

Extraction of Cadmium(II) Using Cloud-Point Method and Determination by FAAS

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A new, simple and versatile cloud-point extraction (CPE) methodology has been developed for type equation here for the separation and pre-concentration of cadmium. The metals in the initial aqueous solution were complexed with 2-(2-benzothiazolylazo)-2-*p*-cresol (BTAC) and Triton-X-114 was added as surfactant. Dilution of the surfactant-rich phase with acidified methanol was performed after phase separation and the cadmium contents were measured by flame atomic absorption spectrometry. The variables affecting the cloud-point extraction were optimized using a Box-Behnken design. Under the optimum experimental conditions, enrichment factors of 29 and 25 were achieved for cadmium. The accuracy of the method was evaluated and confirmed by the inductively coupled plasma atomic emission spectrometric analysis. The limits of detection expressed to solid sample analysis were $0.1 \mu\text{g g}^{-1}$ (Cd). The precision for 10 replicate measurements of $75 \mu\text{g L}^{-1}$ Cd was 0.9. The method has been successfully applied to the analysis of water samples.

Key Words: Cadmium, Cloud-point extraction, Water samples.

INTRODUCTION

In recent years, pollution of the environment by heavy metals has received considerable attention. These elements accumulate in living organisms and are of high toxic potential. Their wide technological use (fertilizers, mining, pigments), as well as their production from burning oil and coal and incineration of waste causes an extensive anthropogenic contamination of soil, air and water¹. Several analytical techniques such as flame atomic absorption spectrometry (FAAS)^{2,3}, inductively coupled plasma atomic emission spectrometry (ICP-AES)⁴ and inductively coupled plasma mass spectrometry (ICP-MS)⁵ are available for the determination of trace metals with enough sensitivity for the most applications. Despite good developments in the modern analytical instruments, which allow great enhancement in aspects of analysis, in many cases the available analytical instrumentation does not have enough sensitivity for the analysis of natural samples. Sample preparation is still a bottleneck for overall throughput because the involved steps often employ large volumes of hazardous organic solvents, are time consuming and/or expensive¹. Although, the determination of trace metal ions in natural waters is difficult due to

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various factors, particularly their low concentrations and matrices effects. Pre-concentration and separation can solve these problems and can lead to a higher confidence level and easy determination of the trace elements. Several procedures have been developed for the separation and preconcentration of contaminants from environmental matrices, such as liquid-liquid extraction (LLE)⁶⁻⁸, co-precipitation⁹⁻¹¹ and solid phase extraction (SPE)¹²⁻²⁰.

Although, disadvantages such as significant chemical additives, solvent losses, complex equipment, large secondary wastes, unsatisfactory enrichment factors and high time consumption, limit the application of these techniques. These problems could be addressed by the development of modular and compact processes that provide adequate separation and preconcentration without complex processes. The solvent microextraction technique effectively overcomes these difficulties by reducing the amount of organic solvent as well as allowing sample extraction and preconcentration to be done in a single step. The technique is faster and simpler than conventional methods. It is also inexpensive, sensitive and effective for the removal of interfering matrices. Solvent microextraction is a form of solvent extraction with phase ratio values higher than 100. Compared with the conventional solvent extraction, microextraction may provide poorer analyte recovery, instead the concentration in the organic phase greatly enhances. In addition, the amount of the used organic solvent is highly reduced and only one step of manipulation is necessary, therefore, problems of contamination and loss of analytes vanishes. Cloud point extraction (CPE)²¹⁻²⁴, homogeneous liquid-liquid extraction (HLLLE)^{25,26} and single drop microextraction (SDME)²⁷⁻³⁰ are fairly new methods of sample preparation which are used in separation and preconcentration of metals and can solve some of the problems encountered with the conventional pretreatment techniques. In the previous researches, we demonstrated a novel microextraction technique, named dispersive liquid-liquid microextraction (DLLME), which was successfully used, for the extraction and determination of polycyclic aromatic hydrocarbons (PAHs), organophosphorus pesticides (OPPs) and chlorobenzenes in water samples^{31,32}. Dispersive liquid-liquid microextraction is a modified solvent extraction method and its acceptor-to-donor phase ratio is greatly reduced comparing with the other methods. In DLLME, the appropriate mixture of the extraction and disperser solvents is rapidly injected by syringe into aqueous samples containing analytes, thereby, cloudy solution forms. In fact, the cloudy state results from the formation of fine droplets of the extraction solvent, which disperse in the sample solution. Then, this cloudy solution should be centrifuged and the fine droplets sediment at the bottom of the conical test tube. The determination of analytes in sedimented phase can be performed by instrumental analysis. In this extraction method any component in the solution, directly or indirectly after previous (or simultaneous) derivatization reaction, interacts with the fine droplets of the extraction solvent and consequently gets extracted from the initial solution and concentrates in the small volume of the sedimented phase. Simplicity of the operation, rapidity, low sample volume, low cost, high recovery and high enrichment factor are some advantages of DLLME.

Dispersive liquid-liquid microextraction is a miniaturized sample pre-treatment technique. On the other hand, graphite furnace atomic absorption spectrometry (GFAAS) is a microamount sample analysis technique. Therefore, it makes it perfect when a combination of both DLLME and GFAAS is used. The applicability of the approach has been demonstrated for the determination of cadmium in water samples. This element was selected for evaluation of the procedure because cadmium is one of the principal heavy metals of analytical interest due to its extreme toxicity even at relatively low concentrations³³⁻³⁷.

The aim of this work is to apply a thiazolylazo reagent, 2-(2-benzothiazolylazo)-2-*p*-cresol (BTAC) in a cloud-point extraction (CPE) procedure for determination of cadmium in water samples. Some analogous compounds of BTAC such as 2-(6-methyl-2-benzothiazolylazo)-5-diethyl aminophenol (MBTAE)^{38,39} and 2-(2-thiazolylazo)-5-diethylaminophenol (TADAP)^{40,41} have been applied as a precolumn derivatizing reagent in high-performance liquid chromatography (HPLC). Another analogue, 2-(6-bromo-2-benzothiazolylazo)-5-diethyl aminophenol, was synthesized and applied to the spectrophotometric determination of nickel⁴² and zirconium⁴³. According to our best of knowledge, the use of BTAC in analytical procedures has not been reported before.

EXPERIMENTAL

Determination of Cd²⁺ contents in working samples were carried out by a Varian spectra A.200 model atomic absorption spectrometer equipped with a high intensity hollow cathode lamp(HI-HCl) according to the recommendations of the manufacturers. Separation of sorbent was assisted using a centrifuge (centurion scientific model: K 240R, West Sussex, U.K.). The pH measurements were carried out by an ATC pH meter (EDT instruments, GP 353).

All chemicals and reagents used in this study were of analytical-reagent grade. Deionized water was used to prepare all solutions. The laboratory glassware was kept in dilute nitric acid at least overnight and subsequently washed with deionized water. Solutions of the non-ionic surfactant Triton-X-114 (Merck) were prepared in high purity deionized water. 2-(2-Benzothiazolylazo)-2-*p*-cresol solutions were prepared by dissolving appropriate amounts of 2-(2-benzothiazolylazo)-2-*p*-cresol (BTAC) prepared in absolute ethanol (Merck, Darmstadt, Germany). 1000 mg/L stock standard solutions of Cd(II) were purchased from Merck. Nitric acid solutions were prepared by direct dilution with deionized water from the concentrated solutions (Merck). Methanol (Merck) was used to decrease the viscosity of surfactant-rich phase. Acetate (4.6), phosphate (6.3) and borate (7.5 and 8.0) buffers were used to adjust the sample pH. Sodium carbonate (Merck), 2-ethylhexyl-amine (Sigma, Aldrich), sodium nitrite (Merck), ethanol (Merck) were used for synthesis of BTAC. The accuracy of the method was assessed by analyzing the following certified reference materials (CRM): NIST 1515 Apple Leaves, NIST 1570a Spinach Leaves and NIST 1573a Tomato Leaves from the National Institute of Standards and Technology (Gaithersburg, MD, USA).

Synthesis of BTAC: BTAC was obtained by coupling diazotized 2-amino-benzothiazole with *p*-cresol in acid solution at 0.5 °C as described previously⁴⁴. 2-Aminobenzothiazole was dissolved (6.0 g) in 120 mL of a 6 mol L⁻¹ sulfuric acid solution. To this solution was added dropwise a solution of 3.0 g of sodium nitrite in 20 mL of water at 0.5 °C and the mixture was stirred and kept at 0.5 °C for 1 h. For coupling, 4.3 g of *p*-cresol was added to 20 mL of a 1 mol L⁻¹ sodium carbonate solution and the mixture was cooled to 0.5 °C. This solution was added dropwise to the above diazotized solution with vigorous stirring. The system was allowed stand overnight in refrigerator at 4 °C. The orange color precipitate formed was filtered and purified by recrystallization with isopropyl alcohol. A yield of about 90 % was obtained. Melting point was determined five times and varied in the range of 158-160 °C. Elemental analysis (%): C₁₄H₁₁N₃OS requires C 62.46, H 4.09, N 15.60, S 11.91; found: C 62.12, H 3.99, N 15.53, S 11.85.

Samples and sample pretreatment: Water samples were taken from the Tap water (Tehran, taken after 10 min operation of the tap), rain water (Tehran, 26 January, 2007), snow water (Saveh , 6 February, 2007) and sea water (taken from Caspian sea, near the Mahmoud-Abad shore) at Iran. After cooling at room temperature these solutions were adjusted to pH with a 10 % (w/v) sodium hydroxide. The solution was made up to required volume with deionized water into a 50 mL volumetric flask. The same procedure was used for the blank solutions. Spiked samples were also prepared in order to calculate the recovery of the digestion procedure.

Cloud-point extraction procedure: An aliquot (10 mL) of a cadmium buffered solution was transferred to a 15 mL centrifuge tube. Appropriate amounts of BTAC and Triton-X-114 solutions were added. The sample solution was left to equilibrate in a thermostated bath for 15 min at 40 °C. The system was then centrifuged at 3500 rpm for 20 min to achieve complete separation of the two phases. The phases were cooled down in an ice bath in order to increase the viscosity of the surfactant-rich phase. After the cloud-point extraction, the aqueous phase was decanted by inverting the tube. Later 200 µL of a 1:1 (v/v) methanol; 1 mol L⁻¹ HNO₃ solution was added to the surfactant-rich phase. The resultant solution was directly introduced into the FAAS by conventional aspiration and the cadmium content was measured.

RESULTS AND DISCUSSION

Optimization of the experimental conditions for CPE: In order to investigate the influence of the main variables in CPE procedure, a Box. Behnken design was employed²⁸. The variables: pH, BTAC concentration (BC) and surfactant concentration (SC) were optimized. Table-1 shows the experiments executed for this design. The data in Table-1 are described by a quadratic model in the three factor variables. The equation that represents the relationship among the analytical signal for Cd (RCd) and pH, BTAC concentration and surfactant concentration are the following:

TABLE-1
VARIABLES FOR THE BOX-BEHNKEN DESIGN WITH REAL AND CODED VALUES

Experiment	pH	Surfactant concentration (% v/v)	BTAC concentration (mol L ⁻¹)	Analytical signal for cadmium
1	4.6 (-1.0)	1.0 (-1.0)	1.84 × 10 ⁻³ (0.0)	0.1700
2	8.0 (1.0)	1.0 (-1.0)	1.84 × 10 ⁻³ (0.0)	0.1800
3	4.6 (-1.0)	2.0 (1.0)	1.84 × 10 ⁻³ (0.0)	0.1720
4	8.0 (1.0)	2.0 (1.0)	1.84 × 10 ⁻³ (0.0)	0.1680
5	4.6 (-1.0)	1.5 (0.0)	1.23 × 10 ⁻³ (-1.0)	0.1660
6	8.0 (1.0)	1.5 (0.0)	1.23 × 10 ⁻³ (-1.0)	0.1810
7	4.6 (-1.0)	1.5 (0.0)	2.46 × 10 ⁻³ (1.0)	0.1680
8	8.0 (1.0)	1.5 (0.0)	2.46 × 10 ⁻³ (1.0)	0.1840
9	6.3 (0.0)	1.0 (-1.0)	1.23 × 10 ⁻³ (-1.0)	0.1900
10	6.3 (0.0)	2.0 (1.0)	1.23 × 10 ⁻³ (-1.0)	0.1990
11	6.3 (0.0)	1.0 (-1.0)	2.46 × 10 ⁻³ (1.0)	0.1840
12	6.3 (0.0)	2.0 (1.0)	2.46 × 10 ⁻³ (1.0)	0.2000
13	6.3 (0.0)	1.5 (0.0)	1.84 × 10 ⁻³ (0.0)	0.2160
14	6.3 (0.0)	1.5 (0.0)	1.84 × 10 ⁻³ (0.0)	0.2240
15	6.3 (0.0)	1.5 (0.0)	1.84 × 10 ⁻³ (0.0)	0.2280

$$\begin{aligned}
 \text{RCd} = & 0.2227 * + 0.0046 (\text{pH}) - 0.0343 * (\text{pH})^2 \\
 & + 0.0019 (\text{SC}) - 0.0158 * (\text{SC})^2 - 0.0136 (\text{BC})^2 \\
 & - 0.0035 (\text{pH}) (\text{SC}) + 0.0002 (\text{pH}) (\text{BC}) \\
 & + 0.0017 (\text{SC}) (\text{BC})
 \end{aligned}
 \tag{1}$$

values marked with * are significant. Considering eqn. 1, a maximum point is predicted for the codified factor levels of 0.065 for pH, 0.052 for surfactant concentration and 0.0039 for BTAC concentration. Corresponding real values from data of zinc are: 6.4 (pH), 1.53 % (v/v) (surfactant concentration) and 1.84 × 10⁻³ mol L⁻¹ (BTAC concentration). The results from mathematical adjustment by surface response graphs can be seen in Fig. 1. The critical point of surface response from data of cadmium was characterized as a saddle point. This point is a stationary point of a surface response which presents the maximum response for the levels of some variables and simultaneously the minimum response for the levels of other variables of the system⁴⁵. The results obtained separately for cadmium agree with the results calculated using the multiple response function which were: 7.5 (pH), 1.0 % (v/v) (surfactant concentration) and 1.9 × 10⁻³ mol L⁻¹ (BTAC concentration)^{45,46}.

Analytical features: Calibration graphs obtained with preconcentration gave good linearity over the concentration range of 1.0-100.0 µg L⁻¹ of cadmium (Abs = 8.67 × 10⁻³ + 2.84 × 10⁻³ C). Abs is the absorbance and C is the metal concentration, in µg L⁻¹. By using metal standard solutions with cadmium concentrations in the range from 50.0 to 2000.0 µg L⁻¹, calibration graphs for metals without preconcentration were also obtained. These calibration graphs were: Abs = 2.20 × 10⁻³ + 9.80 × 10⁻⁵ C for cadmium. The enrichment factors of metals were based on

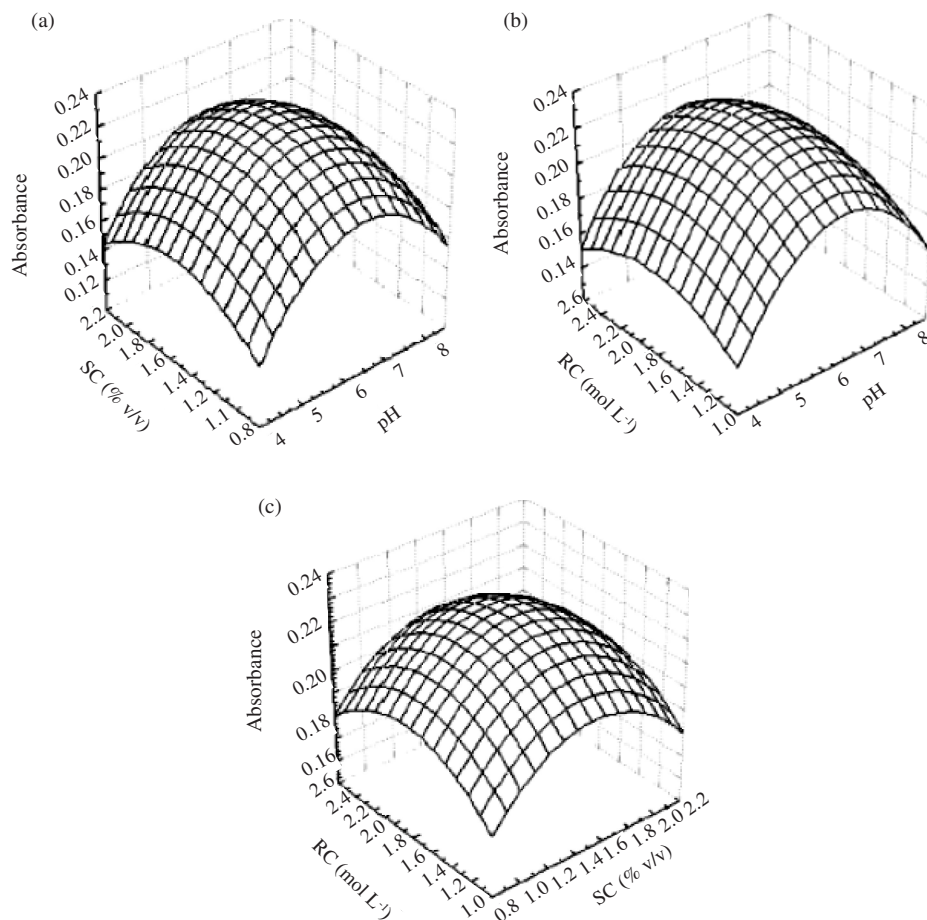


Fig. 1. Response surface plot of the cadmium analytical signal as a function of (A) pH and surfactant concentration (B) pH and reagent concentration and (C) surfactant concentration and reagent concentration

matching the slopes of calibration graphs of preconcentration procedure and those by a conventional FAAS nebulization method. The enrichment factors of cadmium was 29-fold. The limit of detection (LOD) was calculated based on three times ($3s$) of standard deviation of blank signal by 11 replicate measurements⁴⁷. LODs was $0.9 \mu\text{g L}^{-1}$ for cadmium. The limit of quantification (LOQ) is the concentration that gives a response equivalent to ten times the standard deviation of blank signal ($n = 11$) and define the lower limit of the range. The limit of quantification was also calculated for cadmium ($1.5 \mu\text{g L}^{-1}$). The precision (determined as the relative standard deviation) for 10 replicate measurements of $75 \mu\text{g L}^{-1}$ cadmium was 0.9.

Interference: The interference of foreign ions on the CPE determination of $100.0 \mu\text{g L}^{-1}$ Cd was studied. The tolerance limits of various foreign species on the

sorption of the metal ions are given in Table-2. These tolerance limits were taken as that value which caused an error of not more than 5 % in the absorbance reading. Most of the metal ions are tolerated up to 400 mg L⁻¹. The potential interferences from some common matrix cations such as Na(I), K(I) and Mg(II) were also investigated. These substances are tolerated at concentrations at least up to 30 g L⁻¹. The results obtained in these experiments demonstrate that the presence of large amounts of species commonly present in water samples have no significant effect on the CPE of cadmium.

TABLE-2
EFFECT OF FOREIGN IONS ON THE CLOUD-POINT
EXTRACTION OF 100.0 µg L⁻¹ CADMIUM

Substance	Concentration cadmium	Substance	Concentration cadmium
Al ³⁺	250.0 mg L ⁻¹	K ⁺	55.0 g L ⁻¹
Ba ²⁺	45.0 g L ⁻¹	Mg ²⁺	50.0 g L ⁻¹
Br ⁻	40.0 g L ⁻¹	Na ⁺	70.0 g L ⁻¹
Cu ²⁺	300.0 mg L ⁻¹	Ni ²⁺	4.0 mg L ⁻¹
Cl ⁻	50.0 g L ⁻¹	NO ₃ ⁻	30.0 g L ⁻¹
Co ²⁺	240.0 mg L ⁻¹	Pb ²⁺	600.0 mg L ⁻¹
Cr ₂ O ₇ ²⁻	12.0 mg L ⁻¹	SO ₄ ²⁻	30.0 g L ⁻¹
CrO ₄ ²⁻	12.0 mg L ⁻¹	Ca ²⁺	300.0 mg L ⁻¹
Fe ³⁺	18.0 mg L ⁻¹	-	-

Analysis of water samples: To assess the applicability of the method to real samples, it was applied to the extraction and determination of cadmium from different water samples. Tap water (Tehran, taken after 10 min operation of the tap), rain water (Tehran, 26 January, 2007), snow water (Saveh , 6 February, 2007) and Sea water (taken from Caspian sea, near the Mahmoud-Abad shore) samples were analyzed (Table-3). As is seen, the recovered cadmium ion reveals that the results are quite reliable and are in satisfactory agreement with those obtained by ICPAES.

TABLE-3
RECOVERY OF CADMIUM ADDED TO 1000 mL OF DIFFERENT WATER SAMPLES
(AFTER COOLING AT ROOM TEMPERATURE THESE SOLUTIONS WERE
ADJUSTED TO pH WITH A 10 % (w/v) SODIUM HYDROXIDE)

Sample	Cd ²⁺ added (µg)	Cd ²⁺ determined (ng mL ⁻¹)	ICP-AES
Tap water	0.0	1.75 (0.8) ^a	Not detected
	10.0	11.95 (0.6)	11.9
Snow water	0.0	4.84 (1.2)	Not detected
	10.0	14.90 (1.1)	14.4
Rain water	0.0	2.67 (1.3)	Not detected
	10.0	12.79 (1.0)	12.1
Sea Water	0.0	12.69 (1.1)	12.2
	10.0	22.98 (1.0)	23.6

^aValues in parentheses are % RSDs based on five individual replicate analysis.

Application: The results demonstrate the applicability of the method for interference-free determination of the metals. Applicability of the method to real samples was checked by the determination of cadmium in waters. The results of this investigation are given in Table-2. It can be seen that the recovery of spiked samples is good. The results indicate that the proposed method is applicable for routine monitoring of cadmium in these matrices. The limits of detection and limits of quantification expressed to solid sample analysis were: Cd (LOD = 0.1 $\mu\text{g g}^{-1}$, LOQ = 0.51 $\mu\text{g g}^{-1}$).

Conclusion

The reagent BTAC was successfully employed in a CPE procedure for determination of cadmium in water samples by FAAS. This study allowed the development of a rapid, easy to use, safe, environmentally friendly and inexpensive methodology for the preconcentration and separation of trace metals. The method significantly improved the performance of the FAAS detection for Cd. The method developed was simple, reliable and precise for determining cadmium in water. The proposed method was also free of interference compared to conventional procedures to determine cadmium⁴⁷⁻⁵².

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