

TLC-Densitometric Determination of Vitamins B₁, B₆ and B₁₂ in Pure and Pharmaceutical Formulations Using Treated Aleppo Bentonite

ABDUL AZIZ RAMADAN*, AMER BODAKJI and IBRAHIM MAHMOUD
Department of Chemistry, Faculty of Science, University of Aleppo, Aleppo, Syria
Fax: (963)(21)2633136; E-mail: dramadan@scs-net.org

Determination of vitamins B₁, B₆ and B₁₂ in pure and in pharmaceutical formulations by thin layer chromatography (0.30 mm thickness) using acid treated Aleppo bentonite and mobile phase consisted of iso-butyl alcohol:methanol:chloroform:acetic acid:ammonia (7.5:1:3.5: 0.4:0.5, v/v/v/v/v) has been applied. The specific surface area of bentonite was 168.25 m²/g and mean pore size was 52.3 Å determined using nitrogen adsorption (BET). Quantification was carried out densitometrically at $\lambda = 275$ nm for vitamins B₁, B₆ and B₁₂ (if their concentrations are in close proximity) and at $\lambda = 525$ nm for vitamin B₁₂ (when $C_{B_6} \approx C_{B_1} \gg C_{B_{12}}$ as in pharmaceutical products). This method is successful for vitamin B₁, B₆ and B₁₂ formulation even in the ratios between vitamin B₁₂ and others 1:100. The retardation factors (R_f) of B₁, B₆ and B₁₂ were 0.37, 0.68 and 0.26, respectively. Calibration curves were obtained in the range of 5.0-40.0 μ g/spot for vitamins B₁, B₆ and B₁₂ (at $\lambda = 275$ nm) for standard solutions, while for pharmaceutical products, vitamins B₁, B₆ were determined at $\lambda = 275$ nm and vitamin B₁₂ was determined at $\lambda = 525$ nm in the range of 0.5- 4.0 μ g/spot.

Key Words: Aleppo bentonite, BET, Thin layer chromatography, Vitamins B₁, B₆ and B₁₂.

INTRODUCTION

Aleppo bentonite is rocky clay which originates from volcanic ash and consists of 47 % SiO₂, 14.4 % Al₂O₃ and some other oxides as Fe₂O₃, MgO, CaO and Na₂O^{1,2}. Bentonite has large pore volumes and high specific surface area³. The thermal treatment causes decreasing of its specific surface area with increasing in the temperature of thermal treatment⁴. Bentonite clays are used in many industrial products and processes, drilling fluids, a certain lubricating grease^{5,6} and it can be used as chromatographic supports in gas chromatography to separate many mixtures after grafting with different methods^{7,8}. Bentonite is used as stationary phase in thin layer chromatography to separate some metal ions⁹.

Vitamins are organic compounds present mostly in food and are needed in very small amounts for various metabolic processes and other functions¹⁰. The two types of vitamins are named as fat-soluble and water soluble. The water-soluble vitamins are thiamine (vitamin B₁), riboflavin (vitamin B₂), niacin, pyridoxine (vitamin B₆),

folate, vitamin B₁₂, biotin and pantothenic acid. These vitamins are widely distributed in foods. They function as coenzymes that help the body obtain energy from food. They are also important for normal appetite, good vision, healthy skin, healthy nervous system and red blood cell formation¹¹.

Determination of water-soluble vitamins has always been a peculiar problem largely because of the instability of these compounds and the complexity of the matrices in which they usually exist. As their chemical structure is not related, a considerable number of publications have appeared using different physical, chemical and biological methods. For the determination of the compounds in single- and multi-component mixtures the number of papers is more restricted. They include, among others, the determination of vitamin B₁₂ as cobalt by electrothermal atomic absorption spectrometric methods¹², higher order derivative spectrophotometry¹³ and capillary electrophoresis¹⁴. Also, the determination of the B-complex mainly in tablets have been described extensively using HPLC methods^{15,16} or HPLC after solid-phase extraction¹⁷. Most methods are successful in single component preparations. Other chromatographic systems separate B₁, B₆ and B₁₂ when they exist in the same range but are hampered when the amount of B₁ and B₆ exceeds by a hundred or even a thousand times the amount of B₁₂ present in the complex^{18,19}.

EXPERIMENTAL

Surface area and pore size measurement (BET) were recorded using a Micromeritics Gemini III 2375 under nitrogen atmosphere (USA). Scanner-densitometer CD60 (Desega, Germany) equipped with mercury, tungsten and deuterium lamps, CAMAG Hand Operated TLC Coater for preparation of TLC plates (Switzerland), CAMAG UV Cabinet for assessing and marking thin layer chromatograms under UV light (Switzerland) and different size of syringe (Hamilton, Switzerland) were used.

Thiamine hydrochloride (vitamin B₁) (99.0 %, Jiangsu, China), pyridoxine hydrochloride (vitamin B₆) (99.6 %, Sinochem, China) and cyanocobalamine (vitamin B₁₂) (99.1 %, Hebei, China) were used. Methanol, chloroform, acetic acid were of analytical grade, Merck, Germany. Ammonia solution and iso-butyl alcohol were of analytical grade, BDH, England.

Preparation of stationary phase: Bentonite was crushed to obtain small pieces, which have diameter less than 45 µm, followed by washing with concentrated hydrochloric acid at boiling point for 30 h to remove soluble oxides especially iron oxide. Then it was washed several times with distilled water and dried at 120 °C for 3 h.

Preparation of TLC plates: For preparation of thin layer chromatography, 8.8 g treated bentonite was mixed with 0.7 g fluorescence substance (F₂₅₄), then the mixture was added to 20 mL hot water containing 0.5 g corn starch as binder to obtain homogeneous slurry. The slurry was spread over glass plates by an applicator to form uniform thin layer 0.30 mm thick. The plates were dried at 105 °C.

Mobile phase: Iso-butyl alcohol:methanol:chloroform:acetic acid:ammonia (7.5:1:3.5:0.4:0.5, v/v/v/v/v) were used for the development method as mobile phase.

Standard solutions: Stock solution (**A_i**) was prepared by dissolving 2.5 g of each vitamins B₁, B₆ and B₁₂ in 25 mL mixture of distilled water and methanol (1:1), then transferred into a 50 mL volumetric flask and the final volume was completed to 10 mL with the same mixture of solvents. Volumes 1, 2, 3, 4, 5, 6, 7 and 8 mL from the former solution were transferred into 50 mL volumetric flasks and completed to the mark with the same mixture of solvents (these solutions content: 5, 10, 15, 20, 25, 30, 35 and 40 mg mL⁻¹, respectively for each vitamin).

Stock solution (**B_i**) was prepared by dissolving 12.5 g of each vitamins B₁ and B₆ and 0.1250 g of vitamin B₁₂ in 40 mL mixture of distilled water and methanol (1:1), then transferred into a 50 mL volumetric flask and the final volume was completed to 50 mL with the same mixture of solvents. Volumes 1, 2, 3, 4, 5, 6, 7 and 8 mL from former solution were transferred into 10 mL volumetric flasks and completed to the mark with the same mixture of solvents (these solutions content: 0.25, 0.50, 0.75, 1.00, 1.25, 1.50, 1.75 and 2.00 mg mL⁻¹ of vitamin B₁₂).

Sample preparation: Twenty tablets were weighed and the average tablet weight determined (each tablet contains: 100 mg B₁, 100 mg B₆ and 1 mg B₁₂). The tablets were finely powdered and a portion of powder equivalent to the weight of eight tablets was dissolved in 20 mL mixture of distilled water and methanol (1:1) and vigorously shaken for a 20 min on a mechanical shaker, then filtrated and transferred into a 25 mL volumetric flask and the final volume was completed to 25 mL with the same mixture of solvents. This solution (**T_i**) contains: 32 mg B₁ mL⁻¹, 32 mg B₆ mL⁻¹ and 0.32 mg B₁₂ mL⁻¹).

The ampoules were directly used for analysis (each ampoule contains: 100 mg B₁, 100 mg B₆ and 1 mg B₁₂ in 3 mL).

Procedure (chromatographic conditions): 1 µL of standard solutions **A_i** were spotted on TLC-glass plates 20 cm × 10 cm pre-coated treated bentonite (F₂₅₄ with 0.30 mm thickness). Mobile phase were used for development method, then the plates were dried at room temperature and quantification was carried out densitometrically at λ = 275 nm for vitamins B₁, B₆ and B₁₂. This process was repeated five times for each concentrations and calibration curves were obtained in the range 5-40 µg/spot for B₁, B₆ and B₁₂.

2 µL of standard solutions **B_i** were spotted on TLC-glass plates 20 cm × 10 cm pre-coated treated bentonite (F₂₅₄ with 0.30 mm thickness). Mobile phase were used for development method, then the plates were dried at room temperature and quantification was carried out densitometrically at λ = 525 nm for vitamin B₁₂. This process was repeated five times for each concentrations and calibration curves were obtained in the range 0.50-4.00 µg/spot for B₁₂.

Pharmaceutical formulations (tablets and ampoules): 1 µL of solution (**T_i**), for tablets or 1 µL for ampoules were spotted on TLC-glass plates for determination of vitamins (B₁ and B₆) and quantification was carried out densitometrically at λ = 275 nm. The concentrations were calculated from standard curves in the range 5-40

$\mu\text{g}/\text{spot}$ for B₁ and B₆. 2 μL of solution (T_i), for tablets or 2 μL for ampoules were spotted on TLC-glass plates for determination of vitamin B₁₂ and quantification was carried out densitometrically at $\lambda = 525 \text{ nm}$. The concentrations were calculated from standard curves in the range 0.50-4.00 $\mu\text{g}/\text{spot}$ for vitamin B₁₂.

RESULTS AND DISCUSSION

Surface properties of treated bentonite: Surface areas of treated bentonite was determined by the adsorption of nitrogen at 77 K (BET). For determination of textural properties, the adsorption was carried out until near saturation ($P/P_0 \approx 1.0$), then the desorption was completed until closure of the hysteresis loop. Representative adsorption-desorption isotherms of nitrogen are shown in Fig. 1. The isotherms are IV type of Sing (1982) and BDDT (1940) classifications, which indicate the presence of mesoporous structure. Application of the linear BET equation to the nitrogen adsorption data obtained was within the range of relative pressures (0.02-0.25). The representation linear BET plots are shown in Fig. 2. From these plots, it is found that the BET surface areas (S_{BET}) was 168.25 m^2/g . The data also allows the determination of pore volume distribution (Fig. 3). The total pore volume v_p (0.440 mL/g) was determined from the adsorbed volume at $P/P_0 = 0.95$ in the liquid form. The mean pore radii r_a (52.3 \AA), were determined from the equation: $r_a = 2 \times 10^4 \times v_p/S_{\text{BET}}$. Results of surface properties for treated bentonite can be showed in Table-1.

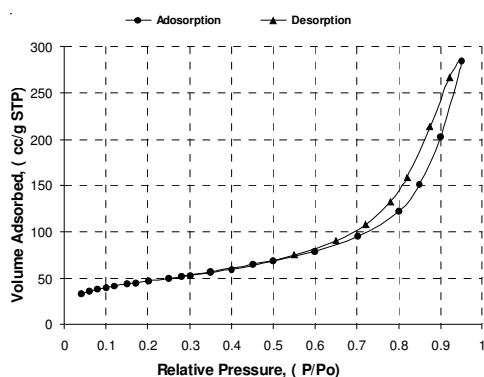


Fig. 1. Adsorption-desorption isotherm of nitrogen at 77 K on treated bentonite

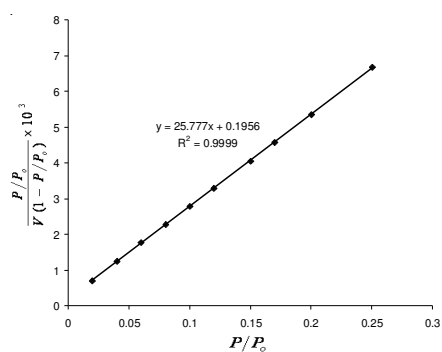


Fig. 2. BET plots for treated bentonite

TABLE-1
SURFACE PROPERTIES OF TREATED ALEPPO BENTONITE

S_{BET} (m^2/g)	v_p (mL/g)	r_a (\AA)
168.25	0.440	52.3

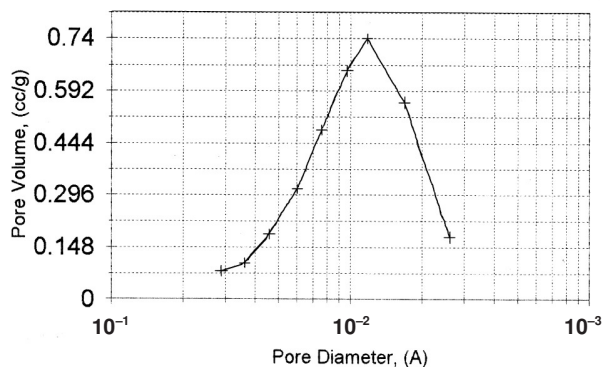


Fig. 3. Pore volume distribution of treated bentonite

Chromatograms processing: The position of the spots from the front on the chromatographic plate for different concentrations (5 to 40 $\mu\text{g}/\text{spot}$) of B₁, B₆ and B₁₂ at $\lambda = 275 \text{ nm}$ was shown in Fig. 4.

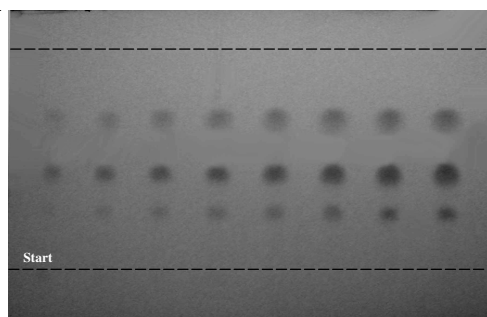


Fig. 4. TLC plate of standard vitamin B₁, B₆ and B₁₂ for concentrations of each vitamin 5, 10, 15, 20, 25, 30, 35 and 40 $\mu\text{g}/\text{spot}$ (mobile phase: iso-butyl alcohol:methanol:chloroform:acetic acid:ammonia 7.5:1:3.5:0.4:0.5 v/v/v/v/v)

The chromatogram of mixture of B₁, B₆ and B₁₂ (20 $\mu\text{g}/\text{spot}$ for each vitamin) can be observed with three peaks at different wavelengths (λ) at 235 to 315 nm (Fig. 5). The first peak for B₁₂ increase to $\lambda = 275 \text{ nm}$ then decrease, the second for B₁ decrease with increasing of λ and the third for B₆ increase to $\lambda = 275 \text{ nm}$ then sharply decrease (Fig. 6). It is inferred from the Figs. 4 and 5 that, the best wavelength to determine the vitamins B₁, B₆ and B₁₂ is 275 nm. The retardation factors (R_f) of B₁, B₆ and B₁₂ were 0.37, 0.68 and 0.26, respectively.

It is seen from the previous study that the peak areas of the three vitamins at the wavelength 275 nm were convergent, while showing only peak of vitamin B₁₂ at the wavelength 525 nm. When the concentration of vitamins B₁ and B₆ very larger than the concentration of vitamin B₁₂ (as in pharmaceutical formulations: $C_{B_6} = C_{B_1} = 100 C_{B_{12}}$) become identified vitamin B₁₂ very difficult and interfere with the peak of vitamin B₁. Therefore be possible to select the wavelength at $\lambda = 525 \text{ nm}$.

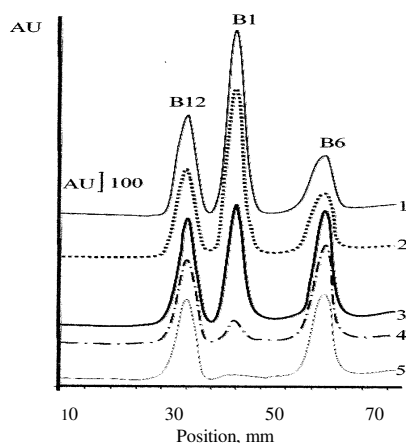


Fig. 5. Chromatograms of mixture of B₁, B₆ and B₁₂ (20 µg/spot) disposed at different wavelengths (λ) at 235 to 315 nm: 1- λ = 235, 2- λ = 255, 3- λ = 275, 4- λ = 295 and 5- λ = 315 nm

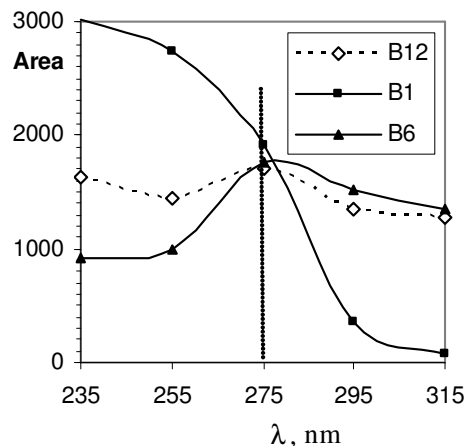


Fig. 6. Effect of wavelengths (λ) on peak areas of vitamins B₁, B₆ and B₁₂ by TLC-densitometric method using acid treated Aleppo bentonite and mobile phase: iso-butyl alcohol:methanol:chloroform:acetic acid: ammonia (7.5:1:3.5:0.4:0.5, v/v/v/v/v) ($C_{B_6} = C_{B_1} = C_{B_{12}} = 20$ µg/spot)

Quantitative evaluation: The method being validated through precision, linearity and accuracy for the determination of different standard mixtures of vitamins B₁, B₆ and B₁₂ in the range of 5.0 to 40.0 µg/spot using $\lambda = 275$ nm (Fig. 7, Table-2). Determination of vitamin B₁₂, when $C_{B_1} \gg C_{B_{12}}$; when it is not possible to use the wavelength 275 nm, the wavelength 525 nm was used (only the peak of B₁₂ appears). The calibration curve for vitamin B₁₂ at $\lambda = 525$ nm was studied. It was found that the equation of this curve in the concentration range of 0.50 to 4.0 µg/spot was as the following: $y = 158.11x + 10.902$ and $R^2 = 0.9996$.

Application: The results of the validation verify the fitness of the proposed analytical procedure for the identification and quantitative determination of the three vitamins in the mixture. Two different commercial products (that contained of the three studied vitamins) were analyzed in order to assign the values of their contents using $\lambda = 275$ nm for B₁ and B₆ and $\lambda = 525$ nm for B₁₂, because $C_{B_6} \approx C_{B_1} \gg C_{B_{12}}$. The pharmaceutical formulations were selected for the study as the following:

Tablets: Each tablet contains 100 mg vitamin B₁, 100 mg vitamin B₆ and 1 mg (1000 µg) vitamin B₁₂ (+ 20 % over dose of all vitamins for stability) of brand Vitaneurin Fort (manufactured by Asia Pharmaceutical Industries, Aleppo-Syria).

Ampoules: Each ampoule contains 100 mg vitamin B₁, 100 mg vitamin B₆ and 1 mg (1000 µg) vitamin B₁₂ (+ 20 % over dose of all vitamins) in 3 mL of brand Vitaneurin Fort (manufactured by Asia Pharmaceutical Industries, Aleppo-Syria).

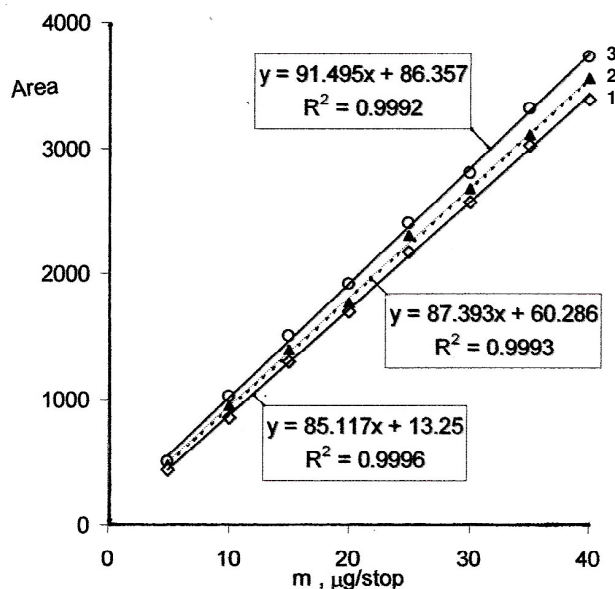


Fig. 7. Calibration curves for determination of vitamins B₁ (3), B₆ (2) and B₁₂ (1) in pure forms by TLC-densitometric method using acid treated Aleppo bentonite and mobile phase: iso-butyl alcohol:methanol:chloroform:acetic acid:ammonia (7.5:1:3.5:0.4:0.5, v/v/v/v/v) at $\lambda = 275$ nm

The three vitamins in the mentioned pharmaceutical formulations were simultaneously evaluated, on the same plate, reducing the time and the amount of the materials required for the analysis. The obtained results and their confidence intervals are listed in Table-3.

The results are in good agreement with the results of HPLC²⁰. It can be observed that the difference between the results by HPLC²⁰ and the found values by this method are less than 5 % in the worst case and the relative standard deviation is did not exceed ± 4.8 %. The proposed method has been successfully applied to determine vitamins B₁, B₆ and B₁₂ formulation even in the ratios between vitamin B₁₂ and others 1:100.

Conclusion

In the preceding method, determination of vitamins B₁, B₆ and B₁₂ in pure and pharmaceutical formulations by TLC-densitometric method using acid treated Aleppo bentonite and mobile phase: iso-butyl alcohol:methanol:chloroform:acetic acid:ammonia (7.5:1:3.5:0.4:0.5, v/v/v/v/v) has been applied. Quantification was carried out densitometrically at $\lambda = 275$ nm for vitamins B₁, B₆ and B₁₂ (if their concentrations were close) and at $\lambda = 525$ nm for vitamin B₁₂ (when $C_{B_6} \approx C_{B_1} \gg C_{B_{12}}$ as in pharmaceutical formulations). This method is successful for determination of vitamins B₁, B₆ and B₁₂ formulation even in the ratios between vitamin B₁₂ and others 1:100. The retardation factors (R_f) of B₁, B₆ and B₁₂ were 0.37, 0.68 and 0.26,

TABLE-2
 DETERMINATION OF VITAMINS B₁, B₆ AND B₁₂ IN PURE FORMS BY TLC-DENSITOMETRIC METHOD USING ACID TREATED ALEPPO BENTONITE
 AT $\lambda = 275$ nm (MOBILE PHASE: ISO-BUTYL ALCOHOL:METHANOL:
 CHLOROFORM:ACETIC ACID:AMMONIA 7.5:1:3.5:0.4:0.5 v/v/v/v/v)

Taken standard m ($\mu\text{g}/\text{spot}$)	Vitamin	Found $\bar{m} \pm \text{SD}$ ($\mu\text{g}/\text{spot}$)	RSD (%)	$\frac{\text{SD}}{\sqrt{n}}$ ($\mu\text{g}/\text{spot}$)	$\bar{m} \pm \frac{\text{SD}}{\sqrt{n}} \times t$ ($\mu\text{g}/\text{spot}$)	Recovery (%)
5.00	B ₁	4.83 \pm 0.15	3.1	0.067	4.83 \pm 0.19	96.7
	B ₆	4.96 \pm 0.13	2.6	0.058	4.96 \pm 0.16	99.2
	B ₁₂	4.88 \pm 0.17	3.5	0.076	4.88 \pm 0.21	97.6
10.00	B ₁	10.14 \pm 0.30	3.0	0.134	10.14 \pm 0.37	101.4
	B ₆	9.93 \pm 0.26	2.6	0.116	9.93 \pm 0.32	99.3
	B ₁₂	9.94 \pm 0.34	3.4	0.152	9.94 \pm 0.42	99.4
15.00	B ₁	15.20 \pm 0.44	2.8	0.197	15.86 \pm 0.55	101.3
	B ₆	15.02 \pm 0.36	2.3	0.161	15.02 \pm 0.45	100.1
	B ₁₂	14.80 \pm 0.49	3.3	0.218	14.80 \pm 0.61	98.7
20.00	B ₁	19.86 \pm 0.50	2.5	0.224	19.86 \pm 0.62	99.3
	B ₆	20.10 \pm 0.40	2.0	0.180	20.10 \pm 0.50	100.5
	B ₁₂	20.07 \pm 0.62	3.1	0.278	20.07 \pm 0.77	100.3
25.00	B ₁	24.87 \pm 0.57	2.3	0.255	24.87 \pm 0.71	99.5
	B ₆	25.23 \pm 0.48	1.9	0.215	25.23 \pm 0.60	100.9
	B ₁₂	24.92 \pm 0.75	3.0	0.334	24.92 \pm 0.93	99.7
30.00	B ₁	29.89 \pm 0.66	2.2	0.294	29.89 \pm 0.82	99.6
	B ₆	30.17 \pm 0.58	1.9	0.259	30.17 \pm 0.72	100.6
	B ₁₂	29.93 \pm 0.90	3.0	0.402	29.93 \pm 1.12	99.8
35.00	B ₁	34.49 \pm 0.86	2.5	0.386	34.49 \pm 1.07	98.5
	B ₆	34.52 \pm 0.72	2.1	0.322	34.52 \pm 0.89	98.6
	B ₁₂	34.46 \pm 1.14	3.3	0.510	34.46 \pm 1.41	98.5
40.00	B ₁	38.62 \pm 1.27	3.2	0.570	38.62 \pm 1.58	96.6
	B ₆	39.01 \pm 0.94	2.4	0.421	39.01 \pm 1.17	97.5
	B ₁₂	38.24 \pm 1.83	4.8	0.821	38.24 \pm 2.28	95.6

*n = 5, t = 2.776.

TABEL-3
 DETERMINATION OF VITAMINS B₁, B₆ AND B₁₂ IN PHARMACEUTICAL FORMULATIONS BY TLC-DENSITOMETRIC METHOD USING ACID TREATED ALEPPO BENTONITE AT $\lambda = 275$ nm FOR B₁ AND B₆ AND $\lambda = 525$ nm FOR B₁₂
 (MOBILE PHASE: ISO-BUTYL ALCOHOL:METHANOL:CHLOROFORM:
 ACETIC ACID:AMMONIA 7.5:1:3.5:0.4:0.5 v/v/v/v/v)

Product	Compound and dose	Found $\bar{m} \pm \text{SD}$ ($\mu\text{g}/\text{spot}$)	RSD (%)	$\frac{\text{SD}}{\sqrt{n}}$ ($\mu\text{g}/\text{spot}$)	$\bar{m} \pm \frac{\text{SD}}{\sqrt{n}} \times t$ ($\mu\text{g}/\text{spot}$)	Recovery (%)
Vitaneurin Fort (Tablets)	B ₁ 100 mg/tab.	113.0 \pm 3.630	3.2	1.620	113.0 \pm 4.500	113.0
	B ₆ 100 mg/tab.	114.1 \pm 3.420	3.0	1.530	114.1 \pm 4.250	114.1
	B ₁₂ 1 mg/tab.	1.197 \pm 0.051	4.3	0.023	1.197 \pm 0.064	119.7
Vitaneurin Fort (Ampoules)	B ₁ 100 mg/amp.	115.5 \pm 3.580	3.1	1.600	115.5 \pm 4.440	115.5
	B ₆ 100 mg/amp.	116.2 \pm 3.250	2.8	1.450	116.2 \pm 4.020	116.2
	B ₁₂ 1 mg/amp.	1.198 \pm 0.050	4.2	0.022	1.198 \pm 0.061	119.8

*n = 5, t = 2.776.

respectively. Calibration curves were obtained in the range of 5.0- 40.0 µg/spot for vitamins B₁, B₆ and B₁₂ (at $\lambda = 275$ nm) for standard solutions, while for pharmaceutical formulations, vitamins B₁, B₆ were determined at $\lambda = 275$ nm and vitamin B₁₂ was determined at $\lambda = 525$ nm in the range of 0.5-4.0 µg/spot.

REFERENCES

1. M. Lahmek, M.Sc. Thesis in Chemistry, Aleppo University, Syria (1987).
2. A. Alhaj Sakur, M.Sc. Thesis in Chemistry, Aleppo University, Syria (1995).
3. A.A. Ramadan, S. Antakli and I. Mahmoud, *Res. J. Aleppo Univ.*, **55**, 297 (2007).
4. M. Martini, M.Sc. Thesis in Chemistry, Aleppo University, Syria (1990).
5. C. Abdelouahab, H. Ait-Amar, T.Z. Obretenov and A. Gaid, *Analysis*, **16**, 292 (1988).
6. M.N. Shaikh Esmail Zadah, Ph.D. Thesis in Chemistry, Aleppo University, Syria (2005).
7. T. Sahlabji, M.Sc. Thesis in Chemistry, Aleppo University, Syria (2003).
8. A.A. Ramadan, S. Antakli and I. Mahmoud, *Res. J. Aleppo Univ.*, **56**, 97 (2007).
9. A.A. Ramadan, H. Al-Akra and O. Abdul Ghafour, *Res. J. Aleppo Univ.*, **27**, 135 (1998).
10. R.T. Sane, V.J. Doshi and S.K. Joshi, *J. Assoc. Off. Anal. Chem.*, **66**, 158 (1983).
11. G. Yanling, G. Fei, G. Sumangala, C. Andrew, S. Qinghai and G. Mingruo, *J. Food Chem.*, **110**, 769 (2008).
12. K. Akatsuka and I. Atsuya, *Fresenius J. Anal. Chem.*, **335**, 200 (1989).
13. B. Morelli, *Fresenius J. Anal. Chem.*, **354**, 97 (1996).
14. D. Lambert, C. Adjalla, F. Felden, S. Benhayoun, J.P. Nicolas and J.L. Gueant, *J. Chromatogr.*, **608**, 311 (1992).
15. M. Kimara and Y. Itokawa, *J. Chromatogr.*, **332**, 181 (1985).
16. I.N. Papadoyiannis, G.K. Tsioni and V.F. Samanidou, *J. Liq. Chromatogr.*, **20**, 3203 (1997).
17. J. Dalbacke and J. Dahlquist, *J. Chromatogr.*, **541**, 383 (1991).
18. M. Amin and J. Reusch, *J. Chromatogr.*, **390**, 448 (1987).
19. B. Urgessa, M.Sc. Thesis in Pharmaceuticals Analysis and Quality Assurance, Addis Ababa University, Ethiopia (2008).
20. British Pharmacopoeia, London: Stationery Office (2005); British Pharmacopoeia, London: Stationery Office (2007).