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Assay of Lercanidipine Hydrochloride with Azocaramine-G, Fe(III)/K₃[Fe(CN)₆] and Folin Ciocalteu Reagent

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Three simple and sensitive visible spectrophotometic methods (A-C) have been developed for the assay of lercanidipine hydrochloride in bulk and formulations. Method **A** is based on the formation of ion-association complex with azocaramine-G (ACG) which was extracted into chloroform ($\lambda_{max} = 540$ nm). Method **B** is based on the formation of coloured complex with K₃[Fe(CN)₆] in presence of Fe(III) ($\lambda_{max} = 730$ nm). Method **C** is based on the oxidation of drug with Folin-Ciocalteu reagent ($\lambda_{max} = 750$ nm). The results of analysis have been validated statistically. Recoveries range from 99.4-100.5 % for the routine assay of lercanidipine hydrochloride formulations.

Key Words: Lercanidipinehydrochloride, Azocaramine-G, Fe(III)/K₃[Fe(CN)₆], Folin Ciocalteu reagent.

INTRODUCTION

Lercanidipine hydrochloride (LER) is chemically 2-{(3,3diphenylpropyl) methylamine}-1,1-dimethylethyl-methyl-1,4dihydro-2,6-dimethyl-4-(3-nitrophenyl)-3,5-pyridine carboxylic ester hydrochloride (Fig. 1). This drug is used as a calcium channel blocker in the treatment of hypertension¹. The drug is cited in Merck Index² and Martindale³. In literature, a number of analytical methods have been described for estimation of LER. These include HPLC^{4,5}, electrophoresis⁶, LC-MS^{7,8-11}, extractive spectrophotometric¹² and few visible spectrophotometric¹³⁻²² methods. With an aim to develop relatively cheap, sensitive and useful for the laboratories with modest infra structure, the authors developed a simple, sensitive, accurate, reproducible, reliable and economical analytical method for estimation of LER in bulk drug and formulations.

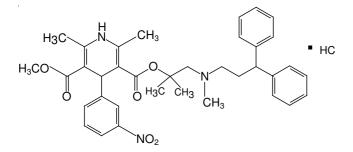


Fig. 1. Chemical structure of lercanidipine

EXPERIMENTAL

All spectral and absorbance measurements were made on a Elico SL-177 model visible spectrophotometer with 1 cm matched glass cells and on Unicam UV 500 spectrophotometer made by Thermo Electron Corporation. All pH measurements were made on a Elico LI 120 digital pH meter.

All the reagents were of analytical grade and all solutions were prepared in double distilled water. Aqueous solutions of ACG (Gurr; 0.05 %, 5.50 × 10⁻⁴ M), Fe(III) (Wilson; 0.054 %, 3.32 × 10⁻³ M), K₃Fe(CN)₆ (BDH; 0.1 %, 3.03 × 10⁻³ M), Na₂CO₃ (BDH; 10 %, 9.43 × 10⁻¹ M), were prepared by dissolving requisite quantities in double distilled water. Folin-Ciocalteu (FC) (Loba; 2N) solution was used as it is. Buffer solution (pH 1.5) was prepared by mixing appropriate concentrations of glycine and HCl (Qualigen, AR Grade).

Standard drug solution: The stock solution (mg/mL) was prepared by dissolving 100 mg of drug in 100 mL of chloroform. Aliquot portion of (25.0 mL) the above stock solution was diluted stepwise with same solvent to obtain working standard solution of 40 μ g/mL (method **A**). About 100 mg of bulk drug was dissolved in 10.0 mL methanol and reduced using standard literature method²³. The reduced drug solution was in methanol was evaporated to dryness. The residue was dissolved and diluted stepwise with distilled water to obtain working standard solutions of concentrations 100 μ g/mL (method **B** and **C**).

Recommended procedures

Method A: Into a series of 125.0 mL separating funnels containing aliquots of standard drug solution [LER: 1.0-3.0 mL, $40 \mu g/mL$], 6.0 mL of buffer solution (pH 1.5) and 2.0 mL of 5.50 × 10^4 M dye solution were added. The total volume in separating funnel was adjusted to 15.0 mL with distilled water. To each separating funnel 10.0 mL of chloroform was added and the contents were shaken for 2 min. The two phases were allowed to separate and the absorbance of the separated chloroform layer was measured at 540 nm against blank. The coloured species were stable for 1 h. The amount of drug present (LER) was deduced from the calibration curve.

Method B: In to a series of calibrated tubes, aliquots of standard drug solution (LER: 1.0-3.0 mL, 100 µg/mL) was transferred and 1.0 mL of 3.32×10^{-3} M FeCl₃ solution was added. The tubes were stoppered immediately and shaken well for 5 min. Then 0.5 mL of 3.03×10^{-3} M potassium ferricyanide solution was added into each tube and was closed with lids immediately. After 5 min, 1.0 mL of 1 N HCl was added and the final volume was made upto 20.0 mL with distilled water. The absorbance of the solution in each tube was measured immediately at 730 nm for (LER). The stability of coloured species was found to be 1 h. The amount of the drug (LER) was calculated from its calibration graph.

Method C: Aliquots of drug solutions (LER: 0.5-2.5 mL, 100 μ g mL⁻¹) were delivered into a series of calibrated tubes. To each tube, 1.5 mL of Folin-Ciocalteu reagent (2.0 N) was added. After 3 min, 5.0 mL of 9.43 × 10⁻¹ M Na₂CO₃ (10 %) solution was added and kept aside for 15 min (LER) and then diluted up to the mark (20.0 mL) with distilled water. The absorbance of the blue coloured solution was measured at 750 nm (LER) against corresponding reagent blank. The stability of coloured species was found to be 45 min. The amount of drug was computed from its appropriate calibration graph.

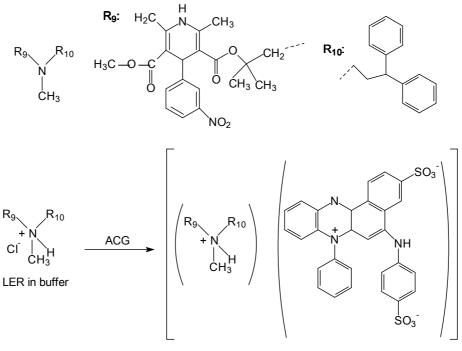
Pharmaceutical formulations: Since only two formulations (Larez and Lerka) are available for LER (tablets), these formulations of different batches were collected and analyzed as 4 sets to verify the validity of proposed methods. Accurately weighed quantity of tablet powder equivalent to 100 mg of LER was extracted with warm chloroform $(3 \times 25.0 \text{ mL} \text{ portions})$ and filtered. The volume of combined extract was evaporated to dryness, reduced as described in the preparation of standard drug solution. Working standard solutions of concentrations 40 µg/mL (method **A**) and 100 µg/mL (method **B** and **C**) were prepared to test the validity of methods developed. Further, the UV spectrophotometric method which was suggested for the identification of LER has been moulded for its assay and chosen as the reference method for ascertaining the accuracy of the proposed methods.

RESULTS AND DISCUSSION

The optimum conditions for each method were established by varying one parameter at a time²⁴ and keeping the others fixed and observing the effect produced on the absorbance of the coloured species.

Optimum conditions and chemistry of coloured species

Method A: 6.0 mL buffer (pH = 1.5), 2.0 mL of azocaramine-G solution (5.50×10^{-4} M), chloroform as solvent for extraction of coloured species (3:2 ratio of aqueous to organic phase) were identified as optimum conditions. The absorbance of coloured species was measured at 540 nm after 5 min. The drug (LER) possess tertiary amino group in the side chain and this group was involved in ion-association complex formation with azocaramine-G which is extractable into chloroform. The colour formation by azocaramine-G reagent with LER may be explained in the following manner based on the analogy reports²⁵ (**Scheme-I**). The quantitative measure of the effect of complexation on acid-base equilibrium is most likely to be



Scheme-I

interpretable in terms of electronic, steric and other effect of complexing. The protonated nitrogen (positive charge) of the drug (LER) under appropriate experimental conditions is expected to attract the oppositively charged part of dye (negative charge) and behaves as a single unit being held together by electrostatic attraction. This was further confirmed by the mole-ratio of drug to dye (1:1) obtained by slope-ratio studies.

Method B: The optimum conditions in this method were fixed as: 1.0 mL of Fe(III) solution $(3.32 \times 10^{-3} \text{ M})$, 0.5 mL of potassium ferricyanide $(3.03 \times 10^{-3} \text{ M})$, 1.0 mL of 1 N HCl. The order of addition of reagents is: drug, Fe(III), potassium ferricyanide and 1 N HCl. The role of HCl is for the maintenance of acidity prior to dilution. The absorbance of coloured species was measured at 730 nm after 10 min. In this method, the formation of the coloured complex obtained is due to formation of ferrous ferricyanide complex. Fe(II) produced by the reduction of Fe(III) in the oxidation of drug (LER). This Fe(II) subsequently reacts with potassium ferricyanide to give coloured ferrous ferricyanide complex (**Scheme-II**).

Step 1

 $\begin{array}{c} Drug + Fe(III) & \longrightarrow \\ Oxidation \ products \ of \ drug + Fe(II) + \ Fe(III) \\ (LER) & (Unreacted) \end{array}$

Step 2

 $3Fe^{2+} + 2Fe(CN)_6^{3-} \longrightarrow Fe_3[Fe(CN)_6]_2$ Scheme-II

Method C: The method involves the reduction of Folin-Ciocalteu reagent by the drug in the alkaline medium. The optimum conditions found in this method were 1.5 mL of Folin-Ciocalteu reagent (2 N), 5.0 mL of 10 % Na₂CO₃ (9.43 × 10^{-1} M) and order of addition of reagents are: drug, Folin-Ciocalteu reagent and Na₂CO₃. The absorbance of coloured species was measured at 750 nm after 15 min. The colour formation by Folin-Ciocalteu reagent with LER may be explained in the following manner based on the analogy with the reports of the earlier workers²⁶. The mixed acids in the Folin-Ciocalteu preparation are the final chromogen and involve the following chemical species

 $3H_2O\cdot P_2O_5\cdot 13WO_3\cdot 5MoO_3\cdot 10H_2O$ and $3H_2O\cdot P_2O_5\cdot 14WO_3\cdot 4MoO_3\cdot 10H_2O$

Lercanidipine hydrochloride probably effects a reduction of 1, 2 or 3 oxygen atoms from tungstate and/or molybdate in Folin-Ciocalteu reagent, thereby producing one or more of the possible reduced species which have a characteristic intense blue colour.

Analytical data: The optical characteristics such as Beer's law limits, molar absorptivity, Sandell's sensitivity and regression characteristics, the relative standard deviation and percentange of error at 95 % confidence level are given in Table-1. The accuracy of the method for the drug was ascertained by comparing the result by proposed methods and reference method (UV), statistically²⁴ by t- and F- tests (Table-2). An additional check of accuracy of the proposed method, recovery experiments were performed by adding a fixed amount of the drug to the pre analyzed formulation and results are given in Table-2. This comparison show that there is no significant difference between the results obtained by the proposed methods and reference method.

Interference studies: The ingredients usually present in the preparation of formulations such as, yellow oxide of iron and titanium dioxide anticipated in pharmaceutical formulations did not interfere with the assay of LER by proposed methods. Commercial formulations such as Lerez and Lerka (tablets) containing LER were successfully analyzed by the proposed methods. The values obtained by the proposed and reference methods for formulations were compared statistically with t- and F-tests and found not to differ significantly. The results are summarized in Table-2.

Conclusion

Unlike in GLC and HPLC procedures, spectrophotometer involved in present procedure is cheap and simple to operate. The high sensitivity, selectivity and simplicity of the methods

	TABLE-1								
OPTICAL CHARACTERISTICS, PRECISION, ACCURACY OF THE PROPOSED METHODS									
	Azocaramine-G Fe(III)/K ₃ [Fe(CN		N ₆] Folin-Ciocalteu reagent						
Optical characteristics -	Method A	Method B	Method C						
λ_{max} (nm)	540	730	750						
Beer's law limits (µg/mL)	4-12	5-15	2.5-12.5						
Detection limit (µg/mL)	7.55-10-2	$1.05 - 10^{-1}$	5.14-10-2						
Molar absorptivity (1 mol ⁻¹ cm ⁻¹)	$3.27-10^4$	$2.94-10^4$	3.13-10 ⁴						
Sandell's sensitivity (µg/cm ² /0.001 absorbance unit)	1.98-10-2	2.2-10-2	$2.07 - 10^{-2}$						
Regression equation $(y = a + bC)^*$									
Slope (b)	5.06-10-2	4.58-10-2	4.88-10-2						
Standard deviation on slope (S_b)	1.5-10-4	1.5-10-4	$1.01 - 10^{-4}$						
Intercept (a)	-2.0-10-4	-1.2-10-3	-7.0-10 ⁻⁴						
Standard deviation on intercept (S_a)	$1.27 - 10^{-3}$	$1.606 - 10^{-3}$	8.35-10-4						
Standard error of estimation (S_e)	9.49-10-4	$1.197 - 10^{-3}$	7.96-10-4						
Correlation coefficient (r)	0.9992	0.9994	0.9997						
Optimum photometric range (µg/mL)	4.2-11.5	4.8-14.4	5.2-12.2						
Relative standard deviation**	0.46	0.62	0.65						
Percentage range of error (confidence limit)									
0.05 level	0.48	0.66	0.68						
0.01 level	0.76	1.03	1.07						
Percentage error in bulk sample***	-0.24	0.21	-0.27						

y = a + bC where C = concentration of analyte in $\mu g/mL$ and y = absorbance unit. **Calculated from six determinations. ***Average error of three determinations.

TABLE-2										
ASSAY OF LERCANIDIPINE HYDROCHLORIDE PHARMACEUTICAL FORMULATIONS										
Formulations* am	Labelled	Amount found by proposed method (mg)**		Deferment	Recovery by proposed methods*** (%)					
	amount (mg)	M-A (ACG)	M-B (Fe(III)/ K ₃ (Fe(CN) ₆)	M-C (F-C)	Reference - method	M-A (ACG)	M-B (Fe(III)/K ₃ (Fe(CN) ₆)	M-C (F-C)		
Tablet I	10	10.02 ± 0.08 F = 1.39 t = 0.67	9.98 ± 0.07 F = 2.26 t = 0.32	10.13 ± 0.11 F = 1.12 t = 1.49	9.96 ± 0.1	100.3 ± 0.8	99.76 ± 0.66	101.3 ± 1.1		
II	10	10.03 ± 0.11 F = 1.27 t = 0.55	9.90 ± 0.07 F = 2.60 t = 1.53	9.88 ± 0.14 F = 1.39 t = 1.27	10.07 ± 0.12	100.3 ± 1.1	99.02 ± 0.74	98.83 ± 1.4		
Ш	10	9.91 ± 0.09 F = 1.76 t = 0.26	9.98 ± 0.13 F = 1.25 t = 1.18	9.87 ± 0.10 F = 1.42 t = 2.15	9.92 ± 0.12	99.07 ± 0.9	99.78 ± 1.34	98.65 ± 1.0		
IV	10	10.10 ± 0.16 F = 1.89 t = 1.31	10.00 ± 0.11 F = 1.04 t = 0.61	9.98 ± 0.09 F = 1.60 t = 0.46	9.92 ± 0.12	101.0 ± 1.6	100.0 ± 1.1	99.78 ± 0.9		

*Four different samples of tablets. **Average \pm standard deviation of six determinations; the t and F-test values refer to comparison of the proposed method with the reference method. Theoretical values of 95 % confidence limit, F = 5.05, t = 2.57. ***: After adding 3 different amounts of the pure labeled to the pharmaceutical formulations, each value is an average of 3 determinations.

proposed make them ideally suitable for the assay of LER in pharmaceutical formulations. The proposed methods exploit the various functional groups in LER. The ingredients usually present in pharmaceutical formulations do not interfere in the colour development by proposed methods. All the proposed methods are simple, economical and do not require much of instrumentation.

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REFERENCES

- 1. K.J. McClellan and B. Jarvis, Drugs, 60, 1123 (2000).
- S. Budavari, Merck Index, Merck & Co. Inc., New York, edn. 12, p. 5469 (1996).
- J.E.F. Reynolds, The Complete Drug Reference, Martindale Extra Pharmacopoeia, The Pharmaceutical Press, London, edn. 32, p. 893 (1999).
- A. Alvarez-Lueje, S. Pujol, J.A. Squella and L.J. Nunez-Vergara, J. Pharm. Biomed. Anal., 31, 1 (2003).
- M. Barchielli, E. Dolfini, P. Farina, B. Leoni, G. Targa, V. Vinaccia and A. Tajana, J. Cardiovasc. Pharmacol., 29, S1 (1997).
- T. Christians, D. Diewald, C. Wessler, Y. Otte, J. Lehmann and U. Holzgrabe, J. Chromatogr. A, 853, 455 (1999).
- J. Fiori, R. Gotti, C. Bertucci and V. Cavrini, *J. Pharm. Biomed. Anal.*, 41, 176 (2006).
- V.A. Jabor, E.B. Coelho, D.R. Ifa, P.S. Bonato, N.A. Dos-Santos and V.L. Lanchote, J. Chromatogr. B; Anal. Technol. Biomed. Life Sci., 796, 429 (2003).

- 9. I.I. Salem, J. Idress, J.I. Al-Tamimi and P. Farina, J. Chromatogr. B; Analy. Technol. Biomed. Life Sci., 803, 201 (2004).
- A.B. Baranda, C.A. Mueller, R.M. Alonso, R.M. Jimenez and W. Weinmann, *Ther. Drug Monit.*, 27, 44 (2005).
- M. Kalovidouris, S. Michalea, N. Robola, M. Koutosopoulou and I. Panderi, *Rapid Commun. Mass Spectrom.*, 20, 2939 (2006).
- 12. N. Erk, Pharmazie, 58, 801 (2003).
- 13. B.S. Sundar, M.E. Gupta and G.D. Rao, *Acta Cien. Indica*, **31C**, 25 (2005).
- 14. M.E. Gupta, B.S. Sundar and G.D. Rao, *Acta Cien. Indica*, **31C**, 27 (2005).
- 15. M.E. Gupta, B.S. Sundar and G.D. Rao, *Acta Cien. Indica*, **31C**, 51 (2005).
- N.R. Rao, P. Nagaraju, C. Srinivasulu, P. Radhakrishnamurthy, D. Sireesha and S.U. Bhaskar, *Asian J. Chem.*, 16, 1950 (2004).
- 17. M.E. Gupta, B.S. Sundar and G.D. Rao, *Indian Pharmacist*, **29**, 77 (2004).
- V.S. Saradhi, V. Himabindu and G.D. Rao, *Acta Cien. Indica*, **31C**, 305 (2005).
- 19. V.S. Saradhi, V. Himabindu and G.D. Rao, Asian J. Chem., 18, 718 (2006).
- 20. V.S. Saradhi, V. Himabindu, G.D. Rao, Asian J. Chem., 18, 1551 (2006).
- C.K.V. Aradhya, B.K. Koteswar Rao, N.Y. Manohara, S.A. Raju S., Int. J. Chem. Sci., 4, 135 (2006).
- S. Mihaljica, D. Radulovic and J. Trbojevic, *Chromatographia*, 61, 25 (2005).
- B.S. Furniss, A.J. Hannaford, P.W.G. Smith and A.R. Tatchell, Text Book of Practical Organic Chemistry, (Reprinted), Longman, England edn. 5, p. 893 (1994)
- D.L. Massart, B.G.M. Vandeginste, S.N. Doming, Y. Michotte and L. Kaufman, Chemometrics: A Text Book, Elsevier, Amsterdam (1988).
- 25. M.D. Santos, C.M. Isabel, and C.L. Goncalves, *Bol. Fac. Farm. Coimbra*, **1**, 37 (1976).
- 26. L.G. Peterson, Anal. Biochem., 100, 20 (1979).