{max}}^{\text{MeOH}}$ nm 250, 310sh, 340; + NaOMe 265, 327sh, 356; AlCl_3 270, 320, 385, + AlCl_3 + HCl 270, 317, 385; NaOAc 270, 330 and NaOAc/ H_3BO_3 251, 315sh, 337.

Hydrolysis of the glycoside: The glycoside (0.05 g. 0.02 m mol) was dissolved in hot aqueous methanol (2 mL, 50 %) and an equal volume of H_2SO_4 (7 %) was added to it. The reaction mixture was refluxed at 100°C for 2 h. The excess of alcohol was distilled off from the hydrolyzate and the resulting aqueous solution was diluted with more water and left under chilled conditions for 2 h. A yellow solid separated was filtered washed and dried. The aqueous filtrate and the washings were extracted with diethyl ether. The dry yellow residue on the filter paper was combined with the residue from the dried diethyl ether and studied for the aglycone.

Identification of aglycone: (flavonol:galangin): The yellow coloured aglycone on recrystallization from methanol afforded a yellow crystalline solid, m.p. 278-280 °C which was identified as galangin by colour reactions, behaviour under UV and R_f (Table-1).

Identification of sugar: (glucose): The aqueous hydrolyzate after the removal of the aglycone was neutralized with $BaCO_3$ and filtered. The concentrated filtrate on PC gave R_f values as shown in Table-2 corresponding to those of glucose. The identification of the sugar was further confirmed by direct comparison with an authentic sample of glucose as also by preparation its osazone and recording its m.p. A quantitative hydrolysis of the same by the Folin-Wu's micro method revealed it to be a monoside.

TABLE-2
 R_f (X100) VALUES OF THE SUGAR FROM THE GLYCOSIDE OF
Parthenium hysterophorus
(Whatmann filter paper No. 1, Ascending 30 ± 2 °C)

Sugar	Developing solvents*			
	E	F	G	H
Sugar from the hydrolyzate of EtOAc fraction	77	9	39	90
Glucose (authentic)	77	9	39	90

*Solvent key; E = 60 % Acetic acid; F = *n*-BuOH:AcOH:H₂O = 4:1:5 (upper phase) (BAW); G = Water saturated phenol; H = AcOH:conc. HCl:H₂O = 30:3:10 (Forestal).

Microbial activity: Fresh blood was collected from healthy sheep and mixed with equal volume of sterilized solution containing 2 % dextrose, 0.8 % sodium citrate, 0.05 % citric acid and 0.42 % NaCl and used within 5 h. Hyposaline (0.36 % 2 mL), phosphate buffer (0.15 M, pH = 7.4, 1 mL) and SRBC (1 %, 0.5 mL) were taken in a number of tubes. Solutions of different concentrations of the drug were added in all tubes except one tube. The last tube served as control in which instead of the drug, isosaline (0.85 %, 1 mL) was added. The contents were incubated at 37 °C for 0.5 h and then centrifuged. The intensity of the supernatant was measured at 560 nm using a photoelectric colourimeter. The control tube that contained no drug was taken as representing 100 % SRBC lysis. The percentage of prevention of drug-treated hyposaline-induced SRBC lysis was calculated from the difference in absorbance readings of the control and drug-treated tubes using the following relation:

$$100 - \frac{\text{Reading corresponding to drug treated sample}}{\text{Reading corresponding to control}} \times 100$$

The results are given in Table-3.

TABLE-3
EFFECT OF ETHYL ACETATE ISOLATE AGAINST
HYPOTONICITY INDUCED HAEMOLYSIS

Conc. of the drug (μg)	Prevention (%)	Conc. of the drug (μg)	Prevention (%)
15	82	65	74
25	85	75	67
35	85	85	60
45	82	95	53
55	79	105	45

RESULTS AND DISCUSSION

The fresh flowers of *P. hysterophorus* have been found to contain galangin and its 3-O-glucoside.

The UV spectrum of the flavonol aglycone obtained from Et₂O fraction exhibited two major peaks at 359 nm (band I) and 267 nm (band II) which showed flavonol skeleton. A bathochromic shift of 53 nm on addition of NaOMe revealed the absence of a free 4'-OH in the B-ring. A shift of 53 nm on the addition of AlCl₃/HCl showed the presence of a free 5-OH in the A-ring. The presence of a free OH at C-7 was ascertained by a shift of + 8 nm (band II) on the addition of NaOAc. The AlCl₃ spectrum was almost same as that of AlCl₃/HCl revealing the absence of a catechol type of substitution in B-ring. The H₃BO₃ spectrum also confirmed it as there was only + 2 nm shift on the addition of NaOAc/H₃BO₃.

The UV absorption of the glycoside is at 340 nm. A comparison of band I absorption of the glycoside and the aglycone reveals that there may be 3-glycosylation in the flavonol. A bathochromic shift of only 25 nm (band I) ascertained the absence of a free OH at C-3. The presence of 5-OH groups was confirmed by the bathochromic shift of + 45 nm on the addition absorption peaks to reveal the absence of an *ortho*-dihydroxy grouping in B-ring. The presence of a free OH at C-7 was evidenced from the + 20 nm the required bathochromic shift indicating the absence of catechol type of substitution in B-ring. On this basis the identify of the pigments obtained from Et₂O and EtOAc solubles can be confirmed as galangin and its 3-glucoside respectively.

The SRBC membrane stabilization effects of EtOAc fraction can be inferred from the data presented in Table-3 which showed a dose-dependent activity against heamolysis. It has maximal inhibition of hypotonicity (haemolysis) induced SRBC lysis at a concentration of 30 μg .

REFERENCES

1. K.R. Kirtikar, B.D. Basu and An I.C.S Indian Medicinal Plants, Periodical Experts Book Agency, New Delhi, India, Vol. 2, p. 1337 (1990).
2. D.A. Bustos, A.A. Tapia, G.E. Feresion and L.A. Espinar, *Fitotesapia*, **67**, 411 (1996).
3. C.W. Wilson, *J. Am. Chem. Soc.*, **61**, 2303 (1939).
4. L. Horhammer and R. Hansel, *Arch. Pharm., Beryl*, **288**, 315 (1955).
5. F.E. King, T.T. King and L.C. Manning, *J. Chem. Soc.*, 563 (1957).

(Received: 23 December 2006;

Accepted: 29 September 2007)

AJC-5928

SYNTHETIC HETEROCYCLIC CHEMISTRY DESIGN SYNTHESSES, DEVELOP PROCESSES

13 — 14 OCTOBER 2008

LONDON, UK

Contact:

Scientific Update LLP, Maycroft Place, Stone Cross
Mayfield, East Sussex, TN20 6EW, United Kingdom
Telephone +44 (0) 1435 873062
Fax Number +44 (0) 1435 872734
Email sciup@scientificupdate.co.uk

POLYMER COLLOIDS: FROM DESIGN TO BIOMEDICAL AND INDUSTRIAL APPLICATIONS

20 — 24 JULY 2008

INSTITUTE OF MACROMOLECULAR CHEMISTRY AS CR
HEYROVSKÉHO NÁM. 2 PRAGUE, CZ 162 06, THE CZECH
REPUBLIC

Contact:

Dr. Daniel Horák
Institute of Macromolecular Chemistry AS CR, v. v. i.
Heyrovského nám. 2, Prague, CZ 162 06, The Czech Republic
(+420) 296 809 332; (+420) 296 809 410
email: pcol08@imc.cas.cz